

## PHYSICOCHEMICAL AND SENSORY CHARACTERISTICS OF KASHK AS INFLUENCED BY QUINOA FLOUR ADDITION

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### ABSTRACT

The current work was aimed to study the application of quinoa flour in the production of kashk, a fermented dairy product and the resulting physicochemical and sensory properties. Three kashk samples were formulated with quinoa flour (1,2 and 3%). Pretreatment of the kashk samples with quinoa flour had significantly increased the viscosity (from 4040 to 4127 cP), acidity value (increased 0.3) and reduced the syneresis of the samples ( $P < 0.05$ ). Concerning color, no significant changes were observed in the “b” and “L” values in all the samples which remained constant following 60 days of storage whereas an increase in the “a” value was observed upon supplementation of quinoa flour (from  $-3.00 \pm 0.50$  to  $-0.11 \pm 0.51$ ). Sensory analysis revealed that increasing the quinoa concentration increased consistency but decreased flavor. Scanning electron micrographs revealed that the denser microstructure was seen in kashk containing quinoa flour. This could suggest that quinoa flour could be used to improve the microstructure of the kashk. Moreover, adding quinoa flour to kashk could compensate lysine depleted in the kashk thermal process. These results indicated that kashk supplementation with quinoa flour could be suggested as an effective means to improve the functional properties of kashk.

**Keywords:** Quinoa, Viscosity, Kashk, Lysin depletion

### INTRODUCTION

Different beneficial effects such as enhanced digestion, tolerance to lactose, antimicrobial properties, hypocholesterolemic, control of blood glucose, antihypertensive consequence, anti-inflammatory, antioxidant action, anti-carcinogenic and anti-allergenic activities have been currently suggested the fermented dairy product (Kukhtyn *et al.*, 2018).

Kashk is a very old popular fermented Iranian dairy product that is traditionally prepared in dried form and made industrially in a liquid form. The Kashk is a common fermented dairy product in many geographical areas of Iran especially in rural communities (Golestan *et al.*, 2016). However, similar products have been named in other countries as Tarhana (Turkey and Greece), Kishk (Lebanon and Egypt), Kushuk (Iraq), Madeer-Oggt (Saudi Arabia), Kichk (India), Talkuna (Finland), Tahanya (Hungary) and Atole (Scotland) (Mashak, Sodagari, Mashak, & Niknafs, 2014). Kashk is a product riched in protein and minerals thus, it is considered as one of the important sources of high-quality dairy products recommended using as a dietary supplemented product in the diets of children, pregnant and lactating women (Soltani & Güzeler, 2013). In order to produce the industrial liquid kashk, concentrated yogurt as a byproduct of the industrial dairy companies is mainly used. Few characteristics such as low pH (less than 4.5), 20–25% nonfat solid, 1% fat, 3% salt, and at least 13% protein in the liquid Kashk are currently approved by the Institute of Standard and Industrial Research of Iran (Shiroodi *et al.*, 2012). There are only a few studies on the enhancement of kashk texture. For example, rice flour and tragacanth gum were used in kashk as texturizers (Mashak *et al.*, 2014; Shiroodi *et al.*, 2012). Additionally, the long-term heat treatment of kashk may cause nutritional value reduction specially lysin. Therefore, it is necessary to add a substance for improving texture and nutritional values.

Quinoa (*Chenopodium quinoa* Willd) is a pseudocereal plant, belongs to the family of Chenopodiaceae and is widely suggested as a beneficial grain. Few unique characteristics such as enriched essential amino acids, lack of gluten, presence of most essential amino acids, lack of gluten and high content and high content of microelements such as Ca, Mg, Fe, flavonoids as important health-promoting compounds, linoleic (52%) and oleic (25%) unsaturated fatty acids make the quinoa seed interesting for consumers. The antioxidant/ antiradical activities of

quinoa seed are associated with the presence of tocopherols and carotenoids (Ruales & Nair, 1993; Ruiz *et al.*, 2014; Zhang *et al.*, 2014). Moreover, many validated functional properties of quinoa such as high water holding capacity, gelation and emulsification have allowed its increasing demands (Graf *et al.*, 2015; Valcárcel-Yamani & Lannes, 2012).

One of the main purposes of adding these ingredients to fermented dairy products is to improve their nutritional values in terms of mineral, vitamins and protein contents (Zare *et al.*, 2011). However, in the food industry, quinoa flour has been used as thickeners and fat replacers in yogurt (Codinã *et al.*, 2016; Curti *et al.*, 2017; Lorusso *et al.*, 2018; Zannini *et al.*, 2018) cream cheese (Lemes *et al.*, 2016) beverages (El-Deeb *et al.*, 2014; Ludena Urquiza *et al.*, 2017) and ice-cream (Jacobsen *et al.*, 2003), although there is no investigations have been conducted on kashk. Consequently, we have supplemented kashk with different concentrations of quinoa flour to see whether any overall acceptability of the product is improved or not.

The objective of our study was kashk supplementation with different concentrations of quinoa flour to see whether any overall acceptability of the product is improved or not.

### MATERIALS AND METHODS

#### Chemicals

The chemicals, solvents and standards include hydrochloric acid, sodium acetate, OPA, β- mercaptoethanol, sodium hydroxide, sodium acetate, methanol, borate buffer, and the other standard reagents were provided by Merck (Darmstadt, Germany). Norovaline was obtained from Sigma Chemicals Co. (St. Louis, USA).

#### Quinoa flour, milk and kashk proximate value

Different factors including moisture, energetic values and macronutrient contents such as fat, ash, proteins and carbohydrates were analyzed in the samples as previously described by AOAC (AOAC, 2016). The following equations were used to evaluate total carbohydrates and energetic values according to (Cardoso *et al.*, 2019).

Total carbohydrates ( $\text{g} \cdot 100 \text{ g}^{-1}$ ) =  $100 - (\text{m fat} + \text{m ash} + \text{m proteins})$  (I);  
 Energy ( $\text{kcal} \cdot 100 \text{ g}^{-1}$ ) =  $4 \times (\text{m proteins} + \text{m carbohydrates}) + [9 \times (\text{m fat})]$  (Ludena Urquiza *et al.*, 2017).

Few nutrient components including fat, protein, pH, solid nonfat materials, density and freezing point in the fresh milk were analyzed using Cryostar I Funke Gerber 2001 (Berlin, Germany) and Lactostar Funke Gerber 2001 (Berlin, Germany). The standard test methods for milk and dairy products (Iranian National Standards, 2852 and 10154) were used to report the acidity and microbial counting and Copan kit (Christian Hansen, Hoersholm, Denmark) was employed to determine the antibiotic residues. For kashk analysis, fat, protein, pH and SNF were measured based on the Iranian Standards Institute (NO. 2452). The enumeration of *mold*, *yeast*, *Escherichia coli*, *coliform*, *Staphylococcus aureus* are currently compulsory in the Kashk samples.

### Kashk preparation

In order to prepare the kashk, the standardized milk fat (2.5% w/v) was initially homogenized at  $100\text{--}200 \text{ kg} \cdot \text{cm}^{-2}$ , pasteurized (at  $90^\circ \text{C}$  for 15 min) and cooled to  $43^\circ \text{C}$ .

Milk was then supplemented with the lyophilized starter culture, CH1 (50U) DVS, followed by incubation at  $43^\circ \text{C}$  to set the acidity at  $150^\circ$  Dornic. The product was then stored at  $4\text{--}5^\circ \text{C}$ , overnight. The preparation was then heated to  $80^\circ \text{C}$  for 4 h until its color changed to light brown. The sample was stirred constantly to prevent burning on the interface of the tank. The infrared moisture analyzer MA100 was employed to adjust the total solid to 20% w-w<sup>-1</sup> (SNF 14.28% w-w<sup>-1</sup>) (Sartorius, Germany) (Zhang *et al.*, 2014). The mixture was finally divided into four equal segments. The supplemented kashk sample consisted of quinoa flour (1, 2, 3) % were added to kashk samples 5 min before homogenization at  $75^\circ \text{C}$  at  $100\text{--}150 \text{ kg} \cdot \text{cm}^{-2}$ . Control (without quinoa flour) and supplemented product were packed in a hot filling manner at  $80^\circ \text{C}$  in plastic containers and stored at  $4^\circ \text{C}$ . Physicochemical and textural, sensorial properties and apparent viscosity were assessed through the 15 days of storage intervals for 60 days.

### pH, total titratable acidity and acid value

The pH of yogurt samples was measured at 1, 15, 30, 45 and 60 days of storage at  $4^\circ \text{C}$  using a pH meter (Greisinger electronic, Regenstauf, Germany) with a glass electrode over the range 6.8 to 4.0. The titration of samples with  $1/9 \text{ mole} \cdot \text{L}^{-1}$  NaOH and phenolphthalein was then used to record the dornic acidity. Acid value was principally defined as the required amounts of potassium hydroxide to neutralize the free fatty acid in each gram of fat. The acid value was determined based on the AOCs: Cd 3a-63 method (Lemes *et al.*, 2016).

### Lysin assessment

Lysin was measured in quinoa flour, yogurt, control kashk and the sample containing 3% quinoa. In order to derivatize the lysin, the following reagents were used:  $0.01 \text{ mol} \cdot \text{L}^{-1}$  sodium acetate in water (mobile phase A) and methanol (mobile phase B). The borate buffer ( $0.5 \text{ mol} \cdot \text{L}^{-1}$ ; pH: 10.2); for preparation of [2ME; OPA: 2-Mercaptoethanol; O-Phthalaldehyde] reagent (pH: 9.3): 0.05 gr of the OPA was dissolved in 500  $\mu\text{L}$  borate buffer (pH: 9.9) with 4.5  $\mu\text{L}$  methanol and 25  $\mu\text{L}$  of 2-ME, and Norovaline reagent: 50  $\mu\text{L}$  Norovaline (concentration 1000  $\mu\text{M}$ ) was added to 950  $\mu\text{L}$  HCl  $0.01 \text{ mol} \cdot \text{L}^{-1}$ . The method was performed for lysin analysis including the hydrolysis of quinoa flour samples with  $6 \text{ mol} \cdot \text{L}^{-1}$  HCl and  $0.5 \text{ g} \cdot \text{L}^{-1}$  of  $\beta$ -mercaptoethanol in sealed vacuum-evacuated tubes at  $110^\circ \text{C}$  for 21 h. The outcome precipitate was removed following centrifugation at 6000 g for 30 min. The sample was neutralized using NaOH  $3.0 \text{ mole} \cdot \text{L}^{-1}$ . A consecutive sampling of 100  $\mu\text{L}$  of borate buffer and 25  $\mu\text{L}$  of the sample was then mixed twice for 0.5 min. Subsequently, 50  $\mu\text{L}$  of 2ME; OPA reagent was added. 100  $\mu\text{L}$  of HCl  $0.1 \text{ mole} \cdot \text{L}^{-1}$  was taken up and thoroughly mixed six times. After that, 50  $\mu\text{L}$  of this solution was added to 200  $\mu\text{L}$  mobile phase A, mixed twice and 20  $\mu\text{L}$  of the mixture was finally injected. The 10  $\mu\text{L}$  of injection volume used for samples. The separation was carried out using a 4 mm x 25 cm with precolumn (PS Spheribond 80-5 ODS 2), AK351, Vertex Plus column,  $40^\circ \text{C}$ , with a flow rate of  $1.0 \text{ ml} \cdot \text{min}^{-1}$  in HPLC system an autosampler system (Perkin Elmer, Australia) equipped with a fluorescence detector (RF-20A series) at  $\lambda$  excitation 348nm and  $\lambda$  emission 450 nm. The reporting of lysin content was made compared to the standard. The retention time for lysin was 4.3 min. The lysin content was expressed in  $\text{mg} \cdot (\text{kg} \cdot \text{d} \cdot \text{m})^{-1}$ .

### Syneresis calculation

The syneresis (the free whey) was defined by the standard centrifugation method in which, 20g of the sample was centrifuged (640 g, 20min,  $4^\circ \text{C}$ ), the clear pellet was then collected and weighed. The following equation was finally employed to calculate the syneresis (Eq. 1).

$$\text{Syneresis} = [\text{weight of supernatant (g)} / \text{Weight of sample (g)}] \times 100 \quad (\text{Eq. 1})$$

### Sensory assessment

The sensory evaluation including color, taste, texture and overall acceptance based on 5-point hedonic scales (1: dislike extremely; 5: like extremely), was performed by forty trained panelists (aged between 25 to 35 years old). Each sample was scored, individually. The samples were accessible to the panelists in the individual plastic containers. Samples were coded with three digits which were randomly tested by the panel group.

### Color

The digital imaging method was applied to evaluate the color using a Chroma meter CR-400, Konica Minolta, INC (Osaka, Japan). The Chroma meter was calibrated using the white tile. A total of five grams of each sample was placed in a special chamber, the standard color parameters were comprised of brightness ( $L^* = 23.92$ ), redness ( $a^* = -1.29$ ) and yellowness ( $b^* = 1.19$ ). The average color measurement for each sample was then reported at various sites.

### Apparent viscosity

The Brookfield cone and plate viscometer (DV2 Pro II, Brookfield Engineering Laboratories, USA) were used to report the rheological measurement of the samples. The viscometer was equipped with Spindle CP51 and a temperature controller. 0.5 ml of each sample was then transferred into the plate. To achieve a constant solution, the probe was rotated at  $50 \text{ s}^{-1}$  for 3 min. The device was tuned at  $25^\circ \text{C}$  for 10 min. The shear rate increased linearly from 1 to  $300 \text{ s}^{-1}$  for 3 min. To describe the flow behavior of the kashk mixture, five apparent viscosity equations were used. The best qualities which revealed the highest fitness were finally chosen. The viscosity of the samples was measured at  $27^\circ \text{C}$  upon 1, 15, 30, 45 and 60<sup>th</sup> days of storage. For each sample, apparent viscosity records were completed in duplicate. Flow behavior was described by the fitting of the Power law (Eq. 2), Bingham (Eq. 3) and Casson models (Eq. 4)

$$\tau = k \dot{\gamma}^n, \quad (\text{Eq. 2})$$

where  $\tau$  is the shear stress ( $\text{D} \cdot \text{cm}^{-2}$ ),  $\dot{\gamma}$  is the shear rate ( $\text{s}^{-1}$ ),  $k$  is the Power law consistency coefficient (cP), and  $n$  is the Power law flow behavior index (dimensionless) (Eq. 2).

$$\tau = \tau_{0B} + \eta_B \dot{\gamma}, \quad (\text{Eq. 3})$$

where  $\tau_{0B}$  is the Bingham yield stress ( $\text{D} \cdot \text{cm}^{-2}$ ) and  $\eta_B$  is the Bingham plastic viscosity (cP) (Eq. 3).

$$(\tau)^{0.5} = k_{0c} + k_c (\dot{\gamma})^{0.5} \quad (\text{Eq. 4})$$

where,  $k_{0c}$  and  $k_c$  are the intercept and slope of the plot of  $(\dot{\gamma})^{0.5}$  versus  $(\tau)^{0.5}$ , respectively. Therefore, the Casson yield stress ( $\text{D} \cdot \text{cm}^{-2}$ ) and Casson viscosity ( $\eta_c$ , cP) were calculated as the square of the intercept and slope, respectively [ $\tau_{0c} = (k_{0c})^2$ ,  $\eta_c = (k_c)^2$ ] (Eq. 4).

### Texture

The Texture Analyser (TA-XT2, Stable Micro System Ltd., Surrey, UK) was employed to analyze the texture of the samples. A two-bite compression test at pretest speed of  $1 \text{ mm} \cdot \text{s}^{-1}$ , test speed of  $1 \text{ mm} \cdot \text{s}^{-1}$ , the time interval of 10s and strain deformation of 25% was conducted for the Texture Profile Analysis test. Then, the rest of the sample was tested using a cylindrical probe with a diameter of 80 mm.

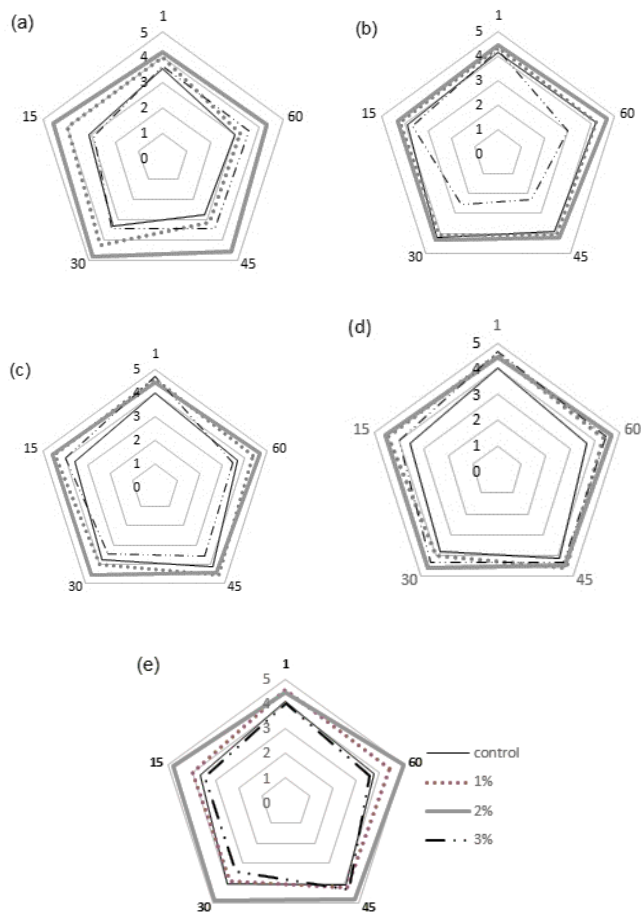
### Scanning electron microscopy

The freeze-dried samples were fixed on the aluminum holder and covered with gold in a sputter coater (Desk Sputter Coater DSR1, Nanostructural Coating Co., Iran) before an examination on the Scanning Electron Microscopy (SEM, VEGA3, TESCAN, Czech Republic). Samples were then observed under the accelerating voltage of 10.0 kV. The working distance of the microscope lens and sample surface was 7.03- 8.91 mm.

### Statistical analysis

Data were analysed by SPSS (ver. 21). For statistical analysis, one-way analysis of variance (ANOVA) was used to analyse the data. The Duncan post-test was used to find the significant difference between the means values ( $p < 0.05$ ).





**Figure 3** Sensory attributes of kashk samples supplemented with quinoa flour. Odor (a); Taste (b); colour (c); consistency (d) total acceptance (e).

### Color

**Table 2** Illustrates the color parameters in different kashk during the storage period. Concerning  $L^*$  and  $b^*$  parameters, no considerable change was observed in the following supplementation of the samples with quinoa flour. The quinoa flour did not alter the light scattering properties of the sample and thus no differences in the lightness of the samples were observed. When assessing the  $a^*$  parameter, kashk containing three percent quinoa flour presented a value closer to 0 demonstrating higher proximity to red, which is significantly different ( $p \leq 0.05$ ) from the control sample. A similar result was seen in yogurt supplemented with three percent of lentil flour which revealed a reddish appearance to some extent (Zare et al., 2011).

**Table 1** Free fatty acid, syneresis and color parameters of kashk samples during storage time.

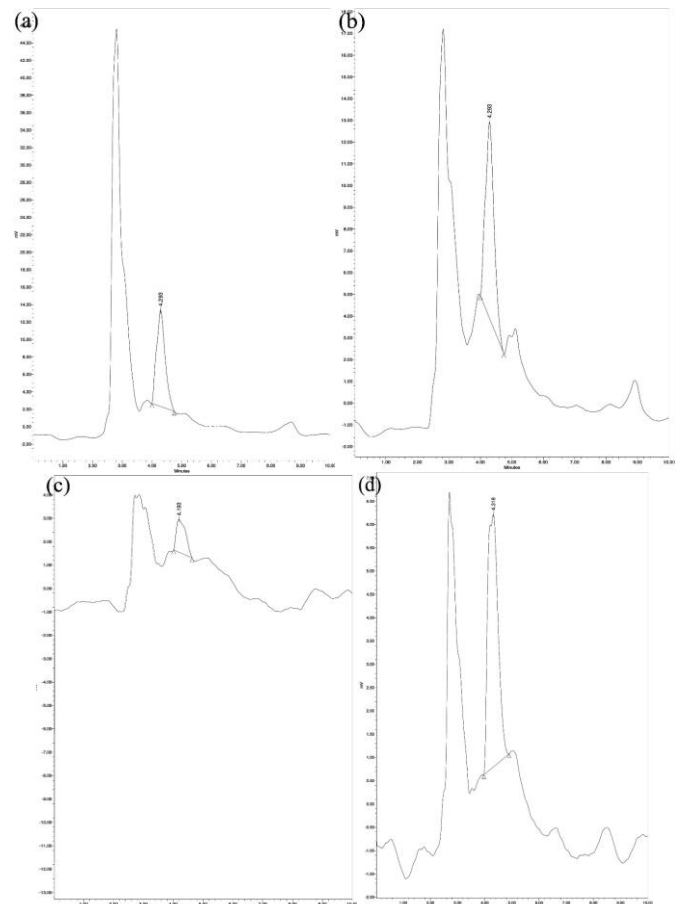
$T$	Day	Control	1%	2%	3%
ADV	1	0.60±0.01 <sup>D</sup> a	0.40±0.04 <sup>C</sup> b	0.70±0.04 <sup>C</sup> a	0.60±0.01 <sup>C</sup> a
	15	0.90±0.02 <sup>C</sup> a	0.80±0.03 <sup>B</sup> c	0.90±0.04 <sup>B</sup> b	1.00±0.04 <sup>B</sup> b
	30	1.00±0.03 <sup>B</sup> a	1.10±0.05 <sup>A</sup> a	1.00±0.01 <sup>A</sup> Ba	1.10±0.04 <sup>B</sup> a
	45	1.10±0.01 <sup>A</sup> ab	1.00±0.03 <sup>A</sup> b	1.10±0.03 <sup>A</sup> ab	1.30±0.04 <sup>A</sup> a
	60	1.10±0.02 <sup>A</sup> b	1.00±0.03 <sup>A</sup> b	1.00±0.04 <sup>A</sup> Bb	1.40±0.04 <sup>A</sup> a
Syneresis	1	2.00±0.05 <sup>A</sup> a	1.00±0.11 <sup>A</sup> b	0.50±0.05 <sup>A</sup> c	0.00±0.00 <sup>A</sup> d
	15	1.50±0.11 <sup>B</sup> a	1.00±0.11 <sup>A</sup> b	0.50±0.05 <sup>A</sup> c	0.00±0.00 <sup>A</sup> d
	30	1.00±0.05 <sup>C</sup> a	0.80±0.11 <sup>A</sup> a	0.50±0.05 <sup>A</sup> b	0.00±0.00 <sup>A</sup> d
	45	1.00±0.11 <sup>C</sup> a	0.70±0.05 <sup>A</sup> b	0.30±0.11 <sup>A</sup> c	0.00±0.00 <sup>A</sup> d
	60	1.10±0.11 <sup>C</sup> a	0.60±0.02 <sup>B</sup> b	0.50±0.11 <sup>A</sup> b	0.00±0.00 <sup>A</sup> d
$L^*$	1	70.11±3.56 <sup>A</sup> a	69.88±3.99 <sup>A</sup> a	66.00±3.02 <sup>A</sup> a	67.66±2.77 <sup>A</sup> a
	15	-3.22±0.46 <sup>AB</sup> b	-3.11±0.75 <sup>B</sup> b	-0.11±0.69 <sup>A</sup> a	-0.66±0.72 <sup>A</sup> a
	30	14.00±1.09 <sup>B</sup> b	17.33±0.66 <sup>A</sup> a	15.33±0.83 <sup>B</sup> a	17.66±1.67 <sup>A</sup> a
	45	61.66±2.04 <sup>A</sup> a	65.88±3.04 <sup>A</sup> a	70.33±3.60 <sup>A</sup> a	68.55±3.14 <sup>A</sup> a
	60	-0.55±0.50 <sup>AB</sup> b	-1.22±0.77 <sup>B</sup> b	0.88±0.38 <sup>A</sup> a	0.44±0.64 <sup>A</sup> a
$b^*$	1	17.88±0.45 <sup>A</sup> a	18.00±0.50 <sup>A</sup> a	17.55±0.62 <sup>A</sup> a	17.00±0.76 <sup>A</sup> a
	15	60.88±4.99 <sup>A</sup> a	59.66±3.75 <sup>B</sup> a	63.77±3.00 <sup>A</sup> a	57.88±3.84 <sup>B</sup> a
	30				
	45				
	60				

$T$	Day	Control	1%	2%	3%
a*	30	-0.33±0.33 <sup>A</sup> a	-0.44±0.41 <sup>A</sup> a	0.44±0.24 <sup>A</sup> a	-0.11±0.26 <sup>A</sup> a
	45	14.44±1.34 <sup>AB</sup> b	18.22±0.68 <sup>A</sup> a	18.33±1.08 <sup>A</sup> a	18.00±0.94 <sup>A</sup> a
	60	63.22±3.40 <sup>A</sup> a	64.11±1.82 <sup>AB</sup> a	62.44±2.61 <sup>A</sup> a	65.66±3.04 <sup>AB</sup> a
b*	30	-2.00±0.44 <sup>B</sup> a	-0.66±0.64 <sup>A</sup> a	-0.66±0.33 <sup>A</sup> a	-0.55±0.58 <sup>A</sup> a
	45	15.55±0.62 <sup>AB</sup> a	17.66±0.40 <sup>A</sup> a	17.55±0.68 <sup>AB</sup> a	17.22±0.95 <sup>A</sup> a
	60	65.11±2.63 <sup>A</sup> a	57.11±2.81 <sup>B</sup> a	63.33±5.09 <sup>A</sup> a	62.88±5.09 <sup>AB</sup> a
L*	30	-3.00±0.50 <sup>AB</sup> b	0.11±0.65 <sup>A</sup> a	-0.33±0.66 <sup>A</sup> a	-0.11±0.51 <sup>A</sup> a
	45	17.33±1.57 <sup>AB</sup> a	17.11±0.45 <sup>A</sup> a	16.77±0.72 <sup>AB</sup> a	15.33±0.72 <sup>A</sup> a
	60				

**Notes:**  $L^*$  ranges from 0 (black) to 100 (white),  $a^*$  ranges from red (+  $a^*$ ) to green (-  $a^*$ ) and  $b^*$  ranges from yellow (+  $b^*$ ) to blue (-  $b^*$ );  
 ‡ Different capital letters in the column (A-C) differ significantly ( $P \leq 0.05$ ); different letters (a-c) in the row differ significantly ( $P \leq 0.05$ ).

### Lysin assessment

The Maillard (nonenzymatic glycation) is described as a complicated chemical reaction that may occur between the amino and the carbonyl groups which gives a browned appearance to some foodstuffs. This reaction usually develops during food processing and/or storage. Generally, in the Maillard reaction, lactose essentially reacts with the Gamma-amino group of residual lysine that exists in milk proteins. Thus, the degree of the disappearance of lysine is correlated with the severity of heat treatment (Shimamura & Ukeda, 2012). In this study, the lysin content in quinoa flour, yogurt without any heat treatment, control kashk and kashk containing 3% quinoa were 308.29, 80.23, 55.33, 89.07 ppm separately (Figure 4). Thus, adding quinoa flour to kashk can compensate lysine lost in the kashk thermal process.



**Figure 4** High-performance liquid chromatograms of the Lysin. Quinoa flour (a), yogurt before heat process (b), control kashk (c), kashk containing 3% quinoa (d).

### Apparent Viscosity

The apparent viscosity of quinoa supplemented kashk samples is presented in Table 3. The power law, Bingham and Casson models were found appropriate in this study to express the following behavior of kashk samples. The non-Newtonian behavior occurred in the following manner a reduction in the apparent viscosity of the kashk samples with increasing shear rate (Figure 5). Following a rapid reduction, the apparent viscosity changed slightly and became stable at higher shear rates. This could be due to the decrease in particle size as the consequence of increasing the shear rate (Sekhvatizadeh et al., 2019). The viscosity increased during the storage time among all samples. It is worth mentioning that post



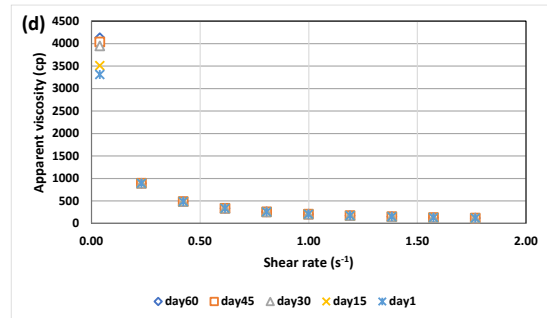
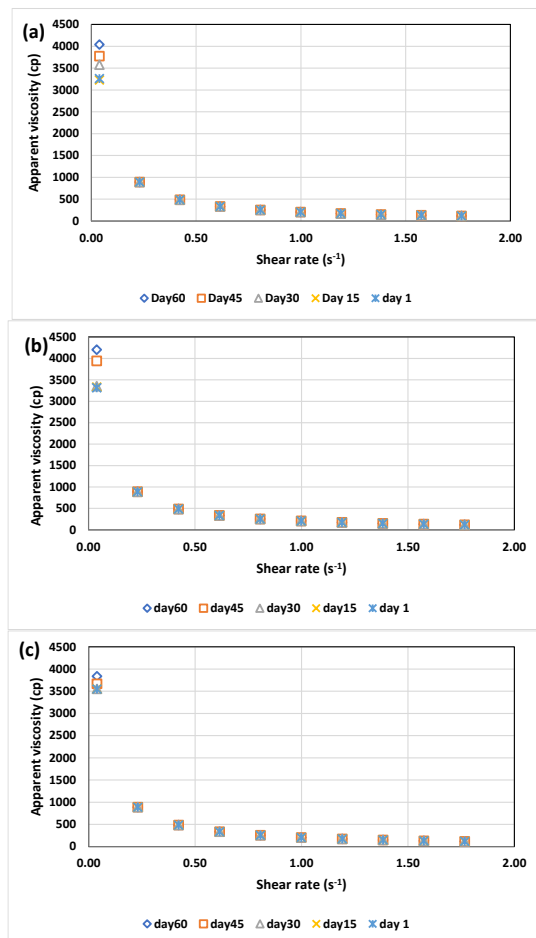
acidification caused the deterioration of the negative electric charge of the casein micelles resulted from dissolving the calcium and inorganic phosphate which attenuates colloidal stability and subsequently the casein becomes insoluble near its isoelectric pH (about 4.6). This phenomenon, strengthening of protein-protein complexes leads to enhance viscosity (Dönmez *et al.*, 2017). For the power law, Bingham and Casson models, the determination coefficient ( $R^2$ ) values were nearly 0.95, 0.93, 0.96 which showed the appropriate findings for these models (Table 3). As can be viewed, no significant differences were found in the flow index values between all samples at the initial and end of the storage ( $p > 0.05$ ). In general, the lower flow index ( $n$ ) values ( $<1$ ) and higher consistency coefficient cause the occurrence of pseudo-plastic (non-Newtonian) materials at every concentration of quinoa flour (Yang *et al.*, 2012).

Yield stress is suggested as a significant quality control value in the industrial procedures, typically for matching the comprehensive features of products made on different production sites (Haminiuk *et al.*, 2006).

The shear rate ranged between 3.84 to 176.64 ( $s^{-1}$ ) as resulted from the influence of quinoa flour proportions on shear stress (cP) of kashk (151.33 to 127.22 cP). A decrease in the level of shear stress values was denoted a clear decline in the shear stress value after increase the concentration of quinoa flour from 1 to 3%. These findings could result from the gelation of quinoa flour which formed from the aggregation of casein-particle. Fewer intermolecular bonds formed, due to the lack of enough interstitial solvents, after the addition of quinoa to the samples (Codină *et al.*, 2016).

In the power-law equation, the results showed that the control kashk and the sample containing 3% quinoa flour had the highest consistency coefficient. But 1% quinoa flour sample had the lowest, on the first day of storage. While at the end of the storage, the kashk sample contained 3% quinoa flour and the control had the highest; the lowest flow index was at 15681 and 13601 (cP), respectively. The higher value of  $k$  indicates that a higher firmness of the samples that occur after limiting molecular movement had increased as a result of the interaction between polymer chains (Karazhiyan *et al.*, 2009).

The results (Casson and Bingham models) showed that a high level of supplemented quinoa flour (3%) had a lower plastic viscosity in the samples upon 60 days of the storage time. An increase in the viscosity of samples was observed following the addition of up to 1.0% of quinoa flour. This was likely associated with the increase in bond water in the samples. This is beneficial as the product may keep its stability for a longer period (Codină *et al.*, 2016).



**Figure 5** The apparent viscosity of the kashk samples containing different quinoa flour during storage. Control (a); 1% quinoa flour (b); 2% quinoa flour (c); 3% quinoa flour (d).

**Table 3** Viscosity parameters of kashk fortified with quinoa

Viscosity parameter	Day	Control	1%	2%	3%
Bingham	$\eta_p$ (cP)				
	1	14.14 $\pm$ 5.25Aa	21.55 $\pm$ 0.35Aa	19.50 $\pm$ 1.90Aa	14.20 $\pm$ 3.70Ba
	15	20.20 $\pm$ 2.80Aa	20.65 $\pm$ 0.35Aa	19.50 $\pm$ 0.10Aa	20.00 $\pm$ 0.90ABa
	30	17.00 $\pm$ 3.40Aa	19.60 $\pm$ 0.30Aa	17.30 $\pm$ 1.10Aa	15.80 $\pm$ 0.60ABa
	45	22.65 $\pm$ 0.55Aa	15.22 $\pm$ 6.17Aa	16.35 $\pm$ 1.55Aa	22.10 $\pm$ 1.10Ba
	60	18.05 $\pm$ 0.15Aa	12.37 $\pm$ 4.43Aa	18.25 $\pm$ 3.05Aa	13.20 $\pm$ 1.10Ba
Bingham	$\tau_{0B}$ (D $\cdot$ cm $^{-2}$ )				
	1	187.20 $\pm$ 6.60Aa	177.80 $\pm$ 0.20Aa	180.60 $\pm$ 2.40Aa	187.20 $\pm$ 4.70Aa
	15	179.65 $\pm$ 3.55Aa	179.10 $\pm$ 0.50Aa	180.55 $\pm$ 0.15Aa	179.95 $\pm$ 1.15ABa
	30	183.65 $\pm$ 4.25Aa	180.45 $\pm$ 0.35Aa	183.35 $\pm$ 1.35Aa	185.15 $\pm$ 0.75ABa
	45	176.50 $\pm$ 0.70Aa	185.85 $\pm$ 7.75Aa	184.50 $\pm$ 1.90Aa	177.25 $\pm$ 1.35Ba
	60	180.95 $\pm$ 1.25Aa	189.45 $\pm$ 5.55Aa	182.10 $\pm$ 3.80Aa	188.45 $\pm$ 1.35Aa
Casson	$R^2$				
	1	94.85 $\pm$ 2.15Aa	91.65 $\pm$ 0.05Aa	92.65 $\pm$ 0.85Aa	94.90 $\pm$ 1.50Aa
	15	92.30 $\pm$ 1.30Aa	92.15 $\pm$ 0.15Aa	92.65 $\pm$ 0.05Aa	92.45 $\pm$ 0.45ABa
	30	93.75 $\pm$ 1.45Aa	92.65 $\pm$ 0.15Aa	93.65 $\pm$ 0.45Aa	94.30 $\pm$ 0.30ABa
	45	91.15 $\pm$ 0.25Aa	94.35 $\pm$ 2.55Aa	94.05 $\pm$ 0.65Aa	91.45 $\pm$ 0.55Ba
	60	92.50 $\pm$ 0.80Aa	95.60 $\pm$ 1.70Aa	93.20 $\pm$ 1.40Aa	95.35 $\pm$ 0.45Aa
Casson	$\eta_c$ (cP)				
	1	1.40 $\pm$ 0.96Aa	3.03 $\pm$ 0.06Aa	2.42 $\pm$ 0.52Aa	1.31 $\pm$ 0.68BCa
	15	2.67 $\pm$ 0.81Aa	2.72 $\pm$ 0.11Aa	2.39 $\pm$ 0.03Aa	2.52 $\pm$ 0.25ABa
	30	1.87 $\pm$ 0.78Aa	2.41 $\pm$ 0.07Aa	1.84 $\pm$ 0.25Aa	1.50 $\pm$ 0.12BCa
	45	3.36 $\pm$ 0.19Aa	1.71 $\pm$ 1.25Aa	1.64 $\pm$ 0.33Aa	3.18 $\pm$ 0.35Aa
	60	2.10 $\pm$ 0.06Aa	1.04 $\pm$ 0.69Aa	2.14 $\pm$ 0.76Aa	1.03 $\pm$ 0.18Ca
Power law	YS (D $\cdot$ cm $^{-2}$ )				
	1	173.20 $\pm$ 12.10Aa	155.75 $\pm$ 0.45Aa	160.90 $\pm$ 4.50Aa	173.20 $\pm$ 8.60Aa
	15	159.15 $\pm$ 6.65Aa	158.20 $\pm$ 0.90Aa	160.90 $\pm$ 0.30Aa	159.80 $\pm$ 2.10ABa
	30	166.65 $\pm$ 7.95Aa	160.70 $\pm$ 0.60Aa	166.10 $\pm$ 2.50Aa	169.55 $\pm$ 1.45ABa
	45	153.30 $\pm$ 1.30Aa	170.65 $\pm$ 14.35Aa	168.25 $\pm$ 3.55Aa	154.70 $\pm$ 2.60Ba
	60	162.80 $\pm$ 2.30Aa	177.35 $\pm$ 10.15Aa	163.85 $\pm$ 7.15Aa	175.50 $\pm$ 2.50Aa
Power law	$R^2$				
	1	97.60 $\pm$ 1.00Aa	96.15 $\pm$ 0.05Aa	96.60 $\pm$ 0.70Aa	97.60 $\pm$ 0.70Aa
	15	96.45 $\pm$ 0.55Aa	96.40 $\pm$ 0.10Aa	96.60 $\pm$ 0.00Aa	96.50 $\pm$ 0.20ABa
	30	97.10 $\pm$ 0.70Aa	96.60 $\pm$ 0.10Aa	97.05 $\pm$ 0.25Aa	97.35 $\pm$ 0.15ABa
	45	95.95 $\pm$ 0.15Aa	97.40 $\pm$ 1.20Aa	97.20 $\pm$ 0.30Aa	96.05 $\pm$ 0.25Ba
	60	96.85 $\pm$ 0.05Aa	98.00 $\pm$ 0.80Aa	96.85 $\pm$ 0.65Aa	97.80 $\pm$ 0.20Aa
Power law	$K_p$ (cP)				
	1	15339.50 $\pm$ 1909.50Aa	12620.00 $\pm$ 67.00Aa	13412.00 $\pm$ 687.00Aa	5318.00 $\pm$ 1353.00Aa
	15	13146.00 $\pm$ 1011.00Aa	12991.50 $\pm$ 133.50Aa	13401.50 $\pm$ 47.50Aa	3239.50 $\pm$ 324.50ABa
	30	14302.50 $\pm$ 1234.00Aa	13373.50 $\pm$ 95.50Aa	14204.50 $\pm$ 392.50Aa	4740.50 $\pm$ 220.50ABa
	45	12258.50 $\pm$ 199.50Aa	14948.50 $\pm$ 2242.00Aa	14539.50 $\pm$ 555.50Aa	12468.00 $\pm$ 390.00Ba
	60	13601.00 $\pm$ 288.00Aa	15982.00 $\pm$ 1615.00Aa	13862.00 $\pm$ 1099.00Aa	15681.00 $\pm$ 394.00Aa
Power law	$n$				
	1	0.065 $\pm$ 0.025Aa	0.110 $\pm$ 0.000Aa	0.090 $\pm$ 0.010Aa	0.065 $\pm$ 0.015BCa
	15	0.095 $\pm$ 0.015Aa	0.100 $\pm$ 0.000Aa	0.090 $\pm$ 0.000Aa	0.095 $\pm$ 0.005ABa
	30	0.080 $\pm$ 0.020Aa	0.095 $\pm$ 0.005Aa	0.080 $\pm$ 0.010Aa	0.075 $\pm$ 0.005BCa
	45	0.115 $\pm$ 0.005Aa	0.070 $\pm$ 0.030Aa	0.075 $\pm$ 0.005Aa	0.110 $\pm$ 0.10Aa
	60	0.090 $\pm$ 0.000Aa	0.055 $\pm$ 0.025Aa	0.085 $\pm$ 0.015Aa	0.055 $\pm$ 0.005Ca
Power law	$R^2$				
	1	96.20 $\pm$ 1.55Aa	93.95 $\pm$ 0.05Aa	94.65 $\pm$ 0.65Aa	96.30 $\pm$ 1.10Aa
	15	94.40 $\pm$ 1.00Aa	94.30 $\pm$ 0.10Aa	94.65 $\pm$ 0.05Aa	94.50 $\pm$ 0.30ABa
	30	95.45 $\pm$ 1.05Aa	94.65 $\pm$ 0.05Aa	95.35 $\pm$ 0.35Aa	95.85 $\pm$ 0.15ABa
	45	93.60 $\pm$ 0.20Aa	95.90 $\pm$ 1.90Aa	95.70 $\pm$ 0.50Aa	93.80 $\pm$ 0.40Ba
	60	94.30 $\pm$ 0.80Aa	96.80 $\pm$ 1.30Aa	95.05 $\pm$ 0.95Aa	96.60 $\pm$ 0.30Aa

**Notes:** †Yield stress; ‡ plastic viscosity

$k_p$ : the Power law consistency coefficient,  $\eta_p$ : the Power law flow behavior index,  $\eta_B$ : the Bingham plastic viscosity,  $\tau_{0B}$ : the Bingham yield stress (D $\cdot$ cm $^{-2}$ ),  $\eta_c$ : Casson viscosity (cP), YS: Casson yield stress (D $\cdot$ cm $^{-2}$ ).

Different capital letters in the column (A-D) differ significantly ( $P \leq 0.05$ ); different letters (a-c) in the row differ significantly ( $P \leq 0.05$ ).

## Texture

The texture of kashk samples are shown in Table 4 indicated the fortifying kashk with 3.0% quinoa flour had a significant effect on the textural properties. Hardness

and gumminess increased and adhesiveness, chewiness, springiness and cohesiveness decreased significantly. The binding of proteins could lead to a rise in the firmness of samples during storage. However, the lowest values of hardness were occurred by the end of the storage (on day 60). The samples enriched with 3% of quinoa flour revealed the highest value of firmness (Hosseini & Ansari, 2019). An increase in the hardness of whole skimmed flavored yogurt after a long storage time was reported (Salvador & Fiszman, 2004). The increase in the hardness of different varieties of kashk samples observed in our study on account of the addition of quinoa flour may be associated with the increase in the dry matter content (Table 1). The presence of amylase/ amylopectin in the starch chain which contributed to the reactive groups in the quinoa flour may produce its emulsifying property which finally tended to a higher level of water absorption, the greater viscosity in the continuous phase and firmer texture in the treated kashk (Hosseini & Ansari, 2019). Moreover, hardness might be related to quinoa flour absorbing more moisture because of its higher water-holding capacity. The highest values of hardness are associated with a greater reorganization of proteins in the gel network. Cohesiveness is described by a large number of broken casein-casein connections during the stress circumstances together with springiness (the ability to recover its initial condition after applying a deformation) (Curti *et al.*, 2017). The Adhesion was significantly decreased during the storage. It is supposed that the long refrigerating storage of kashk samples made amylose more likely to bind casein micelles in quinoa flour, and thus a stronger three-dimensional network was formed with more stiffness which caused reducing the process of adhesion. This result was supported by (Hosseini & Ansari, 2019) regarding three forms of modified tapioca starch which were incorporated into industrial liquid kashk. The result of their study revealed a significant decrease in the adhesiveness and a simultaneous increase in the hardness of samples ( $p \leq 0.05$ ). An adverse relationship between adhesiveness and hardness was therefore reported.

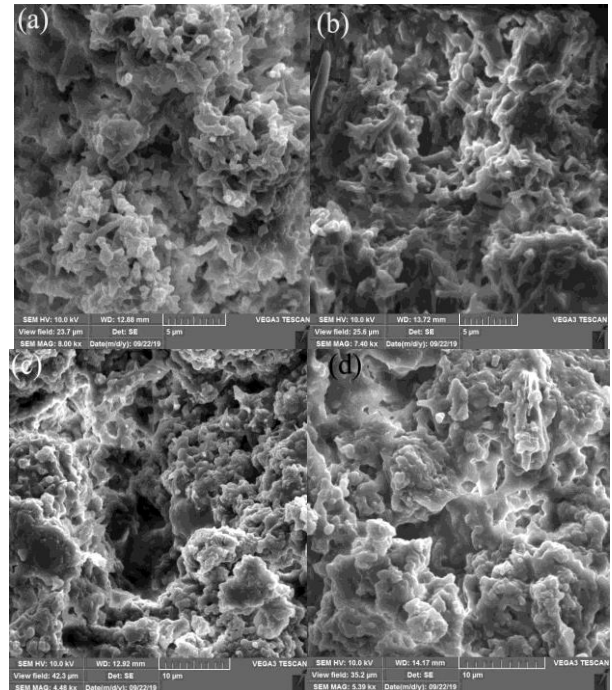
**Table 4** Texture parameters of kashk fortified with quinoa

†T	Day	Control	1%	2%	3%
Hardness	1	0.065±0.001Dd	0.215±0.03Ac	0.356±0.03Bb	0.721±0.03Ba
	15	0.125±0.001Cd	0.213±0.001Ac	0.367±0.001Bb	0.814±0.00Ba
	30	0.141±0.001BCd	0.291±0.01Ac	0.384±0.02Bb	0.871±0.01Ba
	45	0.157±0.001Bb	0.605±0.31Aab	0.579±0.001Aab	1.051±0.02Aa
	60	0.200±0.01Ac	0.325±0.01Ac	0.577±0.01Ab	1.114±0.08Aa
Adhesiveness	1	0.096±0.001Ba	0.188±0.001Ab	0.568±0.01Bc	0.200±0.02Ab
	15	0.165±0.001Ba	0.375±0.05ABb	0.582±0.02Bc	0.350±0.05Ab
	30	0.178±0.001Ba	0.448±0.01Bb	0.305±0.02Ac	0.550±0.05Ac
	45	0.108±0.001Aa	0.532±0.02Bb	0.575±0.02Bb	0.617±0.10Ab
	60	0.384±0.02Ca	0.308±0.13ABa	0.721±0.11Ba	0.417±0.30Aa
Chewiness	1	0.48±0.001Dc	0.131±0.001Ab	0.206±0.01Ba	0.153±0.02Cb
	15	0.065±0.001CDc	0.135±0.03Abc	0.225±0.001Bab	0.247±0.03Ba
	30	0.079±0.001Cb	0.154±0.001Aab	0.175±0.04Ba	0.141±0.00Cab
	45	0.108±0.00Bd	0.184±0.001Ac	0.161±0.001Bb	0.320±0.00AB <sub>a</sub>
	60	0.145±0.001Ab	0.133±0.04Ab	0.347±0.001Aa	0.344±0.02Aa
Cohesiveness	1	0.818±0.001Aa	0.663±0.08Aa	0.634±0.01Aa	0.322±0.03AB <sub>b</sub>
	15	0.675±0.02Aa	0.687±0.15Aa	0.656±0.03Aa	0.437±0.05Aa
	30	0.636±0.08Aa	0.592±0.04Aa	0.536±0.02Aa	0.276±0.00Bb
†T	Day	Control	1%	2%	3%
Cohesiveness	45	0.745±0.05Aa	0.713±0.05Aa	0.330±0.02Bb	0.340±0.02AB <sub>b</sub>
	60	0.799±0.001Aa	0.458±0.07Ab	0.667±0.05Aa	0.399±0.03AB <sub>b</sub>
Gumminess	1	0.052±0.001Dc	0.139±0.001Ab	0.225±0.01Ba	0.231±0.01Ba
	15	0.083±0.001Cc	0.146±0.03Abc	0.240±0.01Bb	0.356±0.04AB <sub>a</sub>
	30	0.089±0.001Cc	0.171±0.001Ab	0.206±0.02Bab	0.236±0.02AB <sub>a</sub>
	45	0.116±0.001Bc	0.201±0.001Ab	0.191±0.01Bb	0.356±0.02AB <sub>a</sub>
	60	0.159±0.001Ac	0.150±0.03Ac	0.862±0.001Aa	0.446±0.06Ab
Springiness	1	0.936±0.001Aa	0.945±0.01Aa	0.917±0.01Aa	0.678±0.14AB <sub>a</sub>
	15	0.918±0.001ABa	0.926±0.01Aa	0.938±0.01Aa	0.693±0.00AB <sub>b</sub>
	30	0.896±0.02Ba	0.903±0.02Aa	0.840±0.10Aa	0.598±0.03Bb
	45	0.933±0.001ABa	0.918±0.001Aa	0.850±0.05Aa	0.905±0.06Aa
	60	0.913±0.001ABa	0.870±0.09Aa	0.862±0.001Aa	0.780±0.06AB <sub>a</sub>

Different capital letters in the column (A-D) differ significantly ( $P \leq 0.05$ ); different letters (a-c) in the row differ significantly ( $P \leq 0.05$ ). The texture parameter consist of Hardness (kg), Adhesiveness (kg·s<sup>-1</sup>), Chewiness (kg·s<sup>-1</sup>), Cohesiveness, Gumminess (kg), Springiness (mm). †Texture parameters

## Scanning electron microscopy

Details of the scanning electron micrograph of kashk samples supplemented with various concentrations of quinoa flour are given in Figure 6. A relatively denser appearance was revealed in the protein matrices of the supplemented samples that resulted in improved gel strength and lessen syneresis. This was confirmed by the textural, sensory and syneresis data (Tables 4, 2 and Figure 3). The micrograph displayed the highest strength of 3% quinoa flour supplemented kashk gels from cross-linked, which is due to a well-organized protein network with smaller pores in the product (Farnsworth *et al.*, 2006). The control kashk sample network had the most pores or void spaces in which the aqueous phase was retained (Lee & Lucey, 2010).



**Figure 6** Scan electron microscopy images of kashk samples. Control (a); 1% quinoa (b); 2% quinoa (c); 3% quinoa (d).

## CONCLUSIONS

In the present study, overall acceptability and lower syneresis of the quinoa-supplemented product were observed. The addition of quinoa flour in yogurt intensified the consistency although the flavor was higher in kashk. The addition of quinoa flour had not altered the odor and color properties of kashk samples. SEM micrographs demonstrated that 3% of quinoa flour could be used in kashk to improve the gel microstructure. Moreover, reducing lysine in the kashk heat process is compensated by the addition of quinoa flour. The results of this study promisingly provide opportunities to apply quinoa flour in food industries. Future research can investigate the possible function of quinoa flour on other dairy products. Specifically, the potential for sample profitability of different variants of kashk supplemented with quinoa flour can also be investigated.

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