

APPLICATIONS OF PLANT LECTINS IN BIOTECHNOLOGY AND THERAPEUTICS

Sanjay Naik, Sanjit Kumar*

Address(es):

Vellore Institute of Technology (VIT), Centre for Bio-Separation Technology (CBST), 632014 Vellore, Tamil Nadu, India. Tel.: +91- 416 220 2374

*Corresponding author: sanjitkrroy@gmail.com sanjitkumar@vit.ac.in

<https://doi.org/10.55251/jmbfs.4224>

ARTICLE INFO

Received 16. 1. 2021
Revised 13. 8. 2021
Accepted 3. 9. 2021
Published 1. 2. 2022

Review



ABSTRACT

Lectins are carbohydrate-binding proteins broadly used in various therapeutic and biomedical applications. The variable affinity of lectins towards variety of carbohydrates has raised attention for the biologist to explore functional aspects of lectins. Lectins express specificity to simple carbohydrates for example mannose, lactose, sialic acid, complex glycan, and glycoproteins. Lectins are classified based on their sugar specificity and are used as a tool to study protein-carbohydrate interactions. Lectins are ubiquitous in nature and identified from all sources such as bacteria, fungi, algae, and animals. Plants are the most abundant source of lectins, and till now, more than three hundred lectins were characterized from plants. These are distributed to various parts of a plant according to their requirements and function. The physiological role of lectins in a plant is still not well understood. The overabundant presence of lectins in plant seeds and storage tissues indicated their role in plant development. Plant lectins shows a broad range of activities like antibacterial, antifungal, insecticidal, anticancerous, antileishmanial, antiviral, and anticoagulants. In this review, we aim to highlight the plant lectins classification and their application in various biological aspects.

Keywords: Lectins, Plant lectins, Sugar specificity, Therapeutic aspect

INTRODUCTION

Lectins are glycoprotein with a divers function capable to bind specific carbohydrates without modifying them (Gemeiner *et al.*, 2009). Lectins are non-immune origin differ from antibody in many aspects. Antibodies are structurally similar exclusively present in animals whereas lectins are structurally different from each other and also present in plants and bacteria where no immunological system present (Sharon, 2008). In the late 18th century Peter Hermann Stillmark observed that the protein from plants can agglutinate erythrocytes and it was named Phytohemagglutinin {(Phyto (plant)+ Hemagglutinin (red blood cells agglutinate))} (Sharon *et al.*, 2004). These observations led to the discovery of the first phytohemagglutinin from seeds of the castor plant which was named ricin (Polito *et al.*, 2019). In 1954 Boyd and Shapleigh recognized that phytohemagglutinins were selectively agglutinate different human blood groups ABO, because of its sugar specificity. The selective nature of phytohemagglutinin named as lectin from the Latin verb that's means "to select" proposed by Boyd and Shapleigh (Hou *et al.*, 2003). Lectins are isolated and characterized by almost all five kingdoms (Monera, Protista, Fungi, Plantae, and Animalia) (Naik *et al.*, 2017). In case of plants, the source of lectins are roots, bulbs, pulps, tubers, rhizome, latex, barks, stems, leaves, flowers, fruits, and seeds (Table 1). Lectins are extracted from different parts of the plants have a wide range of applications in biochemistry, plant biology, agriculture, cell biology, immunology, virology, and biomedical field. The physiological role of lectins in plants remains unclear or controversial. Differences in structural and sugar specificity indicates lectins unlikely to have similar functions inside the plant. Insecticidal, antibacterial effect of plant lectins demonstrated that lectins play crucial role in plant defense system. More in detail lectins show potential role as antibacterial, antifungal, anti-HIV, antitumor, cytotoxic, insecticidal, immunomodulatory, vasorelaxant, histochemical, tumor detection marker, and pathology. This review is mainly on specificity, classification and application of plant lectins.

Table 1 Source of plant lectins

S.No.	Plants	Source
1	<i>Pinellia ternata</i>	Tubers
2	<i>Acacia farnesiana</i>	Seeds
3	<i>Sambucus ebulus L</i>	shoot
4	<i>mulberry</i>	leaf
5	<i>Microgramma vacciniifolia</i>	rhizome
6	<i>Euphorbia trigona</i>	latex
7	<i>Hyacinth</i>	bulbs
8	<i>Punica granatum</i>	fruits
9	<i>Moringa oleifera</i>	Seed cake
10	<i>Sophora flavescens</i>	roots
11	<i>Alpinia purpurata</i>	inflorescences
12	<i>Crataeva tapia</i>	bark
13	<i>Cucumis sativus</i>	phloem exudate

Specificity and Classification of Plant lectin

In recent years mannose-specific lectins were extensively characterized from monocot plants. Sugar specific hemagglutination inhibition assay ensured the specificity of lectins. *Lonchocarpus campestris* lectin (De Freitas Pires *et al.*, 2019), *Diocleinae* lectin (Leal *et al.*, 2018), and *Sauromatum guttatum* lectin (Thakur *et al.*, 2017) inhibited by mannose (Table 2). Structure studies showed Phe, Trp or Tyr mainly formed interaction with mannose. Entadin lectin (Naik *et al.*, 2020) and *Genipa americana* bark lectin (Costa *et al.*, 2018) were inhibited by lactose sugar. Simple sugar needs higher concentration to inhibits agglutination activity of lectin whereas complex sugar or carbohydrates need lesser concentration (Siritapetawee *et al.*, 2018). Hemagglutination assay is a benchmark to identify lectin from any source and the use of erythrocytes in this assay indicates lectins are specific for different erythrocytes. The lectins from *Sambucus ebulus L* (Jiménez *et al.*, 2013), *Morus alba* (white mulberry) (Deepa, *et al.*, 2012), and *Tinospora tomentosa* (Saha *et al.*, 2014) agglutinate human erythrocytes whereas lectins from, *Lonchocarpus campestris* seed (De Freitas Pires *et al.*, 2018), *Genipa americana* bark (Costa *et al.*, 2018), and *Coccinia indica* fruits (Bobbili *et al.*, 2018) agglutinate rabbit erythrocytes. Lectins from *Praecitrullus fistulosus* fruit (Shivamadhhu *et al.*, 2017), *Moringa oleifera* seed (Asaduzzaman *et al.*, 2018), *Kaempferia rotunda* tuberous rhizome (Ahmed *et al.*, 2018).

al., 2017) and *Crotalaria retusa* seeds (Aragão et al., 2017) agglutinate human as well as rabbit erythrocytes. Due to diversity of lectin in respect to function and origin its classification is still not completely evolved. Based on the overall structure and carbohydrate-binding site lectin can be divided into four types i. Merolectins: They are low molecular weight proteins consist of only one carbohydrate domain example Hevein, chitin-binding protein from the latex of the rubber tree (*Hevea brasiliensis*) ii. Hololectins: They contain more than two carbohydrate-binding sites. Hololectins are divalent or multivalent which help to agglutinate cells. Most of the plant lectins belong to this category. iii.

Chimerolectins: They are chimeric proteins composed of carbohydrate-binding domain and catalytic activity domain. Depending upon binding sites they behave as merolectin or hololectin. iv. Superlectins: superlectin can also be considered as chimerolectins consist exclusively of at least two carbohydrate-binding sites, which recognize structurally two unrelated sugars. During the last three decades, advanced biotechnology techniques help to understand the structure and function of various forms of lectin. Based on structural and functional aspects plant lectins are classified into seven different categories (Table 3).

Table 2 Sugar specificity of plant lectin

S.No.	Sugar specificity	Lectin
1	Mannose	<i>Lonchocarpus campestris</i> , <i>Diocleinae</i> , <i>Sauromatum guttatum</i> , <i>Pinellia ternata</i> , wheat, <i>Allium sativum</i> , <i>Microgramma vacciniifolia</i> , <i>Colocasia esculenta</i> , <i>Dioclea lasiophylla</i> mart, <i>Centrolobium microchaete</i> Mart, <i>Machaerium acutifolium</i>
2	Lactose	<i>Genipa americana</i> , <i>Spatholobus Parviflorus</i> , <i>Tinospora tomentosa</i> , Soybean, <i>Trichosanthes anguina</i> , <i>Abrus fruticulosus</i> , <i>Genipa Americana</i>
3	Galactose	<i>Nicotiana benthamiana</i> , <i>Bauhinia unguolata</i> L., <i>Lotus corniculatus</i> , <i>Champedak</i> , <i>Euphorbia trigona</i> Miller, <i>Clathrotropis nitida</i> , <i>Dioclea grandiflora</i> , <i>Morus alba</i> L., <i>Phaseolus lunatus</i> billb, <i>Euphorbia tirucalli</i> , <i>Euphorbia antiquorum</i> , <i>Artocarpus hypargyreus hance</i>
4	Glucose and mannose	<i>Canavalia brasiliensis</i> , <i>Pisum sativum</i> , <i>Dolichos lablab</i> , litchi, <i>Crataeva tapia</i> , <i>Microgramma vacciniifolia</i> , <i>Parkia platycephala</i>
5	n-Acetyl glucosamine	<i>Praecitrullus fistulosus</i> , <i>Apuleia leiocarpa</i> , White kidney bean, <i>Bauhinia forficata</i> , <i>Alpinia purpurata</i> , <i>Remusatia vivipara</i> ,
6	Glycoproteins	<i>Colocasia esculenta</i> , <i>Solanum tuberosum</i> , <i>Lagenaria siceraria</i> , <i>Dypsis decaryi</i>
7	Arabinose	<i>Anacardium occidentale</i>
8	D-galactosamine	<i>Phaseolus lunatus</i> L., <i>cicer aritium</i>
9	Chitin	<i>Cucumis sativus</i> , <i>Moringa oleifera</i> , <i>Praecitrullus fistulosus</i>
10	Fructose	<i>Moringa oleifera</i>

Table 3 Properties and classifications of plant lectins

S.No.	Types of lectin	First discovered	Molecular weight range	Plant source	Sugar specificity	Application
1	Legume	<i>Robinia pseudoacacia</i>	20-42 kDa	<i>Acacia farnesiana</i> , <i>Abrus precatorius</i> , <i>Canavalia ensiformis</i> , <i>Canavalia Brasiliensis</i> , <i>Canavalia grandiflora</i> , <i>Canavalia maritime</i> , <i>Canavalia virosa</i> , <i>Centrolobium microchaete</i> , <i>Clathrotropis nitida</i> , <i>Cratylia argentea</i> , <i>Calliandra surinamensis</i> , <i>Dolichos lablab</i> , <i>Dioclea grandiflora</i> , <i>Dioclea sclerocarpa</i> , <i>Dioclea violacea</i> , <i>Dioclea reflexa</i> , <i>Dioclea lasiocarpa</i> , <i>Vatairea guianensis</i> , <i>Phaseolus vulgaris</i> , <i>Phaseolus acutifolius</i> , <i>Phaseolus lunatus</i> L., <i>Parkia biglobosa</i> , <i>Parkia platycephala</i> , <i>Platypodium elegans</i> , <i>Cratylia mollis</i> , <i>Cratylia argentea</i> , <i>Centrolobium tomentosum</i> , <i>Bauhinia unguolata</i> L., <i>Bauhinia forficata</i> , <i>Bauhinia monandra</i> , <i>Bauhinia variegata</i> , <i>Bauhinia Purpurea</i> , <i>Spatholobus Parviflorus</i> , <i>Cicer arietinum</i> , <i>Lotus</i> , <i>pisum sativum</i> , <i>Dioclea lasiophylla</i> ,	D-Galactose, Lactose, Methyl α-D-galactopyranoside, mannose, glucose,	Antibacterial, Anticancer, Plant defense, Vasorelaxant, Nimeticidal, Seed germination, Antifungal, Amaylase inhibitor, Anti-inflammatory,
2	Chitin-binding	<i>Solanum tuberosum</i>	4-70 kDa	<i>Bauhinia forficata</i> , <i>Microgramma vacciniifolia</i> , <i>Apuleia leiocarpa</i> , <i>Punica granatum</i> , <i>Moringa oleifera</i> , <i>Praecitrullus fistulosus</i>	Glycoprotein, Mannose, Chitin	Anticancer, Anticoagulant, Insecticidal, Antibacterial,
3	Type-2 RIP	<i>Ricinus communis</i>	22-38 kDa	<i>Sambucus ebulus</i> L., <i>Abrus fruticulosus</i> ,		Toxic,
4	Monocot mannose specific	<i>Allium sativum</i>	12-30 kDa	<i>Pinellia ternate</i> , <i>Triticum aestivum</i> , <i>Hippeastrum</i> , <i>Allium sativum</i> , <i>Allium chinense</i> , <i>Allium cepa</i> , <i>Colocasia esculenta</i> , <i>Dioscorea bulbifera</i>	mannose	Anticancer, Antibacterial, Insecticidal, Antifungal,
5	Jacalin-related	<i>Maclura pomifera</i>	Subunits of 2-14 kDa	Jack fruit	Galagtose/mannose	Anticancer
6	Amaranthin	<i>Amaranthus caudatus</i>	33 kDa	<i>Amaranth</i>	Not reported	Anticancer,
7	Cucurbitaceae	<i>Cucurbita maxima</i>	Subunits of 30-35 kDa	<i>Cucumis sativus</i> , <i>Cucurbita maxima</i> , <i>Trichosanthes anguina</i> , <i>Cucumis sativus</i> , <i>Lagenaria siceraria</i> , <i>Praecitrullus fistulosus</i> , <i>Coccinia indica</i> ,	Lactose,	Plant defence

i. **Chitin Binding lectins:** All the proteins in this category contain one hevein domain for the consideration of chitin-binding lectin. ii. **Cucurbitaceae phloem lectins:** These lectins are found in phloem exudates of *Cucurbita*, *Citrullus*, *Cucumis*, *Sechium*, *Luffa*, and *Coccinia* species. iii. **Monocot mannose-binding lectins:** Monocot mannose-binding lectins are extracted and characterised from monocotyledonous plants. Tuhe first monocot mannose binding lectin was characterized by a snowdrop bulb. iv. **Legume lectins:** These lectins are isolated from a large family of Leguminosae (Fabaceae). Till now more than 100 lectins are isolated from this family and most of the lectins are purified from seeds (Table 2). v. **Type 2 RIP and related lectins:** Ribosome-inactivating proteins (RIP) are inhibiting or inactivate eukaryotic ribosomes. RIP is divided in to two category

RIP-I and RIP-II. RIP-II consists of two chain A and B and that having carbohydrate binding sites with lectin activity (Table 2). vi. **Jacalin-related lectins:** Jacalin is a plant seed tetrameric protein specific to either mannose or galactose sugars. Lectins structurally (Similar domain or fold) and evolutionary related considered as jacalin related lectins. vii. **Amaranthin lectin family:** Amaranthin lectin was isolated purified from seed of *Amaranthus caudate*. Amaranthin lectins are homodimeric proteins composed of subunits of approximately 33 kDa, which are not glycosylate (Table 3).

Lectin Applications in Biotechnology

Antibacterial Activity

Since 2012, many of the lectins show inhibitory action against different bacteria, lectin from wheat (*Triticum aestivum*) was able to increase resistance to the infection of *Pseudomonas syringae* (Ma et al., 2013). *Acacia farnesiana* seeds lectin was able to inhibit *Xanthomonas axonopodis* pv. *passiflorae* (Gram-negative) and *Clavibacter michiganensis* (Gram-positive) (Santi-Gadelha et al., 2012). Lectins isolated from stem *Tinospora tomentosa* showed antibacterial properties against *Salmonella* (Saha et al., 2014). *Apuleia leiocarpa* seeds lectins showed antibacterial activity effects on the gram-positive bacteria *B. subtilis*, *B. cereus*, *E. faecalis*, *M. luteus*, *S. pyogenes*, and *S. aureus* with the smallest MIC (45.12 µg/ml). This lectin also showed inhibition against *X.campestris* pv. *campestris*, *X. campestris* pv. *viticola*, *X. campestris*pv. *malvacearum*, *Klebsiella pneumoniae*, *E. coli*, *P. aeruginosa*, and *S. enteritidis* with the lowest minimal inhibition concentration (11.2 µg/ml) against *X. campestris* pv. *Apuleia leiocarpa* lectin was bactericidal (Minimal bactericide concentration of 22.5 µg/ml) only against the three varieties of *X. campestris* (Carvalho et al., 2015). Over expression of lectin from *Arabidopsis* shows less disease symptoms after infection with *Pseudomonas syringae* when it's compare to the wild type of *Arabidopsis* (Van Hove et al., 2015). Lectin from *Bauhinia variegata* showed inhibition against oral bacteria facultatively anaerobic, gram-positive *Streptococcus mutans* and *Streptococcus sanguinis* (Klafke et al., 2016). Lectins from *Canavalia brasiliensis* and *Cratylia argentea* enhanced the antimicrobial immunity of macrophages experimentally infected with *S. typhimurium* (Silva et al., 2016; Batista et al., 2017). Chitin binding lectin from juicy sarcotesta of *Punica granatum* showed antibacterial activity against *Micrococcus luteus*, *Serratia*

marcescens, *Streptococcus mutans*, *Aeromonas* sp., *Enterococcus faecalis*, *Escherichia coli*, *Klebsiella* sp., *Salmonella enterica* serovar. *enteritidis*, *Staphylococcus aureus*, *Staphylococcus epidermidis* and *Staphylococcus saprophyticus*. It was observed that bacteria treated (*Aeromonas* sp., *S. aureus*, *S. marcescens* and *S. enterica*) with *Punica granatum* lectin having reduced adhesion and invasion abilities to HeLa cells were reduced when these bacteria were previously treated with (Silva et al., 2016). Lectin purified and characterized from leaves of *Calliandra surinamensis* showed growth inhibition against *Staphylococcus saprophyticus*, *Staphylococcus aureus* (nonresistant) and the *Staphylococcus aureus* (Oxacillin resistant) whereas against *E.coli* no inhibition was observed (Figueiredo et al., 2017). Lectin isolated from the latex of *Euphorbia tirucalli* was able to inhibit the growth of Gram-negative bacteria *Escherichia coli* (Palharini et al., 2017). The lectin from inflorescence of *Alpinia purpurata* showed bacteriostatic activity against non-resistant *Staphylococcus aureus* with minimal inhibitory concentrations (MIC50) of 50 µg/mL and Oxacillin resistant *Staphylococcus aureus* with minimal inhibitory concentrations (MIC50) of 400 µg/mL. It also showed bactericidal effect on the non-resistant *Staphylococcus aureus* with minimal bactericidal concentrations (MBC50) of 200 µg/mL (Ferreira et al., 2018). The lectin isolated from the crude latex of *Euphorbia antiquorum* L. can inhibits the growth of Gram-positive bacteria such as *Staphylococcus aureus* and *Staphylococcus epidermidis*, without killing them with a minimum inhibitory concentration (MIC) of 2000 µg/ml, *Propionibacterium acnes* (human skin commensal) with MIC of 125 µg/ml and *Streptococcus agalactiae* with MIC of 250 µg/ml. Whereas in case of gram negative bacteria *Samonella typhimurium* which inhibits with a MIC of 1000 µg/ml. (Siritapetawe et al., 2018) (Table 4).

Table 4 Antibacterial activity of plant lectins

Plants	Source	Against Bacteria
<i>Triticum aestivum</i>	Seed	<i>Pseudomonas syringae</i>
<i>Acacia farnesiana</i>	Seed	<i>Xanthomonas axonopodis</i> pv. <i>passiflorae</i> and <i>Clavibacter michiganensis</i>
<i>Tinospora tomentosa</i>	Stem	<i>Salmonella</i>
<i>Apuleia leiocarpa</i>	Seed	<i>B. subtilis</i> , <i>B. cereus</i> , <i>E. faecalis</i> , <i>M. luteus</i> , <i>S. pyogenes</i> , and <i>S. aureus</i> , <i>X. campestris</i> pv. <i>campestris</i> , <i>X. campestris</i> pv. <i>viticola</i> , <i>X. campestris</i> pv. <i>malvacearum</i> , <i>Klebsiella pneumoniae</i> , <i>E. coli</i> , <i>P. aeruginosa</i> , and <i>S. enteritidis</i>
<i>Bauhinia variegata</i>	Seed	<i>Streptococcus mutans</i> and <i>Streptococcus sanguinis</i>
<i>Canavalia brasiliensis</i>	Seed	<i>S. typhimurium</i>
<i>Cratylia argentea</i>	Seed	<i>S. typhimurium</i>
<i>Punica granatum</i>	Fruit	<i>Micrococcus luteus</i> , <i>Serratia marcescens</i> , <i>Streptococcus</i> , <i>Aeromonas</i> sp., <i>Enterococcus faecalis</i> , <i>Escherichia coli</i> , <i>Klebsiella</i> sp., <i>Salmonella enterica</i> serovar. <i>Enteritidis</i> , <i>Staphylococcus aureus</i> , <i>Staphylococcus epidermidis</i> and <i>Staphylococcus saprophyticus</i> .
<i>Calliandra surinamensis</i>	Leaf	<i>Staphylococcus saprophyticus</i> , <i>Staphylococcus aureus</i> and the <i>Staphylococcus aureus</i>
<i>Euphorbia tirucalli</i>	Latex	<i>Escherichia coli</i>
<i>Alpinia purpurata</i>	Inflorescence	<i>Staphylococcus aureus</i>
<i>Euphorbia antiquorum</i> L.	Latex	<i>Staphylococcus aureus</i> and <i>Staphylococcus epidermidis</i>

Anti-Bio-Film Activity

A Lectin from litchi (*Litchi chinensis*) seeds showed antibiofilm activity against *Pseudomonas aeruginosa* (Bose et al., 2016). Lectin isolated from the leaves of *Calliandra surinamensis* is an effective inhibitor of oral multispecies biofilms formed by bacteria from supragingival plaque (Figueiredo et al., 2017). Chitin binding lectin isolation and purified from pomegranate (*Punica granatum*) sarcotesta showed significant antibiofilm activity on *Candida albicans* (pathogenic yeast) at inhibitory concentrations of 0.195 and 0.39 µg/mL (Da Silva et al., 2018). The lectin remarkably inhibited biofilm formation by non-resistant *Staphylococcus aureus* isolate and *Candida albicans* from inflorescence of *Alpinia purpurata* at sub-inhibitory concentrations (Ferreira et al., 2018). *Bauhinia variegata* lectin showed anti-biofilm formation against *Streptococcus mutans* and *S. sanguinis* (Klafke et al., 2016).

Antifungal Activity

Chitin binding lectins are predominant in plants that bind to fungal cell wall which is made of chitin a complex polysaccharide. CBL lectins bind to chitin and stop the growth of fungus. Galctose binding lectin isolated from seeds of *Bauhinia unguolata* L. (*Caesalpinioideae*) showed antifungal activity against *Fusarium solani*, *Fusarium moniliforme*, *Fusarium oxysporum*, *Colletotrichum lindemuthianum* and *Aspergillus niger* (Silva et al., 2014). Lectin isolated and characterized from seed of *Spatholobus parviflorus* showed anti-fungal activity against *Aspergillus niger* and *Fusarium* sp. *Aspergillus niger* hyphal growth inhibition was seen at low concentration approx. 0.04 mg/mL of lectin. Whereas *Fusarium* sp. was inhibited at much higher lectin concentration (Geethanandan

et al., 2013). The lectin from *Phaseolus vulgaris* cv. Chinese pinto bean showed antifungal activity against mycelial growth in *Valsa mali* a destructive fungus of apple, by 30.6 % at 30 µM (Ang et al., 2014). The crude plant lectin isolated from the stem of *Tinospora tomentosa* shows fungal growth inhibition of *Aspergillus niger* and showed a clear inhibition of the growth of another fungal strains *saccharomyces cerevasecis* (Saha et al., 2014). Lectin isolated from leaf *Allium sativum* showed antifungal activity its gene was further expressed in an elite indica rice cv. IR64 by employing *Agrobacterium tumefaciens*-mediated transformation to engineer rice plants conferring resistance to sheath blight which mainly caused by *Rhizoctonia solan* (plant pathogenic fungus) (Ghosh et al., 2016). Lectin isolated from *Phaseolus lunatus* billb seeds inhibited the fungal growth of *Sclerotium rolfsii*, *Fusarium solani*, *Fusarium oxysporum*, and *Botrytis cinerea* (Wu et al., 2016). *Calliandra surinamensis* leaf lectin was assessed against four *Candida* species but only *Candida krusei* was sensitive to this lectin with MIC 125 µg/mL and MFC of 250 µg/mL (Figueiredo et al., 2017). The lectin from Sunflower seeds (*Helianthus annuus* L.) affects the viability of sunflower phytopathogenic fungus *Sclerotinia sclerotiorum* (Del Rio et al., 2018). The pomegranate (*Punica granatum*) sarcotesta contains a chitin-binding lectin showed antifungal activity against *Candida albicans* and *Candida krusei* with MIC50 of 25.0 and 12.5 µg/mL respectively (Da Silva et al., 2018). The lectin from inflorescence of *Alpinia purpurata* showed fungistatic activity against *Candida albicans* and *Candida parapsilosis* (Ferreira et al., 2018). Antifungal protein from *Cicer arietinum* seeds showed antifungal activity against *Candida parapsilosis*, *Candida krusei*, and *Candida tropicalis* (Kumar et al., 2014) (Table 5).

Table 5 Antifungal activity of plant lectins

Plants	Source	Against fungus
<i>Bauhinia unguolata</i> L.	Seed	<i>Fusarium solani</i> , <i>Fusarium moniliforme</i> , <i>Fusarium oxysporum</i> , <i>Colletotrichum lindemuthianum</i> and <i>Aspergillus niger</i>
<i>Spatholobus parviflorus</i>	Seed	<i>Aspergillus niger</i> and <i>Fusarium</i> sp.
<i>Phaseolus vulgaris</i> cv	Seed	<i>Valsa mali</i>
<i>Tinospora tomentosa</i>	Stem	<i>Aspergillus niger</i> and <i>Accharomyces cerevasecis</i>
<i>Allium sativum</i>	Leaf	<i>Rhizoctonia solani</i>
<i>Phaseolus lunatus</i> billb	Seed	<i>Sclerotium rolfsii</i> , <i>Fusarium solani</i> , <i>Fusarium oxysporum</i> , and <i>Botrytis cinerea</i>
<i>Calliandra surinamensis</i>	Leaf	<i>Candida krusei</i>
<i>Helianthus annuus</i> L.	Seed	<i>Sclerotinia sclerotiorum</i>
<i>Punica granatum</i>	Fruit	<i>Candida albicans</i> and <i>Candida kruse</i>
<i>Alpinia purpurata</i>	Inflorescence	<i>Candida albicans</i> and <i>Candida parapsilosis</i>
<i>Cicer arietinum</i>	Seed	<i>Candida parapsilosis</i> , <i>Candida krusei</i> , and <i>Candida tropicalis</i>

Insecticidal Activity

Hippeastrum hybrid (*Amaryllis*) ornamental plant bulb lectin showed insecticidal activity against the larvae of the cotton leaf worm (*Spodoptera littoralis*) (Caccia et al., 2012). Tubers of *Colocasia esculenta* having lectin which is toxic against *Aphis gossypii* (Cowpea aphid), *Dysdercus cingulatus* (Red cotton bug) and *Aphis craccivora* (Cowpea aphid) with the lectin concentration of 9.98 ± 0.239 µg/mL, 16.95 ± 0.279 µg/mL and 15.21 ± 0.274 µg/mL respectively (Das et al., 2013). *Microgramma vacciniifolia* rhizome lectin showed toxicity against *Nasutitermes corniger* (termite) workers and soldiers and acts as termiticidal lectin (Albuquerque et al., 2014). The lectin of *Myracrodruon urundeuva* and its leaf extract showed insecticidal properties against *Sitophilus zeamais* (maize weevil) which is a major pest of maize (Camaroti et al., 2013). The effects of *Dioclea violacea* lectin (DVL) on larval development in flour moth or mill moth (*Anagasta kuehniella*). DVL interfered with larval growth, retarding development and decreasing larval mass without affecting survival (Oliveira et al., 2015). Lectin with insecticidal property against the stored product pest, *Callosobruchus maculatus* was successfully isolated from the seeds of *Canavalia virosa* (Shanmugavel et al., 2016). *Moringa oleifera* seed cake lectin exerted deleterious effects on larvae (LC50: 0.89 mg/mL) and eggs (EC50: 0.14 mg/mL) of *Aedes aegypti* (Yellow fever mosquito) and served as an oviposition-stimulant at the concentration of 0.1 mg/mL (De Oliveira et al., 2016). The chitin-binding lectin from *Moringa oleifera* seeds showed insecticidal activity against *Anagasta kuehniella* (flour moth) (Fernando et al., 2017). Later the same chitin-binding lectin from *Moringa oleifera* seeds were investigated on gamma irradiation a low dose of gamma irradiation (10 mGy) can be used to improve the deleterious effects of moringa seed lectin on *Aedes aegypti* larvae and eggs (Santos et al., 2018). Lectin from *Abelmoschus esculentus* seeds Kunitz-type serine/cysteine protease inhibitor reported its toxic effects on the Mediterranean fruit fly *Ceratitis capitata* (De Lacerda et al., 2017). Lectin extracted from *Polygonum persicaria* L. is able to decrease damages of lepidopterous pests mainly *H. armigera* and *P. brassicae* on agricultural field (Rahimi et al., 2018). The lectin isolated from the leaves of *Schinus terebinthifolius* showed insecticidal activity against maize weevil (*Sitophilus zeamais*). The lectin doses of 100, 200, and 250 mg/g showed mortality rates of 32%, 40%, and 51% respectively on 7th day of treatments. On 12th day, the percentage of dead insects were increased to 94% to 97%. On 20th day of treatment all the insects were died. Whereas in control groups all the insects were survived even on 34th day (Camaroti et al., 2018) (Table 6).

Table 6 Insecticidal activity of plant lectins

Plants	Source	Against insects
<i>Hippeastrum</i> hybrid	Bulb	Cotton leaf worm
<i>Colocasia esculenta</i>	Tuber	Cowpea aphid and Red cotton bug
<i>Microgramma vacciniifolia</i>	Rhizome	Termite
<i>Myracrodruon urundeuva</i>	Leaf	Maize weevil
<i>Dioclea violacea</i>	Seed	Flour moth
<i>Canavalia virosa</i>	Seed	Cowpea weevil
<i>Moringa oleifera</i>	Seed	Yellow fever mosquito and flour moth
<i>Abelmoschus esculentus</i>	Seed	Mediterranean fruit fly
<i>Polygonum persicaria</i> L.	Fruit	Cotton bollworm
<i>Schinus terebinthifolius</i>	Leaf	Maize weevil

Anti-HIV Activity

Dioscorea bulbifera bulbils lectin showed anti-HIV activity and inhibits the HIV-1 reverse transcriptase activity in a dose dependent manner with IC50 of 1.3 µg (Sharma et al., 2017). *Sauromatum guttatum* (*Sauromatum venosum*) plant tuber lectinglycan array analysis and in-silico studies of the coding sequence proposed biological property as a putative anti-HIV agent (Thakur et al., 2017).

Antileishmanial Activity

Lectin isolated from *Abrus fruticulosus* seeds that has type 2 ribosome-inactivating properties showed antileishmanial activity against the promastigote form of *Leishmania* (Vasconcelos et al., 2018).

Nematicidal Activity

The lectin from white kidney beans (*Phaseolus vulgaris*) showed nematicidal effect against *Teladorsagia circumcincta* and *Trichostrongylus colubriformis* which are the parasites of sheep and goats (Ríos-de Álvarez et al., 2012). Seeds of *Dioclea lasiophylla* having lectin which shows less toxicity against *Artemia* sp. nauplii (Pinto et al., 2013). Lectin isolated from *Moringa oleifera* seed was evaluated for the inhibition of nematodes *Strongyloides* genera, *Oesophagostomum*, *Haemonchus* and *Trichostrongylus* in vitro effect of WSMoL (water soluble *Moringa oleifera* lectin) on hatching of eggs and the development of early-stage larvae of gastrointestinal nematodes from naturally infected goats (De Medeiros et al., 2018). Lectin from *Abelmoschus esculentus* seeds Kunitz-type serine/cysteine protease inhibitor reported its toxic effects on the root-knot nematodes *Meloidogyne incognita* and *Meloidogyne javanica* (De Lacerda et al., 2017). Eutirucallin: lectin isolated from latex of *Euphorbia tirucalli* was shown to be effective when tested directly against *Toxoplasma gondii* infection in vitro (Palharini et al., 2017). *Canavalia brasiliensis* lectin inhibits *Haemonchus contortus* also known as the barber's pole worm, in vitro, and through the *in silico* studies suggested that the inhibition of development is directly related to the recognition of the core trimannoside present in the N-glycans of these parasites (Batista et al., 2018).

Lectin application in the therapeutic aspect

Anticancerous Activity

Canavalia ensiformis (ConA) and *Canavalia brasiliensis* (ConBr) lectins showed antiproliferative activity with the IC50 values of approximately 3 µg/mL and 20 µg/mL using human leukemia MOLT-4 and HL-60 after 72 hrs of incubation with the lectins. The lectin was not toxic against normal human peripheral blood lymphocytes even with 200 µg/mL concentration of lectins (Faheina-Martins et al., 2012). Hog plum (*Ximena Americana*) fruit kernels is having a lectin known as Riproximin is specifically bind to two types of glycostructures, the N-linked NA2/NA3 [Galβ1-4GlcNAcβ1-2Manα1-6[Galβ1-4GlcNAcβ1-2Manα1-3]Manβ1-4GlcNAcBSA/Galβ1-4GlcNAcβ1-2Manα1-6[Galβ1-4GlcNAcβ1-2(Galβ1-4GlcNAcβ1-2)Manα1-3]Manβ1-4GlcNAc -BSA } and the O-linked clustered Tn tumor-specific antigen which found in tumor cells (Bayer et al., 2012). Lectin isolated and characterized from mulberry leaf induced apoptosis in human breast cancer cell line (MCF-7) and colon cancer cell line (HCT-15). Lectin required with the concentration of 8.5 µg/mL for MCF-7 and 16 µg/mL for HCT-15 to get 50% inhibition of cell growth after 24 hrs induced with lectin ((Deepa and Priya, 2012; Deepa et al., 2012). Lectin isolated from *Bauhinia unguolata* L. seeds showed in vitro antiproliferative activity against the HT-29 cell line of human colon adenocarcinoma in a dose-dependent manner, which require 160 µg/mL concentration of lectin to inhibit 80% of cell growth after 24 hrs of incubation (Silva et al., 2014). Lectin from pods of *Lotus corniculatus* showed antiproliferative activity towards human leukemic (THP-1) cancer cells and lung cancer (HOP62) cells and colon cancer cells (HCT116) with an IC50 of 39 µg/mL and 50 µg/mL and 60 µg/mL respectively after 24 hrs of lectin dose (Rafiq et al., 2013). Pea (*Pisum sativum* L.) seeds lectin was able to inhibit 84% of Ehrlich ascites carcinoma (EAC) cells after 24 hrs of incubation with the concentration of 120 µg/mL of pea lectin (Kabir et al., 2013). The seeds of *Phaseolus vulgaris* cv. Chinese pinto bean showed anti proliferative activity against nasopharyngeal carcinoma HONE-1 cells with an IC50 of 1mg/mL after 24 hrs of incubation with lectin (Ang et al., 2014). Monocot mannose-binding lectin from tubers of *Pinellia ternata* showed anti-proliferative effects on human liver tumor cell line Bel-7404 (Zhou et al., 2014). Lectin from the bulbs of *Allium chinense* inhibited 50% of the

human hepatoma Hep-3B cells after 48 hrs with the doses of 60 µg/mL of allium lectin (Xiao et al., 2015). Lectin extracted from the seeds of *Canavalia brasiliensis* (ConBr) as a therapeutic agent for melanoma because it inhibit the growth of murine melanoma B16F10 cells (Silva et al., 2014). *Bauhinia forficata* seeds lectin was treated against the breast cancer cells MCF-7, MDA-MB-231, MDA-MB-468 and MCF-10A among of all these cells *Bauhinia forficata* lectin was effective against MCF-7 cells (Silva et al., 2014). Antiproliferative lectin from *Cicer arietinum* seeds was toxic against human oral carcinoma cells (KB) at the concentration of 37.5 µg/mL (IC50) after 48 hrs of incubation and at the same time it was not toxic for normal human peripheral blood mononuclear cells (PBMCs) (Kumar et al., 2014). Toxic effects of *Microgramma vacciniifolia* rhizome lectin on lung mucoepidermoid carcinoma (NCI-H292) cells at IC50 concentration of 25.23 µg/mL, It did not affect the viability of chronic myelocytic leukaemia K562 and larynx epidermoid carcinoma Hep-2 tumour cells, as well as PBMCs normal cells (Albuquerque et al., 2014). Mannose-binding *Polygonatum odoratum* lectin showed anti-proliferative effect on MCF-7 cell growth with IC50 concentration of 10 µg/mL after 24 hrs of inhibition (Ouyang et al., 2014). Lectin isolated from soybean seed showed invitro antitumor effect on HeLa (cervical cancer), Hep2 (oral cancer), HepG2 (liver cancer), MDAMB 231(breast cancer), U373 MG (glioblastoma) with the IC50 concentration of 60, 20, 40, 40, 51 µg/mL, respectively and it did not affect the HaCaT (human keratinocyte). It also showed invitro antitumor effect on Dalton's lymphoma (DL) bearing mice; There was 74% and 82% reduction in cancer cell survival on the 10th day for the 1 and 2 mg/kg body weight doses, respectively (Panda et al., 2014). *Amaranthus mantegazzianus* seed lectin and commercial *Amaranthus caudatus* lectin were capable of inhibiting UMR106 rat osteosarcoma-derived cell proliferation with an IC50 of 0.1 mg/mL and 0.08 mg/mL respectively after 24 hrs of incubation (Quiroga et al., 2015). *Typhonium flagelliforme* (Lodd.) blume having lectin in tuber reported as antiproliferation activity against human breast cancer cell MCF-7 with LC50 of 90.78 ppm and the same tuber was isolated from other region showing LC50 of 130.93 ppm (Biosci et al., 2015). Ribosome-inactivating protein; three types of lectin ETRI1, ETRI2 and ETRI3 from the latex of *Euphorbia trigona* miller showed cytotoxicity against tumoral cell lines HeLa (IC50-11nM, 12nM, 10nM respectively), A549 (IC50-32nM, 32nM, 38nM respectively), H116 (IC50-40nM, 18nM, 21nM respectively), HL-60 (IC50-75nM, 44nM, 44nM respectively), HT-29 (no inhibition) and non-tumoral cell line NIH-3T3 (IC50-875nM, 480nM, 480nM respectively) analyzed by flowcytometry (Villanueva et al., 2015). High-mannose-type glycans of *Maaekia Amurensis* lectin was purchased which is known to interact with the α2,3-sialic acid moiety expressed on the MCF-7 human breast cancer cell line. The researcher groups have deglycosylated the lectin by using PNGase F for the removal of the high-Man-type N-linked glycans which leaving the two paucimannosidic type glycans in place on deglycosylated lectin, structural analysis of both the lectin release the information that there was no difference in both the structure and both the lectin were active. They found that the presence of a single GlcNAc attached to Asn 61 is sufficient for the lectin activity (Kim et al., 2015). *Polygonatum odoratum* lectin induce apoptosis and autophagy in lung cancer A549 cells. They found that the lectin down-regulated the expression of micro RNA (miR-1290) and at the same time upregulated the expression of micro RNA (miR-15a-3p), which leads to the apoptosis and autophagy of A549 cells by ROS-p53 mediated pathway (Wu et al., 2016). White kidney bean (*Phaseolus vulgaris cv.*) seeds lectin showed anti-proliferative activity on HONE1 cells, HepG2 cells, MCF7 cells and WRL68 cells with IC50 values of 18.8 µM, 19.7 µM, 26.9 µM, and >80 µM respectively for a 48 hrs treatment with lectin (Chan et al., 2016). Brazilian lima bean variety (*Phaseolus lunatus L. var. cascavel*) seeds having lectin with anti-tumor activity against melanoma derived cells (A375) at doses of 100, 50 and 25µg/mL, reducing tumor cells after 48 hrs of incubation number of cells was reduced by 73, 41, and 7%, and after 72 hrs of incubation it was reduced by 83, 53, and 0%, respectively (Lacerda et al., 2016). Lectin from *Canavalia virosa* seeds having cytotoxic activity against rat glioma cells (C6). The researcher group found that 100 µg/mL of lectin can able to decrease the cell viability by 25% and 50% after 24 and 48 hrs of treatment with lectin, respectively (Osterne et al., 2017). Lectin from the latex of mulberry (*Morus indica*) showed toxicity against an epithelial cell line MDCK and a breast cancer cell line MCF-7 cell line at the concentration of 0.625 µM of lectin for 72 hrs of treatment the viability for both cell lines reached ~35% and at the concentration of 1.25 µM it reached 17% in case of MDCK cells and 26% in case of MCF-7 (Datta et al., 2016). Lectin from latex of *Lagenaria siceraria* showed invitro cell cytotoxicity against MCF-7, A549, Dalton's lymphoma ascites (DLA), and Ehrlich ascites carcinoma (EAC) with IC50 value of 17.4 µg/mL, 20.4 µg/mL, 12 µg/mL, and 19.5 µg/mL, after 48 hrs of lectin doses but in case of non-cancerous cells NIH-3T3 was not showed toxicity at 50 µg/mL. It also showed invitro antitumor activity in ascites tumor model, Six dose of lectin with the concentration of 25 mg/kg was administered to tumor bearing mice on every alternative days and the found that there were reduction in the size of tumor (Vigneshwaran et al., 2016). *Phaseolus lunatus* billb seed lectin strongly inhibit the proliferation of K562 leukemia cells with an IC50 of 13.7 µM, whereas HeLa and HepG2 cells were only weakly affected after 48 hrs of incubation with the lectin (Wu et al., 2016). *Calliandra surinamensis* leaf lectin was evaluated for cell cytotoxicity on the cancer cell lines K562 (chronic myelogenous leukemia) and T47D (breast cancer) and noncancerous cells human peripheral blood mononuclear (PBMCs) after the treatment of lectin for 72 hrs the

IC50 observed with the concentration of 67.04 ± 5.78 and 58.75 ± 2.5µg/mL for cancerous cells respectively. Lectin did not affect non-cancerous cells at concentrations lower than 100 µg/mL (Figueiredo et al., 2017). The concanavalin A (Con A) and jacalin lectins on colonic adenocarcinoma cells (SW480) evaluated for antiproliferation of cells with pre-incubated lectins with dairy propionibacteria (*Propionibacterium acidipropionici* CRL 1198) as well as post-incubated lectins with the *P. acidipropionici* CRL 1198. They found that there were decreased in the antiproliferation activity of SW480 cells after incubating lectins with *P. acidipropionici* CRL 1198 as compare to the lectins activity before incubating the lectins with *P. acidipropionici* CRL 1198 (Zarate, 2017). Lectin from *Dioscorea bulbifera* bulbils inhibit the cell growth of HT 29, SW 620 and HepG2 cells with IC50 of 110 µg, 9.8 µg, 40 µg respectively after 72 hrs of interaction with lectin (Sharma et al., 2017). The seeds of *Canavalia bonariensis* lectin showed antitumor activity against rat C6 glioma cells with the lectin concentration of 50 and 100 mg/mL decrease cell viability at 40% and 50%, respectively, after 24 hrs treatment where as in case of 48 hrs of treatment with lectin concentration of 30 and 100 mg/mL cell viability decreased upto 30% and 50% respectively (Cavada et al., 2018). Native as well as recombinant lectin of *Cratylia mollis* seed showed toxicity against human prostate adenocarcinoma (PC-3) cells after 24 hrs exposure to the lectin along with the Concanavalin A. They observed that there were more than 10% decreased in the cell viability at 100 µg/mL and 300 µg/mL of lectin concentration in case of both the native as well as recombinant *Cratylia mollis* seed lectin; IC50 were 29.91 µg/mL and 39.69 µg/mL for recombinant and native *Cratylia mollis* seed lectin respectively. But there were no significant decrease in the cell viability in case of Concanavalin A with the concentration of 30 to 300 µg/mL (Figueirôa et al., 2017). Lectins isolated from the seeds of *Canavalia brasiliensis*, *Canavalia maritima*, *Dioclea lasiocarpa* and *Dioclea sclerocarpa* showed anti-proliferative activity against Human ovarian carcinoma (A2780), human Caucasian lung carcinoma (A549), human prostate carcinoma (PC3) and human breast carcinoma (MCF-7), with IC50 values ranging from 52 to 529 nM. Out of the four different lectins, *Dioclea lasiocarpa* lectin showed highly active against A2780 with IC50 value of 52 ± 2 nM. Whereas with MCF-7 IC50 value was 275 nM (Gondim et al., 2017). Later on the same group of researcher worked on the same lectin (*Dioclea lasiocarpa* lectin) showed decrease in the cell viability of rat glioma cells (C6) but decrease in the cell viability starts at the low concentration 10 µg/mL both after 24 hrs and 48 hrs on incubation (Nascimento et al., 2017). Methyl-β-d-galactopyranoside specific lectin from *Kaempferia rotunda* showed 43.7% cell growth inhibition in vitro against EAC cells at 160 µg/mL at the same time it was not inhibited the growth of U87 cell line and in-vivo antitumor effect of the lectin was observed in EAC bearing Swiss albino mice at the doses of 3.0 and 6.0 mg/kg/day for five consecutive days and found that there were decrease in cell growth of EAC by 41 and 59% respectively (Ahmed et al., 2017). The crude lectin from the latex of *Euphorbia tirucalli* showed toxicity against non-tumor cell lines such as peritoneal macrophages, murine bone-marrow-derived macrophages and fibroblasts after 18 hrs of incubation but purified lectin did not showed toxicity against these non-tumor cells. Where as in case of tumor cell lines HeLa: Human cervical cancer cell line, PC-3: Human prostate cancer cell line, MDA-MB-231: Human breast cancer cell line, MCF-7: Human breast adenocarcinoma cell line the crude and purified lectin showed antiproliferative activity; at concentration of 25 and 50 µg/mL, HeLa and PC3 were more susceptible to the harmful effects of purified lectin. MDA-MB-231 and MCF-7 were more susceptible to the harmful effects of crude lectin. There was same effect even at higher concentration. Breast cancer cell lines (MDA-MD-231 and MCF-7) were more sensitive to crude extract (47.9 and 56.9% death rates) than to the purified lectin (25.5 and 32.5%) death rates. Meanwhile, purified lectin killed more PC3 cells (27.5% death rate) than crude extract (19.6% death rate). There was no difference in cytotoxicity in HeLa cells between crude extract and purified lectin. In-vivo antitumor activities of both the crude and purified lectin were evaluated on intraperitoneal administered 100 Ehrlich ascites carcinomas (EAC) on mice. They found that the mice treated with crude and purified lectin were survived and there were no significant difference between the survival rates (Palharini et al., 2017). In vitro cytotoxic effects of *Moringa oleifera* seeds lectin on (B16-F10) melanoma cells; concentration of lectin ranging from 1.5–16 µM was evaluated for the reduction of the cells. The IC50 value was found at 9.72 µM. They determine that the reduction of the number of cells is due to the activation of caspases 3, 8 and 9 (Luz et al., 2017). Lectin from the same *Moringa oleifera* seeds inhibit the growth of Ehrlich ascites carcinoma (EAC) cell in-vitro with a concentration of lectin ranging from 25–200 µg/mL; at 200 µg/mL and 12.5 µg/mL the inhibitory effect was 71.08% and 15.27% respectively. In-vivo antitumor effect of the *Moringa oleifera* lectin on Ehrlich ascites carcinoma (EAC) on mice was observed at a concentration of lectin dose of 2.0 mg/kg/day there were 25.38% of EAC cell growth inhibition but when the concentration was increased to 4.0 mg/kg/day, the growth inhibition was increased to 55% (Asaduzzaman et al., 2018). Lectin from the phloem exudates/sap of fruit of *Praecitrullus fistulosus* showed invitro antiproliferative effects on cell lines HT29 (colon cancer), HeLa (cervical cancer), MCF7 (breast cancer) and K562 (leukemia) with an IC50 value of 43.80 µg, 45.17 µg, 62.80 µg and 76.23 µg respectively. The lectin did not showed cytotoxicity against normal cell line, peripheral blood mononuclear cells (PBMCs) even at a higher concentration of 1 mg/mL. In-vivo anticancerous activity was observed on EAC model 10 mg/kg injection for every alternate day (3

doses on day 7, 9 & 11). When compare it to the control i.e., without injection of lectin; the mice after receiving the first dose itself on day 7 there were gradual decrease in the body weight. After receiving the 3rd dose mice reduced tumor cell proliferation up to 75% (Shivamadhur et al., 2017). High mannose N-glycan binding lectin from *Remusatia vivipara* tubers showed anticancerous activity against the human breast cancer cell lines MDA-MB-468, MCF-7 and MCF-10A. After 48 hrs of treatment of lectin; 67.07% and 34.23% of cell viability was decreased in MDA-MB-468 and MCF-7 cells at 10 µg/mL dose of lectin. Whereas at 100 µg/mL concentration the cell viability decrease was 80.8% and 50.6% respectively. In case of non-tumorigenic MCF-10A cells derived from human fibrocystic mammary tissue, lectin exerted only marginal effect with 18.2% decrease in cell viability at 40 µg/mL concentration of lectin (Sindhura et al., 2017). Lectin from *Dioclea lasiophylla* Mart. Ex Benth seeds showed toxicity against C6 glioma cells at a concentration of 50 and 100 µg/mL of lectin decreased cell viability after 24 hrs of incubation and at 30–100 µg/mL after 48 hrs incubation (Leal et al., 2018). *Canavalia ensiformis* (ConA) and *Dioclea violacea* (DVL) lectins showed antiglioma effect on rat glioma cells (C6). *Dioclea violacea* lectin at concentration of 30 µg/mL decreased cell viability around 30% after 24 hrs of incubation, whereas after 48 hrs of incubation there were 66% of decrease in cell viability. *Canavalia ensiformis* lectin at concentration of 30 µg/mL the decrease in the cell viability was observed only after 24 hrs of incubation, there were only 30% decrease in the cell viability after 48 hrs of incubation (Nascimento et al., 2018). *Pisum sativum* seeds lectin showed inhibition of 17, 35 and 63% in SW480 cells at concentration of 0.25, 0.5 and 1.0 mg/mL after 3 days of treatment. Similarly, 18, 36 and 62% cell growth inhibition were noted in SW48 cells at concentration of 0.25, 0.5 and 1.0 mg/mL (Islam et al., 2018). The purified lectin from soya bean seeds displayed minor inhibition with 30% cell growth inhibition of B16F1 melanoma cells at a concentration of 35 µg/mL (Roy et al., 2018).

Anti-inflammatory and Antinociceptive Activity

Lectin from *Abelmoschus esculentus* seeds showed activity in reducing zymosan-induced temporomandibular joint (TMJ) inflammatory hypernociception in rats along with the mechanism of action like reduced leukocyte influx along with MPO activity as well as lectin reduced TNF-α and IL-1β levels in temporomandibular joint inflammatory tissue and trigeminal ganglion, Which showed lectin exerts anti-inflammatory activity (Freitas et al., 2016). Later lectin from the same plant *Abelmoschus esculentus* significantly reduced formalin-induced TMJ inflammatory hypernociception and decreased Evans blue extravasation. It decreased TNF-α levels in the TMJ tissue, trigeminal ganglion, and subnucleus caudalis. The lectin did not show antinociceptive effects in the presence of naltrindole or nor-binaltorphimine (Alves et al., 2018). *Bauhinia monandra* leaf lectin showed anti-inflammatory activity of 64% on paw edema induced by carrageenan as well as it also exert antinociceptive activity on mice (Campos et al., 2016). Lectins from *Parkia platycephala* showed antinociceptive activity in the mouse model of acetic acid induced and also demonstrated anti-inflammatory effect causing inhibition of leukocyte migration induced by both direct and indirect chemo attractants (Umarao et al., 2016). *Crotalaria retusa* L. seeds lectin inhibited the mice paw edema and neutrophil migration into the peritoneal cavity induced by carrageenan (Aragão et al., 2017). Lectin from *Lonchocarpus campestris* seeds inhibited paw edema and hyperalgesia induced by carrageenan as well as it showed antinociceptive activity in the behavioural tests of Formalin and Writhing (Pires et al., 2018).

Anti-coagulant Activity

Lectin purified from *Bauhinia forficata* seeds showed anticoagulant and antiplatelet properties. Coagulation time was increases in presence of *Bauhinia forficata* lectin and this consequence is not due to inhibition of human plasma kallikrein or human factor Xa. Dose dependent manner this lectin inhibits ADP and epinephrine-induced platelet aggregation. It was observed that at 3.0 µM concentration this lectin completely inhibits the platelet aggregation (Silva et al., 2012). *Moringa oleifera* seed lectin showed anticoagulant protein on in-vitro blood coagulation parameters and at least on activated partial thromboplastin by more than 300 Second and prothrombin times at a concentration ranging from 3 to 60 µg/mL (Luz et al., 2013). *Phthirusa pyrifolia* leaves lectin showed anticoagulant activity. It was showed through coagulation times like activated partial thromboplastin time (APTT) and thrombin time (TT) extrinsic pathway (Costa et al., 2017).

Immune Response Activity

Lectin from banana was cloned and expressed after that it was injected in Balb/c (albino). This experiment showed increased IgG and IgM concentrations in dose-dependent manners (Dimitrijevic et al., 2012). In the year 2016 same group showed banana lectin could be able to increase the concentration of IgA and IgG production in a dose-dependent manner (Marinković et al., 2016). One more recombinant banana lectin binds to murine peritoneal macrophages and affects their functional aspects. Lectin raised the production of IL-10, TGFβ, IL-4 and enhancement of arginase activity with reduction of NO IL-12 formation

(Marinkovic et al., 2017). Again, another lectin from the fruit pulp of banana, when orally administered to the mice, they found that there were, no significant differences were found in the lymphocytes of mouse peripheral blood, although an increase of CD4+ and decrease of CD8+ T-cells were observed in the thymus. However, they found that there were decrease in IL-6 and IFNγ and increase of IL-10, IL-17A and TNFα production in the peripheral blood of mice (Sansone et al., 2016). B-chain of *Korean mistletoe* seeds lectin did not induce apoptosis in bone marrow-derived dendritic cells (BMDCs). Whereas the treatment of lectin on BMDCs increases the expressions of co-stimulatory molecules (CD40, CD80, CD86, and MHC II) and the secretions of cytokines (IL-1β, IL-6, IL-12p70, and TNF-α). The lectin induced naïve CD4+ T cell differentiation toward Th1 cells directly and indirectly (Kim et al., 2014). Lectin from the Onion (*Allium cepa*) plant bulb increase the production of NO (nitric oxide) up to 6–8-fold in macrophage cell line, RAW264.7 and rat peritoneal macrophages. It stimulated the production of cytokines (TNF-α, IL-12, IFN-γ and IL-2) and furthermore enhanced the proliferation of murine thymocytes (Prasanna et al., 2015). Another onion lectin showed immune-protective properties by promoting the development of lymphoid cells count (WBC and lymphocytes count) and immune response in cyclophosphamide induced immune suppressed Wistar rats. Furthermore, it also depressed proinflammatory molecules like COX-2 and NO, and expression levels of immune regulatory molecule TNF-α, and suppressed the Th2-type type cytokine, i.e., serum IL-10 (Kumar et al., 2016). Lectin from the inflorescence of *Alpinia purpurata* induced the release of cytokines belonging to Th1 (IFN-γ, TNF-α, and IL-6) and Th17 (IL-17A) profiles as well as of nitric oxide. It stimulated the production of IL-10 as well as activate both T CD8+ and CD4+ subsets of lymphocytes (Brito et al., 2017). *Microgramma vacciniifolia* fronds lectin induced immunomodulatory properties on human peripheral blood mononuclear cells (PBMCs) by increasing TNFα, IFNγ, IL-6, IL-10, and nitric oxide production. It also stimulates T lymphocytes from PBMCs to differentiate into CD8+ cells (Leite et al., 2017). Lectin from the leaf pinnulae of *Calliandra surinamensis* activates immune cells in BALB/c mice splenocytes. They found that there were changes in in cytosolic calcium concentration ([Ca²⁺]cyt), mitochondrial membrane potential (ΔΨm), and reactive oxygen species (ROS) levels associated with cell viability, proliferation, cytokine and nitric oxide production. It also promotes the production of IL-2 and TNF-α (Procópio et al., 2018).

Mitogenic Activity

In the case of Red Kidney Bean (*Phaseolus vulgaris* L.), wild-type lectin and irradiated (γ-radiation) form lectins showed the mitogenic effect on lymphocyte proliferation. Authors found that the lectin after irradiation at 1kGy showed a decrease in mitogenic activity approx 70%, Whereas at 10kGy showed complete mitogenic activity (Mallikarjunan et al., 2014).

Glycoconjugates Research

Anacardium occidentale bark lectin was Immobilization on CNBr-Activated (cynogen bromide) Sepharose CL-4B. Bound feutin to this column demonstrated that lectin matrix could be used for glycoconjugates research (Maciel et al., 2012). The use of matrices (Sepharose CL-4B) containing *Cratylia mollis* lectins used for isolation of glycoproteins from fetal bovine serum, human colostrum, hen egg white, and human blood plasma (Santos-Filho et al., 2013). Lectin isolated from *Platypodium elegans* seeds when immobilized on CNBr-activated Sepharose 4B and showed feutin binding properties in solution, demonstrated that this lectin active and capable of binding carbohydrates (Alencar et al., 2017). Lectin extracted from *Cratylia mollis* seeds was immobilized on the surface of titanium dioxide nanotubes and found that increase in osteoblast-like cell adhesion on the TNTs-LbL-Cramoll (titanium dioxide nanotubes-layer by layer- *Cratylia mollis* seeds lectin) system when compared to the bare TNTs surfaces (Oliveira et al., 2018).

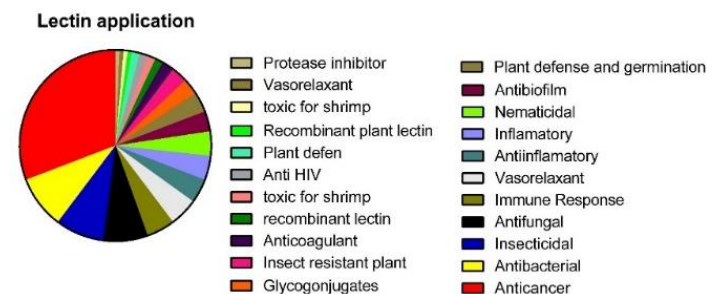


Figure 1 Application of lectins

CONCLUSION

Plant lectins are complex and carbohydrates binding proteins. The physiological role of lectins in the plant is not clear. Although plant lectins differ in their amino acids sequences, structural aspect as well evolutionary origin but most of lectins exhibits similar type of function such as antibacterial, antifungal, insecticidal, anti-

inflammatory (Figure 1). Due to easy availability and requirements simple purification system plants lectins will one of the best tools to study protein-carbohydrates interaction or useful disease biology.

Acknowledgement: Authors thank the Indian Council of Medical Research (ICMR) for research funding. S.K thanks SERB for financial assistance under the Fast-Track Scheme (SB/FT/LS-190/2012).

Conflict of interest: The authors declare that there is no conflict of interest.

REFERENCES

- Ahmed, F. R. S., Amin, R., Hasan, I., Asaduzzaman, A. K. M., & Kabir, S. R. (2017). Antitumor properties of a methyl- β -D-galactopyranoside specific lectin from *Kaempferia rotunda* against Ehrlich ascites carcinoma cells. *International Journal of Biological Macromolecules*, 102, 952–959. <https://doi.org/10.1016/j.ijbiomac.2017.04.109>
- Albuquerque, L. P. de, Pontual, E. V., Santana, G. M. de S., Silva, L. R. S., Aguiar, J. dos S., Coelho, L. C. B. B., Rgo, M. J. B. de M., Pitta, M. G. da R., Silva, T. G. da, Melo, A. M. M. de A., Napoleo, T. H., & Paiva, P. M. G. (2014). Toxic effects of *Microgramma vacciniifolia* rhizome lectin on *Artemia salina*, human cells, and the schistosomiasis vector *Biomphalaria glabrata*. *Acta Tropica*, 138, 23–27. <https://doi.org/10.1016/j.actatropica.2014.06.005>
- Alencar, D., Pinto-junior, V. R., Hadson, A., Neco, B., Queiroz, M., Jose, V., Osterne, S., Freitas, A., Figueiredo, C., Gleiciane, M., Martins, Q., Luiz, J., Correia, A., Guimaraes, R., Bairy, R., Maria, A., Assreuy, S., Santiago, K., & Sousa, B. (2017). Partial characterization and immobilization in CNBR-activated Sepharose of a native lectin from *Platypodium elegans* seeds (PELa) and comparative study of edematogenic effect with the recombinant form. *International Journal of Biological Macromolecules*, 102, 323–330. <https://doi.org/10.1016/j.ijbiomac.2017.03.193>
- Alves de Vasconcelos, M., Sena da Penha, S., Castro e Silva, V. R., Leite, T. A., Bezerra de Souza, E., Silva Souza, B. W., Teixeira, E. H., & Coelho da Silva, A. L. (2018). Fruticulosin: A novel type 2 ribosome-inactivating protein from *Abrus fruticulosus* seeds that exhibits toxic and antileishmanial activity. *Archives of Biochemistry and Biophysics*, 658, 46–53. <https://doi.org/10.1016/j.abb.2018.09.001>
- Alves, S. M., Freitas, R. S., do Val, D. R., Vieira, L. V., de Assis, E. L., Gomes, F. I. F., Gadelha, C. A. de A., Gadelha, T. S., de Lacerda, J. T. J. G., Clemente-Napimoga, J. T., Pinto, V. de P. T., Cristino Filho, G., Bezerra, M. M., & Chaves, H. V. (2018). The efficacy of a lectin from *Abelmoschus esculentus* depends on central opioid receptor activation to reduce temporomandibular joint hypernociception in rats. *Biomedicine and Pharmacotherapy*, 101, 478–484. <https://doi.org/10.1016/j.biopha.2018.02.117>
- Ang, A. S. W., Cheung, R. C. F., Dan, X., Chan, Y. S., Pan, W., & Ng, T. B. (2014). Purification and characterization of a glucosamine-binding antifungal lectin from *Phaseolus vulgaris* cv. Chinese Pinto Beans with antiproliferative activity towards nasopharyngeal carcinoma cells. *Applied Biochemistry and Biotechnology*, 172(2), 672–686. <https://doi.org/10.1007/s12010-013-0542-2>
- Aragão, D. P., da Silva Souza, B., de Brito, T. V., de Araújo Bastos Santana, L., de Paiva Silva, R. M., de Oliveira, A. P., da Cunha Pereira, A. C. T., Ferreira, G. P., dos Reis Barbosa, A. L., & de Oliveira, J. S. (2017). The anti-inflammatory and antinociceptive activity of albumins from *Crotalaria retusa* seeds. *Biomedicine and Pharmacotherapy*, 93, 536–542. <https://doi.org/10.1016/j.biopha.2017.06.078>
- Asaduzzaman, A. K. M., Hasan, I., Chakraborty, A., Zaman, S., Islam, S. S., Ahmed, F. R. S., Kabir, K. M. A., Nurujjaman, M., Uddin, M. B., Alam, M. T., Shaha, R. K., & Kabir, S. R. (2018). *Moringa oleifera* seed lectin inhibits Ehrlich ascites carcinoma cell growth by inducing apoptosis through the regulation of Bak and NF- κ B gene expression. *International Journal of Biological Macromolecules*, 107, 1936–1944. <https://doi.org/10.1016/j.ijbiomac.2017.10.070>
- Batista, J., Ralph, M., Vaz, R., Souza, P., Silva, A., Nascimento, D., Souza, L., Ramos, M., Mastroeni, P., & Lima-Filho, J. (2017). Plant lectins ConBr and CFL modulate expression toll-like receptors, pro-inflammatory cytokines and reduce the bacterial burden in macrophages infected with *Salmonella enterica* serovar Typhimurium. *Phytomedicine*, 25, 52–60. <https://doi.org/10.1016/j.phymed.2016.12.005>
- Batista, K. L. R., Silva, C. R., Santos, V. F., Silva, R. C., Roma, R. R., Santos, A. L. E., Pereira, R. O., Delatorre, P., Rocha, B. A. M., Soares, A. M. S., Costa-Júnior, L. M., & Teixeira, C. S. (2018). Structural analysis and anthelmintic activity of *Canavalia brasiliensis* lectin reveal molecular correlation between the carbohydrate recognition domain and glycans of *Haemonchus contortus*. *Molecular and Biochemical Parasitology*, 225, 67–72. <https://doi.org/10.1016/j.molbiopara.2018.09.002>
- Bayer, H., Essig, K., Stanzel, S., Frank, M., Gildersleeve, J. C., Berger, M. R., & Voss, C. (2012). Evaluation of riproximin binding properties reveals a novel mechanism for cellular targeting. *Journal of Biological Chemistry*, 287(43), 35873–35886. <https://doi.org/10.1074/jbc.M112.368548>
- Biosci, I. J., Alfarabi, M., Rosmalawati, S., & Bintang, M. (2015). Antiproliferative activity of tuber protein from *Typhonium flagelliforme* (Lodd.) blume on MCF-7 cell line. *International Journal of Biosciences*, 6, 52–60. <https://doi.org/10.12692/ijb/6.12.52-60>
- Bobbili, K. B., Pohlentz, G., Narahari, A., Sharma, K., Surolia, A., Mormann, M., & Swamy, M. J. (2018). *Coccinia indica* agglutinin, a 17 kDa PP2 like phloem lectin: Affinity purification, primary structure and formation of self-assembled filaments. *International Journal of Biological Macromolecules*, 108, 1227–1236. <https://doi.org/10.1016/j.ijbiomac.2017.11.024>
- Bose, P. P., Bhattacharjee, S., Singha, S., Mandal, S., Mondal, G., Gupta, P., & Chatterjee, B. P. (2016). A glucose/mannose binding lectin from litchi (*Litchi chinensis*) seeds: Biochemical and biophysical characterizations. *Biochemistry and Biophysics Reports*, 6, 242–252. <https://doi.org/10.1016/j.bbrep.2016.05.001>
- Caccia, S., Van Damme, E. J. M., De Vos, W. H., & Smagge, G. (2012). Mechanism of entomotoxicity of the plant lectin from *Hippeastrum hybrid* (Amaryllis) in *Spodoptera littoralis* larvae. *Journal of Insect Physiology*, 58(9), 1177–1183. <https://doi.org/10.1016/j.jinsphys.2012.05.014>
- Camaroti, J. R. S. L., de Almeida, W. A., do Rego Belmonte, B., de Oliveira, A. P. S., de Albuquerque Lima, T., Ferreira, M. R. A., Paiva, P. M. G., Soares, L. A. L., Pontual, E. V., & Napoleão, T. H. (2018). *Sitophilus zeamais* adults have survival and nutrition affected by *Schinus terebinthifolius* leaf extract and its lectin (SteLL). *Industrial Crops and Products*, 116, 81–89. <https://doi.org/10.1016/j.indcrop.2018.02.065>
- Campos, J. K. L., Arajo, C. S. F., Arajo, T. F. S., Santos, A. F. S., Teixeira, J. A., Lima, V. L. M., & Coelho, L. C. B. B. (2016). Anti-inflammatory and antinociceptive activities of *Bauhinia monandra* leaf lectin. *Biochimie*, 2, 62–68. <https://doi.org/10.1016/j.biopen.2016.03.001>
- Carvalho, A. de S., da Silva, M. V., Gomes, F. S., Paiva, P. M. G., Malafaia, C. B., da Silva, T. D., Vaz, A. F. de M., da Silva, A. G., Arruda, I. R. de S., Napoleo, T. H., Carneiro-da-Cunha, M. da G., & Correia, M. T. dos S. (2015). Purification, characterization and antibacterial potential of a lectin isolated from *Apuleia leiocarpa* seeds. *International Journal of Biological Macromolecules*, 75, 402–408. <https://doi.org/10.1016/j.ijbiomac.2015.02.001>
- Cavada, B. S., Silva, M. T. L., Osterne, V. J. S., Pinto-Junior, V. R., Nascimento, A. P. M., Wolin, I. A. V., Heinrich, I. A., Nobre, C. A. S., Moreira, C. G., Lossio, C. F., Rocha, C. R. C., Martins, J. L., Nascimento, K. S., & Leal, R. B. (2018). *Canavalia bonariensis* lectin: Molecular bases of glycoconjugates interaction and antiangioma potential. *International Journal of Biological Macromolecules*, 106, 369–378. <https://doi.org/10.1016/j.ijbiomac.2017.08.023>
- Chan, Y. S., Xia, L., & Ng, T. B. (2016). White kidney bean lectin exerts anti-proliferative and apoptotic effects on cancer cells. *International Journal of Biological Macromolecules*, 85, 335–345. <https://doi.org/10.1016/j.ijbiomac.2015.12.094>
- Costa, R. B., Campana, P. T., Chambergo, F. S., Napoleão, T. H., Paiva, P. M. G., Pereira, H. J. V., Oliva, M. L. V., & Gomes, F. S. (2018). Purification and characterization of a lectin with refolding ability from *Genipa americana* bark. *International Journal of Biological Macromolecules*, 119, 517–523. <https://doi.org/10.1016/j.ijbiomac.2018.07.178>
- Costa, R. M. P. B., Albuquerque, W. W. C., Silva, M. C. C., Paula, R. A. de, Melo, M. S., Oliva, M. L. V., & Porto, A. L. F. (2017). Can γ -radiation modulate hemagglutinating and anticoagulant activities of PpyLL, a lectin from *Phthirusa pyrifolia*. *International Journal of Biological Macromolecules*, 104, 125–136. <https://doi.org/10.1016/j.ijbiomac.2017.06.007>
- Da Silva, Pollyanna Michelle, de Moura, M. C., Gomes, F. S., da Silva Trentin, D., Silva de Oliveira, A. P., de Mello, G. S. V., da Rocha Pitta, M. G., de Melo Rego, M. J. B., Coelho, L. C. B. B., Macedo, A. J., de Figueiredo, R. C. B. Q., Paiva, P. M. G., & Napoleão, T. H. (2018). PgTeL, the lectin found in *Punica granatum* juice, is an antifungal agent against *Candida albicans* and *Candida krusei*. *International Journal of Biological Macromolecules*, 108, 391–400. <https://doi.org/10.1016/j.ijbiomac.2017.12.039>
- Das, A., Roy, A., Hess, D., & Das, S. (2013). Characterization of a Highly Potent Insecticidal Lectin from *Colocasia esculenta* Tuber and Cloning of Its Coding Sequence. *American Journal of Plant Sciences*, 4, 408–416. <https://doi.org/10.4236/ajps.2013.42A053>
- Datta, D., Pohlentz, G., Schulte, M., Kaiser, M., Goycoolea, F. M., Mthing, J., Mormann, M., & Swamy, M. J. (2016). Physico-chemical characteristics and primary structure of an affinity-purified α -D-galactose-specific, jacalin-related lectin from the latex of mulberry (*Morus indica*). *Archives of Biochemistry and Biophysics*, 609, 59–68. <https://doi.org/10.1016/j.abb.2016.09.009>
- De Andrade Luz, L., Rossato, F. A., Costa, R. A. P. e., Napoleão, T. H., Paiva, P. M. G., & Coelho, L. C. B. B. (2017). Cytotoxicity of the coagulant *Moringa oleifera* lectin (cMoL) to B16-F10 melanoma cells. *Toxicology in Vitro*, 44, 94–99. <https://doi.org/10.1016/j.tiv.2017.06.037>
- De Freitas Pires, A., Bezerra, M. M., Amorim, R. M. F., do Nascimento, F. L. F., Marinho, M. M., Moura, R. M., Silva, M. T. L., Correia, J. L. A., Cavada, B. S., Assreuy, A. M. S., & Nascimento, K. S. (2019). Lectin purified from *Lonchocarpus campestris* seeds inhibits inflammatory nociception. *International Journal of Biological Macromolecules*, 125, 53–60. <https://doi.org/10.1016/j.ijbiomac.2018.11.233>
- De Lacerda, J. T. J. G., e Lacerda, R. R., Assunção, N. A., Tashima, A. K., Juliano, M. A., dos Santos, G. A., dos Santos de Souza, M., de Luna Batista, J., Rossi, C.

- E., de Almeida Gadelha, C. A., & Santi-Gadelha, T. (2017). New insights into lectin from *Abelmoschus esculentus* seeds as a Kunitz-type inhibitor and its toxic effects on *Ceratitis capitata* and root-knot nematodes *Meloidogyne* spp. *Process Biochemistry*, 63, 96–104. <https://doi.org/10.1016/j.procbio.2017.09.003>
- De Medeiros, M. L. S., de Moura, M. C., Napoleão, T. H., Paiva, P. M. G., Coelho, L. C. B. B., Bezerra, A. C. D. S., & da Silva, M. D. C. (2018). Nematicidal activity of a water soluble lectin from seeds of *Moringa oleifera*. *International Journal of Biological Macromolecules*, 108, 782–789. <https://doi.org/10.1016/j.ijbiomac.2017.10.167>
- De Oliveira, A. P. S., de Santana Silva, L. L., de Albuquerque Lima, T., Pontual, E. V., de Lima Santos, N. D., Breitenbach Barroso Coelho, L. C., do Amaral Ferraz Navarro, D. M., Zingali, R. B., Napoleo, T. H., & Paiva, P. M. G. (2016). Biotechnological value of *Moringa oleifera* seed cake as source of insecticidal lectin against *Aedes aegypti*. *Process Biochemistry*, 51(10), 1683–1690. <https://doi.org/10.1016/j.procbio.2016.06.026>
- De Oliveira Figueiróa, E., Aranda-Souza, M. Â., Varejão, N., Rossato, F. A., Costa, R. A. P., Figueira, T. R., da Silva, L. C. N., Castilho, R. F., Vercesi, A. E., & dos Santos Correia, M. T. (2017). pCramoll and rCramoll lectins induce cell death in human prostate adenocarcinoma (PC-3) cells by impairment of mitochondrial homeostasis. *Toxicology in Vitro*, 43, 40–46. <https://doi.org/10.1016/j.tiv.2017.05.016>
- De Santana Brito, J., Ferreira, G. R. S., Klimczak, E., Gryshuk, L., de Lima Santos, N. D., de Siqueira Patriota, L. L., Moreira, L. R., Soares, A. K. A., Barboza, B. R., Paiva, P. M. G., do Amaral Ferraz Navarro, D. M., de Lorena, V. M. B., de Melo, C. M. L., Coriolano, M. C., & Napoleão, T. H. (2017). Lectin from inflorescences of ornamental crop *Alpinia purpurata* acts on immune cells to promote Th1 and Th17 responses, nitric oxide release, and lymphocyte activation. *Biomedicine and Pharmacotherapy*, 94, 865–872. <https://doi.org/10.1016/j.biopha.2017.08.026>
- Deepa, M., & Priya, S. (2012). Purification and Characterization of a Novel Anti-Proliferative Lectin from *Morus alba* L. Leaves. *Protein & Peptide Letters*, 19(8), 839–845. <https://doi.org/10.2174/092986612801619516>
- Deepa, M., Sureshkumar, T., Satheeshkumar, P. K., & Priya, S. (2012). Purified mulberry leaf lectin (MLL) induces apoptosis and cell cycle arrest in human breast cancer and colon cancer cells. *Chemico-Biological Interactions*, 200(1), 38–44. <https://doi.org/10.1016/j.cbi.2012.08.025>
- Del Rio, M., de la Canal, L., Pinedo, M., & Regente, M. (2018). Internalization of a sunflower mannose-binding lectin into phytopathogenic fungal cells induces cytotoxicity. *Journal of Plant Physiology*, 221, 22–31. <https://doi.org/10.1016/j.jplph.2017.12.001>
- Dimitrijevic, R., Stojanovic, M., Mivic, M., Dimitrijevic, L., & Gavrovic-Jankulovic, M. (2012). Recombinant banana lectin as mucosal immunostimulator. *Journal of Functional Foods*, 4(3), 636–641. <https://doi.org/10.1016/j.jff.2012.04.003>
- Faheina-Martins, G. V., da Silveira, A. L., Cavalcanti, B. C., Ramos, M. V., Moraes, M. O., Pessoa, C., & Araújo, D. A. M. (2012). Antiproliferative effects of lectins from *Canavalia ensiformis* and *Canavalia brasiliensis* in human leukemia cell lines. *Toxicology in Vitro*, 26(7), 1161–1169. <https://doi.org/10.1016/j.tiv.2012.06.017>
- Fernando, C., Oliveira, R. De, Celine, M., Moura, D., Henrique, T., Maria, P., Paiva, G., Cassandra, L., Barroso, B., Lígia, M., & Macedo, R. (2017). A chitin-binding lectin from *Moringa oleifera* seeds (WSMoL) impairs the digestive physiology of the Mediterranean flour larvae, *Anagasta kuehniella*. *Pesticide Biochemistry and Physiology*, 1–10. <https://doi.org/10.1016/j.pestbp.2017.01.006>
- Ferreira, G. R. S., Brito, J. de S., Procópio, T. F., Santos, N. D. de L., de Lima, B. J. R. C., Coelho, L. C. B. B., Navarro, D. M. do A. F., Paiva, P. M. G., Soares, T., de Moura, M. C., & Napoleão, T. H. (2018). Antimicrobial potential of *Alpinia purpurata* lectin (ApuL): Growth inhibitory action, synergistic effects in combination with antibiotics, and antibiofilm activity. *Microbial Pathogenesis*, 124, 152–162. <https://doi.org/10.1016/j.micpath.2018.08.027>
- Figueiredo, T., Leite, L., Patriota, D. S., Celine, M., Moura, D., Michelle, P., Patrícia, A., Oliveira, S. De, Soares, T., Diego, T., Cassandra, L., Barroso, B., Galdino, M., Jesus, M., Melo, B. De, Celia, R., Queiroz, B., Figueiredo, D., Maria, P., Henrique, T. (2017). CasuL: A new lectin isolated from *Calliandra surinamensis* leaf pinnulae with cytotoxicity to cancer cells , antimicrobial activity and antibiofilm effect. *International Journal of Biological Macromolecules*, 98, 419–429. <https://doi.org/10.1016/j.ijbiomac.2017.02.019>
- Freitas, R. S., do Val, D. R., Fernandes, M. E. F., Gomes, F. I. F., de Lacerda, J. T. J. G., SantiGadelha, T., de Almeida Gadelha, C. A., de Paulo Teixeira Pinto, V., Cristino-Filho, G., Pereira, K. M. A., de Castro Brito, G. A., Bezerra, M. M., & Chaves, H. V. (2016). Lectin from *Abelmoschus esculentus* reduces zymosan-induced temporomandibular joint inflammatory hypernociception in rats via heme oxygenase-1 pathway integrity and tnf- α and il-1 β suppression. *International Immunopharmacology*, 38, 313–323. <https://doi.org/10.1016/j.intimp.2016.06.012>
- Geethanandan, K., Joseph, A., Sadasivan, C., & Haridas, M. (2013). Carbohydrate binding and unfolding of spatholobus parviflorus lectin: Fluorescence and circular dichroism spectroscopic study. *Applied Biochemistry and Biotechnology*, 171(1), 80–92. <https://doi.org/10.1007/s12010-013-0340-x>
- Gemeiner, P., Mislovičová, D., Tkáč, J., Švitel, J., Pätoprstý, V., Hrabárová, E., Kogan, G., & Kožár, T. (2009). Lectinomics. II. A highway to biomedical/clinical diagnostics. *Biotechnology Advances*, 27(1), 1–15. <https://doi.org/10.1016/j.biotechadv.2008.07.003>
- Ghosh, P., Sen, S., Chakraborty, J., & Das, S. (2016). Monitoring the efficacy of mutated *Allium sativum* leaf lectin in transgenic rice against *Rhizoctonia solani*. *BMC Biotechnology*, 16(1), 24. <https://doi.org/10.1186/s12896-016-0246-0>
- Gondim, A. C. S., Romero-Canelón, I., Sousa, E. H. S., Blindauer, C. A., Butler, J. S., Romero, M. J., Sanchez-Cano, C., Sousa, B. L., Chaves, R. P., Nagano, C. S., Cavada, B. S., & Sadler, P. J. (2017). The potent anti-cancer activity of *Dioclea lasiocarpa* lectin. *Journal of Inorganic Biochemistry*, 175, 179–189. <https://doi.org/10.1016/j.jinorgbio.2017.07.011>
- Hou, F. J., Xu, H., & Liu, W. Y. (2003). Simultaneous existence of cinnamomin (a type II RIP) and small amount of its free A- and B-chain in mature seeds of camphor tree. *International Journal of Biochemistry and Cell Biology*, 35(4), 455–464. [https://doi.org/10.1016/S1357-2725\(02\)00274-1](https://doi.org/10.1016/S1357-2725(02)00274-1)
- Islam, F., Gopalan, V., Lam, A. K. Y., & Kabir, S. R. (2018). Pea lectin inhibits cell growth by inducing apoptosis in SW480 and SW48 cell lines. *International Journal of Biological Macromolecules*, 117(2017), 1050–1057. <https://doi.org/10.1016/j.ijbiomac.2018.06.021>
- Jiménez, P., Gayoso, M., Tejero, J., Cabrero, P., Córdoba-Díaz, D., Basterrechea, J. E., & Gírbés, T. (2013). Toxicity in mice of lectin ebulin f present in dwarf Elderberry (*Sambucus ebulus* L.). *Toxicol*, 61(1), 26–29. <https://doi.org/10.1016/j.toxicol.2012.10.009>
- Kabir, S. R., Nabi, M. M., Haque, A., Zaman, R. U., Mahmud, Z. H., & Reza, M. A. (2013). Pea lectin inhibits growth of *Ehrlich ascites* carcinoma cells by inducing apoptosis and G2/M cell cycle arrest in vivo in mice. *Phytomedicine*, 20(14), 1288–1296. <https://doi.org/10.1016/j.phymed.2013.06.010>
- Kim, B. S., Hwang, H. S., Park, H., & Kim, H. H. (2015). Effects of selective cleavage of high-mannose-type glycans of *Maackia amurensis* leucoagglutinin on sialic acid-binding activity. *Biochimica et Biophysica Acta - General Subjects*, 1850(9), 1815–1821. <https://doi.org/10.1016/j.bbagen.2015.05.011>
- Kim, J. J., Hwang, Y. H., Kang, K. Y., Kim, I., Kim, J. B., Park, J. H., Yoo, Y. C., & Yee, S. T. (2014). Enhanced dendritic cell maturation by the B-chain of Korean mistletoe lectin (KML-B), a novel TLR4 agonist. *International Immunopharmacology*, 21(2), 309–319. <https://doi.org/10.1016/j.intimp.2014.05.010>
- Klafke, G. B., Moreira, G. M. S. G., Pereira, J. L., Oliveira, P. D., Conceição, F. R., Lund, R. G., Grassmann, A. A., Dellagostin, O. A., & da Silva Pinto, L. (2016). Lectin I from *Bauhinia variegata* (BVL-I) expressed by *Pichia pastoris* inhibits initial adhesion of oral bacteria in vitro. *International Journal of Biological Macromolecules*, 93, 913–918. <https://doi.org/10.1016/j.ijbiomac.2016.09.062>
- Kumar, S., Kapoor, V., Gill, K., Singh, K., Xess, I., Das, S. N., & Dey, S. (2014). Antifungal and antiproliferative protein from cicer arietinum: A bioactive compound against emerging pathogens. *BioMed Research International*, 2014, 1–9. <https://doi.org/10.1155/2014/387203>
- Kumar, V. P., & Venkatesh, Y. P. (2016). Alleviation of cyclophosphamide-induced immunosuppression in Wistar rats by onion lectin (*Allium cepa* agglutinin). *Journal of Ethnopharmacology*, 186, 280–288. <https://doi.org/10.1016/j.jep.2016.04.006>
- Leal, R. B., Pinto-Junior, V. R., Osterne, V. J. S., Wolin, I. A. V., Nascimento, A. P. M., Neco, A. H. B., Araripe, D. A., Welter, P. G., Neto, C. C., Correia, J. L. A., Rocha, C. R. C., Nascimento, K. S., & Cavada, B. S. (2018). Crystal structure of DlyL, a mannose-specific lectin from *Dioclea lasiophylla* Mart. Ex Benth seeds that display cytotoxic effects against C6 glioma cells. *International Journal of Biological Macromolecules*, 114, 64–76. <https://doi.org/10.1016/j.ijbiomac.2018.03.080>
- Leite, L., Patriota, D. S., Figueiredo, T., Santana, J. De, Sebag, V., Patrícia, A., Oliveira, S. De, Karine, A., Soares, D. A., Rafael, L., Albuquerque, T. De, Soares, T., Diego, T., Maria, P., Paiva, G., Maria, V., Lorena, B. De, Lagos, M., Melo, D., ... Henrique, T. (2017). *Microgramma vacciniifolia* (Polypodiaceae) fronds contain a multifunctional lectin with immunomodulatory properties on human cells. *International Journal of Biological Macromolecules*, 103, 36–46. <https://doi.org/10.1016/j.ijbiomac.2017.05.037>
- Luz, L. de A., Silva, M. C. C., Ferreira, R. da S., Santana, L. A., Silva-Lucca, R. A., Mentele, R., Oliva, M. L. V., Paiva, P. M. G., & Coelho, L. C. B. B. (2013). Structural characterization of coagulant *Moringa oleifera* Lectin and its effect on hemostatic parameters. *International Journal of Biological Macromolecules*, 58, 31–38. <https://doi.org/10.1016/j.ijbiomac.2013.03.044>
- Ma, Q. H., Zhen, W. B., & Liu, Y. C. (2013). Jacalin domain in wheat jasmonate-regulated protein Ta-JA1 confers agglutinating activity and pathogen resistance. *Biochimie*, 95(2), 359–365. <https://doi.org/10.1016/j.biochi.2012.10.014>
- Maciel, M. I. S., De Mendonça Cavalcanti, M. D. S., Napoleão, T. H., Paiva, P. M. G., De Almeida Catanho, M. T. J., & Coelho, L. C. B. B. (2012). *Anacardium occidentale* bark lectin: Purification, immobilization as an affinity model and influence in the uptake of technetium-99M by rat adipocytes. *Applied Biochemistry and Biotechnology*, 168(3), 580–591. <https://doi.org/10.1007/s12010-012-9798-1>
- Mallikarjunan, N., Marathe, S., Rajalakshmi, D., Mahesh, S., Jamdar, S. N., & Sharma, A. (2014). Effect of ionizing radiation on structural and functional attributes of red kidney bean (*Phaseolus vulgaris* L.) lectin. *LWT - Food Science and Technology*, 59(1), 300–307. <https://doi.org/10.1016/j.lwt.2014.04.052>
- Marinkovic, E., Djokic, R., Lukic, I., Filipovic, A., Inic-Kanada, A., Kosanovic,

- D., Gavrovic-Jankulovic, M., & Stojanovic, M. (2017). Modulation of functional characteristics of resident and thioglycollate-elicited peritoneal murine macrophages by a recombinant banana lectin. *PLoS ONE*, 12(2), 1–21. <https://doi.org/10.1371/journal.pone.0172469>
- Marinković, E., Lukić, I., Kosanović, D., Inić-Kanada, A., Gavrovic-Jankulović, M., & Stojanović, M. (2016). Recombinantly produced banana lectin isoform promotes balanced pro-inflammatory response in the colon. *Journal of Functional Foods*, 20, 68–78. <https://doi.org/10.1016/j.jff.2015.10.019>
- Naik, S., & Kumar, S. (2020). Biochemical Characterization of lactose binding entadin lectin from *Entada rheedii* seeds with cytotoxic activity against cancer cell lines. *ACS Omega*, 5(27), 16430–16439. <https://doi.org/10.1021/acsomega.0c00577>
- Naik, S., Rawat, R. S., Khandai, S., Kumar, M., Jena, S. S., Vijayalakshmi, M. A., & Kumar, S. (2017). Biochemical characterisation of lectin from Indian hyacinth plant bulbs with potential inhibitory action against human cancer cells. *International Journal of Biological Macromolecules*, 105, 1349–1356. <https://doi.org/10.1016/j.ijbiomac.2017.07.170>
- Napoleão, T. H., Belmonte, B. do R., Pontual, E. V., de Albuquerque, L. P., Sá, R. A., Paiva, L. M., Breitenbach Barroso Coelho, L. C., & Paiva, P. M. G. (2013). Deleterious effects of *Myracrodruon urundeuva* leaf extract and lectin on the maize weevil, *Sitophilus zeamais* (Coleoptera, Curculionidae). *Journal of Stored Products Research*, 54, 26–33. <https://doi.org/10.1016/j.jspr.2013.04.002>
- Napoleão, T. H., Gomes Dos Santos-Filho, T., Pontual, E. V., Da Silva Ferreira, R., Coelho, L. C. B. B., & Paiva, P. M. G. (2013). Affinity matrices of *Cratylia mollis* seed lectins for isolation of glycoproteins from complex protein mixtures. *Applied Biochemistry and Biotechnology*, 171(3), 744–755. <https://doi.org/10.1007/s12010-013-0403-z>
- Nascimento, A. P. M., Knaut, J. L., Rieger, D. K., Wolin, I. A. V., Heinrich, I. A., Mann, J., Juarez, A. V., Sosa, L. del V., De Paul, A. L., Moreira, C. G., Silva, I. B., Nobre, C. S., Osterne, V. J. S., Nascimento, K. S., Cavada, B. S., & Leal, R. B. (2018). Anti-glioma properties of DVL, a lectin purified from *Dioclea violacea*. *International Journal of Biological Macromolecules*, 120, 566–577. <https://doi.org/10.1016/j.ijbiomac.2018.08.106>
- Nascimento, K. S., Santiago, M. Q., Pinto-Junior, V. R., Osterne, V. J. S., Martins, F. W. V., Nascimento, A. P. M., Wolin, I. A. V., Heinrich, I. A., Martins, M. G. Q., Silva, M. T. L., Lossio, C. F., Rocha, C. R. C., Leal, R. B., & Cavada, B. S. (2017). Structural analysis of *Dioclea lasiocarpa* lectin: A C6 cells apoptosis-inducing protein. *International Journal of Biochemistry and Cell Biology*, 92, 79–89. <https://doi.org/10.1016/j.biocel.2017.09.014>
- Oliveira, C. T., Kunz, D., Silva, C. P., & Macedo, M. L. R. (2015). Entomotoxic properties of *Dioclea violacea* lectin and its effects on digestive enzymes of *Anagasta kuehniella* (Lepidoptera). *Journal of Insect Physiology*, 81, 81–89. <https://doi.org/10.1016/j.jinsphys.2015.07.007>
- Oliveira, W. F., Silva, G. M. M., Cabral Filho, P. E., Fontes, A., Oliveira, M. D. L., Andrade, C. A. S., Silva, M. V., Coelho, L. C. B. B., Machado, G., & Correia, M. T. S. (2018). Titanium dioxide nanotubes functionalized with *Cratylia mollis* seed lectin, Cramoll, enhanced osteoblast-like cells adhesion and proliferation. *Materials Science and Engineering C*, 90, 664–672. <https://doi.org/10.1016/j.msec.2018.04.089>
- Osterne, V. J. S., Silva-Filho, J. C., Santiago, M. Q., Pinto-Junior, V. R., Almeida, A. C., Barreto, A. A. G. C., Wolin, I. A. V., Nascimento, A. P. M., Amorim, R. M. F., Rocha, B. A. M., Delatorre, P., Nagano, C. S., Leal, R. B., Assreuy, A. M. S., Nascimento, K. S., & Cavada, B. S. (2017). Structural characterization of a lectin from *Canavalia virosa* seeds with inflammatory and cytotoxic activities. *International Journal of Biological Macromolecules*, 94, 271–282. <https://doi.org/10.1016/j.ijbiomac.2016.10.020>
- Ouyang, L., Chen, Y., Wang, X. Y., Lu, R. F., Zhang, S. Y., Tian, M., Xie, T., Liu, B., & He, G. (2014). *Polygonatum odoratum* lectin induces apoptosis and autophagy via targeting EGFR-mediated Ras-Raf-MEK-ERK pathway in human MCF-7 breast cancer cells. *Phytomedicine*, 21(12), 1658–1665. <https://doi.org/10.1016/j.phymed.2014.08.002>
- Palharini, J. G., Richter, A. C., Silva, M. F., Ferreira, F. B., Pirovani, C. P., Naves, K. S. C., Goulart, V. A., Mineo, T. W. P., Silva, M. J. B., & Santiago, F. M. (2017). Eutirucallin: A lectin with antitumor and antimicrobial properties. *Frontiers in Cellular and Infection Microbiology*, 7, 1–13. <https://doi.org/10.3389/fcimb.2017.00136>
- Panda, P. K., Mukhopadhyay, S., Behera, B., Bhol, C. S., Dey, S., Das, D. N., Sinha, N., Bissoyi, A., Pramanik, K., Maiti, T. K., & Bhatia, S. K. (2014). Antitumor effect of soybean lectin mediated through reactive oxygen species-dependent pathway. *Life Sciences*, 111(1), 27–35. <https://doi.org/10.1016/j.lfs.2014.07.004>
- Pereira de Albuquerque, L., Maria de Sá Santana, G., Pontual, E. V., Napoleão, T. H., Breitenbach Barroso Coelho, L. C., & Paiva, P. M. G. (2012). Effect of *Microgramma vacciniifolia* rhizome lectin on survival and digestive enzymes of *Nasutitermes corniger* (Isoptera, Termitidae). *International Biodeterioration and Biodegradation*, 75, 158–166. <https://doi.org/10.1016/j.ibiod.2012.06.030>
- Pinto, V. R., De Santiago, M. Q., Da Silva Osterne, V. J., Correia, J. L. A., Pereira, F. N., Cajazeiras, J. B., De Vasconcelos, M. A., Teixeira, E. H., Do Nascimento, A. S. F., Miguel, T. B. A. R., De Castro Miguel, E., Sampaio, A. H., Do Nascimento, K. S., Nagano, C. S., & Cavada, B. S. (2013). Purification, partial characterization and immobilization of a mannose-specific lectin from seeds of *Dioclea lasiophylla* mart. *Molecules*, 18(9), 10857–10869. <https://doi.org/10.3390/molecules180910857>
- Polito, L., Bortolotti, M., Battelli, M. G., Calafato, G., & Bolognesi, A. (2019). Ricin: An ancient story for a timeless plant toxin. *Toxins*, 11(6), 1–16. <https://doi.org/10.3390/toxins11060324>
- Prasanna, V. K., & Venkatesh, Y. P. (2015). Characterization of onion lectin (*Allium cepa* agglutinin) as an immunomodulatory protein inducing Th1-type immune response in vitro. *International Immunopharmacology*, 26(2), 304–313. <https://doi.org/10.1016/j.intimp.2015.04.009>
- Procópio, T. F., de Siqueira Patriota, L. L., da Silva Barros, B. R., de Souza Aguiar, L. M., de Lorena, V. M. B., Paiva, P. M. G., de Melo, C. M. L., & Napoleão, T. H. (2018). *Calliandra surinamensis* lectin (CasuL) does not impair the functionality of mice splenocytes, promoting cell signaling and cytokine production. *Biomedicine and Pharmacotherapy*, 107, 650–655. <https://doi.org/10.1016/j.biopha.2018.08.043>
- Quiroga, A. V., Barrio, D. A., & Añón, M. C. (2015). Amaranth lectin presents potential antitumor properties. *LWT - Food Science and Technology*, 60(1), 478–485. <https://doi.org/10.1016/j.lwt.2014.07.035>
- Rafiq, S., Majeed, R., Qazi, A. K., Ganai, B. A., Wani, I., Rakhshanda, S., Qurishi, Y., Sharma, P. R., Hamid, A., Masood, A., & Hamid, R. (2013). Isolation and antiproliferative activity of *Lotus corniculatus* lectin towards human tumour cell lines. *Phytomedicine*, 21(1), 30–38. <https://doi.org/10.1016/j.phymed.2013.08.005>
- Rahimi, V., Hajizadeh, J., Zibae, A., & Jalali Sendi, J. (2018). Toxicity and physiological effects of an extracted lectin from *Polygonum persicaria* L. on *Helicoverpa armigera* (Hübner) (Lepidoptera: Noctuidae). *Physiological and Molecular Plant Pathology*, 101, 38–44. <https://doi.org/10.1016/10.1016/j.pmp.2017.09.005>
- Ríos-de Álvarez, L., Jackson, F., Greer, A. W., Grant, G., Jackson, E., Morrison, A. A., & Huntley, J. F. (2012). Direct anthelmintic and immunostimulatory effects of oral dosing semi-purified phytohaemagglutinin lectin in sheep infected with *Teladorsagia circumcincta* and *Trichostrongylus colubriformis*. *Veterinary Parasitology*, 187(1–2), 267–274. <https://doi.org/10.1016/10.1016/j.vetpar.2012.01.005>
- Rodrigues Lacerda, R., Silva do Nascimento, E., Thalles Jocelino Gomes de Lacerda, J., da Silva Pinto, L., Rizzi, C., Marques Bezerra, M., Ribeiro Pinto, I., Mateus Pereira Filho, S., de Paulo Teixeira Pinto, V., Cristiano Filho, G., Alberto de Almeida Gadelha, C., & Santi Gadelha, T. (2016). G Model Lectin from seeds of a Brazilian lima bean variety (*Phaseolus lunatus* L. var. cascavel) presents antioxidant, antitumor and gastroprotective activities. *International Journal of Biological Macromolecules*, 95, 1072–1081. <https://doi.org/10.1016/10.1016/j.ijbiomac.2016.10.097>
- Roy, U. K., Lavignac, N., Rahman, A. M., & Nielsen, B. V. (2018). Purification of lectin and Kunitz trypsin inhibitor from soya seeds. *Journal of Chromatographic Science*, 1–7. <https://doi.org/10.1016/10.1093/chromsci/bmy018>
- Saha, R. K., Acharya, S., Jamiruddin, M., & Roy, P. (2014). Antimicrobial effects of a crude plant lectin isolated from the stem of *Tinospora tomentosa*. *The Journal of Phytopharmacology*, 3(1), 44–51.
- Sansone, A. C. M. B., Sansone, M., dos Santos Dias, C. T., & Oliveira do Nascimento, J. R. (2016). Oral administration of banana lectin modulates cytokine profile and abundance of T-cell populations in mice. *International Journal of Biological Macromolecules*, 89, 19–24. <https://doi.org/10.1016/j.ijbiomac.2016.04.049>
- Santi-Gadelha, T., Rocha, B. A. M., Gadelha, C. A. A., Silva, H. C., Castellon, R. E. R., Gonçalves, F. J. T., Toyama, D. O., Toyama, M. H., de Souza, A. J. F., Beriam, L. O. S., Martins, J. L., Joazeiro, P. P., & Cavada, B. S. (2012). Effects of a lectin-like protein isolated from *Acacia farnesiana* seeds on phytopathogenic bacterial strains and root-knot nematode. *Pesticide Biochemistry and Physiology*, 103(1), 15–22. <https://doi.org/10.1016/j.pestbp.2012.02.003>
- Santos, N. D. L., Napoleão, T. H., Benevides, C. A., Albuquerque, L. P., Pontual, E. V., Oliveira, A. P. S., Coelho, L. C. B. B., Navarro, D. M. A. F., & Paiva, P. M. G. (2018). Effect of gamma irradiation of *Moringa oleifera* seed lectin on its larvicidal, ovicidal, and oviposition-stimulant activities against *Aedes aegypti*. *South African Journal of Botany*, 1–6. <https://doi.org/10.1016/j.sajb.2018.05.001>
- Shanmugavel, S., Velayutham, V., Kamalanathan, T., Periasamy, M., Munusamy, A., & Sundaram, J. (2016). Isolation and analysis of mannose/trehalose/maltose specific lectin from jack bean with antibruchid activity. *International Journal of Biological Macromolecules*, 91, 1–14. <https://doi.org/10.1016/j.ijbiomac.2016.05.093>
- Sharma, M., Reddy, H. V., Sindhura, B. R., Kamalanathan, A. S., Swamy, B. M., & Inamdar, S. R. (2017). Purification, characterization and biological significance of mannose binding lectin from *Dioscorea bulbifera* bulbils. *International Journal of Biological Macromolecules*, 102, 1146–1155. <https://doi.org/10.1016/j.ijbiomac.2017.04.085>
- Sharon, N. (2008). Lectins: Past, present and future. *Biochemical Society Transactions*, 36(6), 1457–1460. <https://doi.org/10.1042/BST0361457>
- Sharon, N., & Lis, H. (2004). History of lectins: From hemagglutinins to biological recognition molecules. *Glycobiology*, 14(11), 53–62. <https://doi.org/10.1093/glycob/cwh122>

- Shivamadhu, M. C., Srinivas, B. K., Jayarama, S., & Angatahalli Chandrashekaraiyah, S. (2017). Anti-cancer and anti-angiogenic effects of partially purified lectin from *Praecitrullus fistulosus* fruit on in vitro and in vivo model. *Biomedicine and Pharmacotherapy*, 96, 1299–1309. <https://doi.org/10.1016/j.biopha.2017.11.082>
- Silva, A. F. B., Matos, M. P. V., Ralph, M. T., Silva, D. L., De Alencar, N. M., Ramos, M. V., & Lima-Filho, J. V. (2016). Comparison of immunomodulatory properties of mannose-binding lectins from *Canavalia brasiliensis* and *Cratylia argentea* in a mice model of Salmonella infection. *International Immunopharmacology*, 31, 233–238. <https://doi.org/10.1016/j.intimp.2015.12.036>
- Silva, F. de O., Santos, P. das N., Figueirôa, E. de O., de Melo, C. M. L., de Andrade Lemoine Neves, J. K., Arruda, F. V. S., Cajazeiras, J. B., do Nascimento, K. S., Teixeira, E. H., Cavada, B. S., Porto, A. L. F., & Pereira, V. R. A. (2014). Antiproliferative effect of *Canavalia brasiliensis* lectin on B16F10 cells. *Research in Veterinary Science*, 96(2), 276–282. <https://doi.org/10.1016/j.rvsc.2014.01.005>
- Silva, H. C., Pinto, L. D. S., Teixeira, E. H., Nascimento, K. S., Cavada, B. S., & Silva, A. L. C. (2014). BUL: A novel lectin from *Bauhinia unguolata* L. seeds with fungistatic and antiproliferative activities. *Process Biochemistry*, 49(2), 203–209. <https://doi.org/10.1016/j.procbio.2013.10.020>
- Silva, M. C. C., De Paula, C. A. A., Ferreira, J. G., Paredes-Gamero, E. J., Vaz, A. M. S. F., Sampaio, M. U., Correia, M. T. S., & Oliva, M. L. V. (2014). *Bauhinia forficata* lectin (BfL) induces cell death and inhibits integrin-mediated adhesion on MCF7 human breast cancer cells. *Biochimica et Biophysica Acta - General Subjects*, 1840(7), 2262–2271. <https://doi.org/10.1016/j.bbagen.2014.03.009>
- Silva, M. C. C., Santana, L. A., Mentele, R., Ferreira, R. S., De Miranda, A., Silva-Lucca, R. A., Sampaio, M. U., Correia, M. T. S., & Oliva, M. L. V. (2012). Purification, primary structure and potential functions of a novel lectin from *Bauhinia forficata* seeds. *Process Biochemistry*, 47(7), 1049–1059. <https://doi.org/10.1016/j.procbio.2012.03.008>
- Silva, Pollyanna M., Napoleão, T. H., Silva, L. C. P. B. B., Fortes, D. T. O., Lima, T. A., Zingali, R. B., Pontual, E. V., Araújo, J. M., Medeiros, P. L., Rodrigues, C. G., Gomes, F. S., & Paiva, P. M. G. (2016). The juicy sarcotesta of *Punica granatum* contains a lectin that affects growth, survival as well as adherence and invasive capacities of human pathogenic bacteria. *Journal of Functional Foods*, 27, 695–702. <https://doi.org/10.1016/j.jff.2016.10.015>
- Sindhura, B. R., Hegde, P., Chachadi, V. B., Inamdar, S. R., & Swamy, B. M. (2017). High mannose N-glycan binding lectin from *Remusatia vivipara* (RVL) limits cell growth, motility and invasiveness of human breast cancer cells. *Biomedicine and Pharmacotherapy*, 93, 654–665. <https://doi.org/10.1016/j.biopha.2017.06.081>
- Siritapetawee, J., Limphirat, W., Wongviriyi, W., Maneesan, J., & Samosornuk, W. (2018). Isolation and characterization of a galactose-specific lectin (EantH) with antimicrobial activity from *Euphorbia antiquorum* L. latex. *International Journal of Biological Macromolecules*, 120, 1846–1854. <https://doi.org/10.1016/j.ijbiomac.2018.09.206>
- Thakur, K., Kaur, T., Singh, J., Rabbani, G., Khan, R. H., Hora, R., & Kaur, M. (2017). *Sauromatum guttatum* lectin: Spectral studies, lectin-carbohydrate interaction, molecular cloning and in silico analysis. *International Journal of Biological Macromolecules*, 104, 1267–1279. <https://doi.org/10.1016/j.ijbiomac.2017.06.123>
- Umáro, A., Queiroz, M., Jose, V., Osterne, S., Pinto-junior, V. R., Paulo, L., Silva-filho, J. C., Debray, H., Anderson, B., Rocha, M., Delatorre, P., Souza, C., Correia, C., Maria, A., Assreuy, S., Santiago, K., & Sousa, B. (2016). Lectins from *Parkia biglobosa* and *Parkia platycephala*: A comparative study of structure and biological effects. *International Journal of Biological Macromolecules*, 92, 194–201. <https://doi.org/10.1016/j.ijbiomac.2016.07.032>
- Van Hove, J., Stefanowicz, K., De Schutter, K., Eggermont, L., Lannoo, N., Al Atalah, B., & Van Damme, E. J. M. (2014). Transcriptional profiling of the lectin ArathEULS3 from *Arabidopsis thaliana* toward abiotic stresses. *Journal of Plant Physiology*, 171(18), 1763–1773. <https://doi.org/10.1016/j.jplph.2014.08.009>
- Van Hove, Jonas, De Jaeger, G., De Winne, N., Guisez, Y., & Van Damme, E. J. M. (2015). The Arabidopsis lectin EULS3 is involved in stomatal closure. *Plant Science*, 238, 312–322. <https://doi.org/10.1016/j.plantsci.2015.07.005>
- Vigneshwaran, V., Thirusangu, P., Madhusudana, S., Krishna, V., Pramod, S. N., & Prabhakar, B. T. (2016). The latex sap of the ‘Old World Plant’ *Lagenaria siceraria* with potent lectin activity mitigates neoplastic malignancy targeting neovasculature and cell death. *International Immunopharmacology*, 39, 158–171. <https://doi.org/10.1016/j.intimp.2016.07.024>
- Villanueva, J., Quirós, L. M., & Castañón, S. (2015). Purification and partial characterization of a ribosome-inactivating protein from the latex of *Euphorbia trigona* Miller with cytotoxic activity toward human cancer cell lines. *Phytomedicine*, 22(7–8), 689–695. <https://doi.org/10.1016/j.phymed.2015.04.006>
- Vishweshwaraiah, Y. L., Prakash, B., & Gowda, L. R. (2018). Expression profiling of the *Dolichos lablab* lectin during germination and development of the seed. *Plant Physiology and Biochemistry*, 124, 10–19. <https://doi.org/10.1016/j.plaphy.2017.12.040>
- Wu, J., Wang, J., Wang, S., & Rao, P. (2016). Lunatin, a novel lectin with antifungal and antiproliferative bioactivities from *Phaseolus lunatus* billb. *International Journal of Biological Macromolecules*, 89(2), 717–724. <https://doi.org/10.1016/j.ijbiomac.2016.04.092>
- Wu, L., Liu, T., Xiao, Y., Li, X., Zhu, Y., Zhao, Y., Bao, J., & Wu, C. (2016). *Polygonatum odoratum* lectin induces apoptosis and autophagy by regulation of microRNA-1290 and microRNA-15a-3p in human lung adenocarcinoma A549 cells. *International Journal of Biological Macromolecules*, 85, 217–226. <https://doi.org/10.1016/j.ijbiomac.2015.11.014>
- Xiao, X., He, H., Ding, X., Yang, Q., Liu, X., Liu, S., Rang, J., Wang, T., Zuo, M., & Xia, L. (2015). Purification and cloning of lectin that induce cell apoptosis from *Allium chinense*. *Phytomedicine*, 22(2), 238–244. <https://doi.org/10.1016/j.phymed.2014.12.004>
- Zárate, G., Sáez, G. D., & Chaia, A. P. (2017). Dairy propionibacteria prevent the proliferative effect of plant lectins on SW480 cells and protect the metabolic activity of the intestinal microbiota in vitro. *Anarobe*, 44, 58–65. <https://doi.org/10.1016/j.anaerobe.2017.01.012>
- Zhou, W., Gao, Y., Xu, S., Yang, Z., & Xu, T. (2014). Purification of a mannose-binding lectin *Pinellia ternata* agglutinin and its induction of apoptosis in Bel-7404 cells. *Protein Expression and Purification*, 93, 11–17. <https://doi.org/10.1016/j.pep.2013.09.018>