COMBATING OF ESCHERICHA COLI STRAINS AGAINST SALINITY BY THE ANTIOXIDANT DEFENSE SYSTEM AND OSMOREGULATION: BIOCHEMICAL AND PHYSIOLOGICAL APPROACHES

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ARTICLE INFO

Regular article

ABSTRACT

The study was designed to elucidate the impact of salinity on biochemical and physiological properties of two Escherichia coli strains (6E and S39) isolated from different sites. The strains were exposed to 600 and 1200 mM NaCl and were harvested after 12h and 24h of incubation. The lipid peroxidation levels, osmolyte accumulation, H2O2 content and antioxidant system (superoxide dismutase (SOD), catalase (CAT), peroxidase (POX), ascorbate peroxidase (APX) and glutathione reductase(GR)) of the bacteria were investigated. While thiorubicin acid reactive substances (TBARS) and H2O2 content of S39 has increased steadily with salinity, however, it has not changed at 600 mM NaCl concentration in strain 6E. Accumulation of the osmolytes in 6E increased depending on salinity and application time. In S39 except for glycine-betaine, contents of other osmolytes were either unchanged or decreased. Activities of SOD, CAT, POX, APX, GR and NOX have increased as compared to the control group in 6E exposed to 600 and 1200 mM salt for 12h. In S39, CAT, POX, and GR activities decreased exposed to salinity. Consequently, it can be argued that (i) the different antioxidant responses of 6E in salinity plays a key role which tend to make 6E the more tolerant (ii) this tolerance is closely related to the increased antioxidant capacity against reactive oxygen species and is related to the increased accumulation of osmolytes.

Keywords: Escherichia coli, salt stress, antioxidant defense, osmolyte, isozyme

INTRODUCTION

Bacteria are exposed to different environmental stresses such as acidity, pH, NaCl, heat, electrical potential, bacteriocins, and competitive flora found in foods except stress factors such as high salinity, drought, light, O2, cold, detergent and disinfectants in nature (Dikici, 2009; McMahon et al., 2007). Bacteria can die due to the stresses they encounter, and they can survive by activating adaptation mechanisms. Consequently, bacteria have developed a panoply of mechanisms in response to environmental stresses, in an endeavor to trap, preserve, and transform the energy essential for their biosynthesis and growth. With such adaptive mechanisms, they can thrive in stressful environments for a long time or die due to damage to their metabolism as a result of this stress. For decades, researchers have wondered how microorganisms live in extreme environments and the nature of subsequent adaptive mechanisms in extremely stressful environments. According to preliminary research in 1933, it has been reported that heat adaptation of osmotic stress-exposed bacteria increase (Fay, 1933). When bacteria encounter a sub lethal stress called mild stress, their number does not decrease. However, it results in a halt or decrease in the reproduction rate. When the microorganism encounters a moderate stress, microbial growth stops as well as a decrease in the viability of bacteria. Extreme stress, either called extreme or severe stress, is a lethal condition for bacterial cells and results in the death of the majority of the bacterial population (Neidhardt & VanBogelen, 2000).

Oxidative and osmotic stresses are parts of important abiotic stress factors that microorganisms encounter in environment and foods. When bacteria are subjected to oxidative stress, reactive oxygen species (ROS) such as superoxide, hydroxyl radicals and hydrogen peroxide can be formed, and the accumulation of such various oxidants causes damage to cellular proteins, nucleic acid and lipid (Munna et al., 2013). To keep ROS at a non-toxic concentration, enzymatic and non-enzymatic cellular antioxidants regulation the balance between their production and scavenging. Enzymatic antioxidants, such as superoxide dismutase (SOD), catalase (CAT), peroxidase (POX), ascorbate peroxidase (APX) and glutathione reductase (GR), used to scavenge excess of internal-external ROS (Staerck et al., 2017). Also, many proteins are known to be activated by oxidative stress, acting as antioxidants. Others are especially necessary to repair oxidative damage, especially nucleic acid damage. Salinity, one of the main abiotic stress factors, affects both agricultural lands and many living things negatively. Salinity causes various disruptions at the cellular, physiological and molecular level as well as various growth processes of bacteria. Bacteria develop various tolerance strategies in response to salt stress. In response to salt stress (i) the activity of various antioxidant enzymes are increased destroying ROS formed as by-product of metabolism, (ii) osmolyte synthesis is promoted, (iii) regulating ion uptake mechanism by gene expression, (iv) activating stress-related genes synthesis of transcription factors, and (v) promoting the production of stress proteins are known to be among the major tolerance strategies. The metabolism best characterized in bacteria under hyperosmotic conditions is the accumulation of so-called intracellular osmolytes. Accumulation of compatible solutes can be achieved via synthesis as well as by extracellular transfer. These liquids are polar and are highly soluble in the cell. Even at very high concentrations, they can cope with osmotic pressure without affecting cellular functions. Glycine betaine, proline, ectoine, carminine, choline and trehalose are the most widely known compatible substances. The accumulation of these compounds in the cytoplasm is regulated either directly by altered enzyme activity or by the level of gene transcription (Bremer, 2000).

Pathogenic and nonpathogenic Escherichia coli have previously been reported to show a stress response to sublethal environmental stresses (Chung et al., 2006). The stress responses may enable survival under more negative conditions, cause resistance and increased virulence. Not only microorganisms may come across stress conditions in the environment but also they come across with those conditions in foods, especially minimally processed foods. Temperature, salinity, water content, and pH are the main extrinsic (environmental) factors which effects the survival and growth of E. coli in foods, also development of bacterium interacts with food related (intrinsic) factors. One of the factors affecting microbial growth is the inclusion of preservative. To additive and preservative in food, sodium chloride has been used. NaCl, known since time immemorial as antimicrobial agent, has frequently been incorporated as a component in meat and meat products (Abdulkarim et al., 2009). Although members of Enterobacteriaceae do not tolerate high salt concentration, specific strains of E. coli are halo-tolerant and able to strive in environments ranging from very dilute aqueous solutions of main nutrients to media including molar concentrations of salts or non-electrolyte solutes. It has always intrigued microbiologists how E. coli cope with such exposure and which physiological changes, cause it to survive and at the same time retaining infectivity of the bacterium. Adaptation to the stress both in environmental and food borne microorganisms have been ignored in the past. However, nowadays the importance of this phenomenon in the field of environmental and food safety is increasing. Thus, understanding the impacts of environmental stresses on the physiological tolerance of E. coli are urgently needed and are important in order to evaluate and minimize the risk of food-borne illness.
This study was therefore designed to determine and compare the physiological and biochemical responses of two *E. coli* strains, isolated and identified from the different sites, under oxidative stress by employing different concentrations of salt (NaCl). The specific objectives set for this study are (i) to evaluate the changes in activities of the enzymes taking part in antioxidant defense system such as SOD, CAT, POX, APX, and GR, (ii) to assess the lipid peroxidation levels and hydrogen peroxide (H$_2$O$_2$) contents and (iii) to understand and highlight the osmoprotectant metabolism under salt stress conditions.

**MATERIAL AND METHODS**

**Sample collection and isolation of *Escherichia coli* strains**

*E. coli* strains used in the present study were isolated from two different sites in 2014. For the S39 numbered strain, well water sample was taken from the water sampling site as determined with the help of the Konya Public Health Laboratory commissioner. For the 6E numbered strain, water and sediment samples were taken from the watercourse between Çihanbeyli-Golyazi (38° 37.270’N and 33° 08.986’E, 904 m). Sterile water sampling bottles contained 50 mg of sodium thioulate to neutralize any residual chlorine in the water were used to collect samples.

The membrane filtration method as described by Uysal et al. (2013) was used to isolate *E. coli* from water samples. After filtration, the membrane containing the bacteria was placed on a selective differential medium (Standard Lactose Tri Tetrazolium Chloride (TTC) Agar with Tergitol 7) and incubated at 35°C for 24 h to resuscitate the injured or stressed bacteria and then at 44°C for 22 h. After incubation, yellow or yellow-brown colonies on TTC agar were chosen to perform oxidase and indole tests. Oxidase (+) and indole (+) colonies were transferred to Eosin Methylene Blue Agar. Finally, Chromocult TBX (Tryptone Bile X-glucuronide) agar medium was used for identification purposes (Uysal et al., 2013).

**Halo tolerance of *E. coli* strains**

The halo tolerance tests were performed in Luria Bertani (LB) medium supplemented with NaCl in the range of 0.3%–14% (w/v) (Cronin et al., 2013). The bacterial growth was calculated by measuring of absorbance at 600 nm wavelength. Also growth was visualized by the addition of 2.3, 5, TTC to culture wells for qualitative evaluation.

**Culture conditions and harvesting of the strains**

After the determination of halo tolerance of strains, two salt concentrations (600 mM and 1200 mM) were selected for the application of salinity. The halo tolerance tests were performed in Luria Bertani (LB) medium supplemented with NaCl in the range of 0.3%–14% (w/v) (Cronin et al., 2013). At the same time, standard LB broth without salt was used as control. The time-depend bacterial growths were measured at 600 nm wavelength by using UV spectrometer (Shimadzu UV-1800, Japan) as absorbance (bacteria were not counted for logarithmic growth) for every hour and growth curve was drawn (Figure 1). Two time intervals were determined (12h and 24h) according to this curve for the harvesting of the bacterial cells in the main experiment. In the main assay the volumes of the LB mediums were prepared to 1000 ml for each group and culture conditions applied as above.

Bacterial cells (1000 ml) were harvested from liquid LB medium at 12th and 24th h of the incubation period. Then cell suspensions were centrifuged at 12000 × g for 10 min, washed twice with ice-cold 0.9% sodium chloride solution, and the specimens were dried in an incubator and stored at -80°C until use (Lin et al., 2009).

**Determination of hydrogen peroxide content**

Determination of H$_2$O$_2$ content was performed according to method described by Beers and Sizer (1952).

**Evaluation of lipid peroxidation levels**

The level of lipid peroxidation was determined by thiobarbituric acid reactive substances (TBARS) according to Heath and Packer (1968).

**Changes of osmoregulatory solutes accumulation**

Determination of proline (Pro) content was done according to the procedure defined by Bates et al. (1973). Total amount of choline (Cho) and glycine betaine (GB) were determined according to Grievie and Gratian (1983).

**Enzyme extraction and analysis of isozyme and/or enzyme compositions**

Antioxidant enzyme assays were performed on freshly collected cells using the method of Beers & Sizer, 1952. Homogenization of cells was done in 50 mM Tris-HCl (pH 7.8) containing 0.1 mM ethylenediaminetetraacetic acid (EDTA), 0.2% Triton X-100, 1 mM phenylmethylsulfonyl fluoride and 2 mM dithiobis (Beers and Sizer (1952). The total soluble protein content of the enzyme extracts was determined (Bradford, 1976) using bovine serum albumin as a standard.

Samples containing equal amounts of protein (25 µg) were subjected to non-denaturing polyacrylamide gel electrophoresis (PAGE) as described by (Laemmlli, 1970) with minor modifications. Superoxide dismutase (SOD) activity was detected by photochemical staining using riboflavin and NBT (Beauchamp & Fridovich, 1971). The different types of SOD were discriminated by incubating gels with different types of SOD inhibitors before staining. Mn-SOD activity was resistant to both inhibitor treatments and Cu/Zn-SOD activity was sensitive to 2 mM KCN. Cu/Zn-SOD and Fe-SOD activities were inhibited by 3 mM H$_2$O$_2$ (Vittoria et al., 2001). The total SOD (EC 1.15.1.1) activity assay was based on the method of Beauchamp and Fridovich (1971) After electrophoresis of samples containing 25 µg protein, catalase (CAT) isozymes were detected according to Woodbury et al. (1971). Total catalase [CAT] (EC 1.11.1.6) activity was estimated according to the method of Bergmeyer and Gawehn (1970). Total peroxidase [POX (EC 1.11.1.7)] activity was based on the method described by Herzog and Fahimi (1973). Electrophoretic ascorbate peroxidase (APX) separation was performed according to Mittler and Zilkens (1993). Total APX (EC 1.11.1.11) activity was measured according to Nakano and Asada (1981). Total glutathione reductase [GR (EC 1.6.4.2)] activity was measured according to Foyer and Halliwell (1976).

**Statistical analysis**

All data obtained were subjected to a one-way analysis of variance (ANOVA). Statistical analysis of the values was performed using SPSS 20.0. Tukey’s post-test was used to compare the treatment groups. Comparisons with $p < 0.05$ were considered significantly different.

**RESULTS AND DISCUSSION**

**Changes of Hydrogen peroxide (H$_2$O$_2$) contents**

One of the main important parameters in determining the effect of salt stress was to evaluate the H$_2$O$_2$ level of the bacteria under salt stress condition. The H$_2$O$_2$ content is usually low in tolerant species. During the study, the application of 600 mM NaCl did not change the amount of H$_2$O$_2$ from the 6E strain (Figure 2A). In this strain, the increase in H$_2$O$_2$ occurred only at a high concentration of 1200 mM. In the strain S39, the increase of H$_2$O$_2$ was determined in parallel with the application time and concentration of salinity (Figure 2B). As a result of the 24-hour harvest of these bacteria, the amount of H$_2$O$_2$ in 600 and 1200 mM NaCl were increased by 77.2% and 62.1%, respectively. When the two strains were compared in terms of H$_2$O$_2$ contents, it was observed that the H$_2$O$_2$ content of S39 was higher than 6E.

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**Figure 1** Time-dependent growth curve of strains 6E and S39
**Figure 2** Mean changes in H$_2$O$_2$ content and lipid peroxidation levels (TBARS) of *E. coli* strains under salt stress (n = 6). A-C) 6E strain B-D) S39 strain. The different letters on the columns indicate statistically different values in the same strain (ANOVA, Tukey’s post-test *p* <0.05).

**Figure 3** Mean changes in osmolytes of *E. coli* strains under salt stress (n = 6). A-C-E) 6E strain B-D-F) S39 strain. The different letters on the columns indicate statistically different values in the same strain (ANOVA, Tukey’s post-test *p* <0.05).
Figure 4 Effects on activity staining of SOD isozymes (A-B) and total SOD activity (C-D) in *E. coli* strains exposed to 600 and 1200 mM NaCl for 12 and 24 hours (samples containing 25 µg protein of the gel were pipetted; std. 0.5 units of SOD). (Data A and C belong to strain 6E; data B and D belong to strain S39. The different letters on the columns indicate statistically different values in the same strain (ANOVA, Tukey’s post-test *p* <0.05).

**Lipid peroxidation levels**

One indicator of damage caused by salt stress at the cell level is lipid peroxidation (TBARS) occurring at the level of the membrane structures. Measurement of TBARS contents and determination of peroxidation levels of membrane lipids is an important parameter in the assessment of the severity of salinity-induced oxidative stress and bacterial susceptibility and tolerance. Changes in malondialdehyde levels of *E. coli* strains isolated from different localities are shown in Figure 2C-D. While the TBARS of the strain (6E) isolated from Golyazi site did not change in both short (12 h) and long-term (24 h) 600 mM NaCl applications, it increased by 23.9% and 51.0% in the 1200 mM salt treated groups, respectively (Figure 2C). The unchanged TBARS content at 600 mM salinity is one of the most important parameters indicating that the bacteria are not affected by the harmful effects of salinity at this concentration. In the strain isolated from the well water (S39), the increase of TBARS level was determined in parallel with the application time and concentration of salinity (Figure 2D). This increase was 2.2 fold of the control at 1200 mM for 12 h and 2.8 fold at 1200 mM for 24 h under stress. The highest lipid peroxidation increase in both strains was determined in bacteria exposed to 1200 mM NaCl for a long time. When these two strains isolated from different localities are compared in terms of lipid peroxidation contents, *E. coli* samples S39 showed significantly higher malondialdehyde content compared to strain 6E.

**Effects of Salinity on Osmoprotectants**

One of the most common osmolytes accumulated under environmental stresses as salt stress is proline (Pro). Pro is not only an osmotic regulator, but also serves as an antioxidant and carbon source. The proline contents of strain 6E increased...
significantly over the entire experiment and treatments when compared with the non-stress (Figure 3A). The highest proline content was 6.76 fold of the control in bacteria subjected to 1200 mM NaCl for 24 h. The proline content of strain S39 did not change as compared to the control in both 600 and 1200 mM salinity applications for a short period (Figure 3B). The proline level of this strain increased by 31.1% in samples at 600 mM NaCl only for 24 h compared to the control. Long-term 1200 mM salinity decreased by 33.5% compared to the control. This decrease in proline content clearly shows that the strain is adversely affected by high salt stress.

As in all living organisms under environmental stress conditions such as salinity, osmotic balance between internal and external cellular environment must be maintained. Accumulation of osmolytes such as choline in the cell is a common physiological response to achieve this balance. It was determined that choline contents of 6E strain increased during the whole application period compared to control. The highest choline enhancement was observed in samples exposed to 1200 mM NaCl for 24 h with a ratio of 93.4% (Figure 3C). It can be said that choline accumulation may be a good strategy for 6E strain to cope with high salinity. In strain S39, only 39.2% increase was observed in bacteria under long-term high salinity (1200 mM), and in all other applications the choline content was either decreased or unchanged (Figure 3D).
Glycine betaine increases the osmotic potential of cells and maintained the turgor state by internally increasing the osmotic pressure. In both strains, the increased in glycine betaine content was directly proportional to the salt stress. The highest glycine betaine content in both strains was observed in bacteria exposed to 1200 mM NaCl for 24 h (Figures 3E-F).

Antioxidant enzyme/isozyme composition

Total SOD activity of the strain 6E increased in all applications except 1200 mM NaCl applied group for 24 h (Figure 4C). This increase in SOD activity may reflect an increase in the production of \( \text{O}_2^− \) radicals in consequence of electron leaks from the electron transport chain. The total SOD (tSOD) activities of the groups treated with 1200 mM salinity for 24 h were not markedly different from the control. The highest increased in SOD was observed in bacteria at 600 mM salinity at both sampling times. These increase rates were 52% and 17.8% respectively. The tSOD activities of these strains, which we considered to be tolerant, were higher under short-term (12 h) salinity than long-term (24 h) treatments. tSOD activities of S39 isolated from well water varied. tSOD activity was significantly increased in all salt stress applications except for the short-term 1200 mM NaCl group. A 29.9% reduction in 1200 mM NaCl was observed for 12 h when compared to the control (Figure 4D).

A total of five different APX isozymes were determined by electrophoretic separation of the APX enzyme and salt applications of both groups of bacteria. Increased APX1 and APX5 isozyme plays an important role in the highest APX activity of strain 6E (Figure 7A). In S39 strain, the lowest APX activity is due to the decrease in the activities of APX3-5 isozymes (Figure 7B).

Nine different SOD isozymes were determined by electrophoretic separation of SOD enzyme from 6E strain (Figure 4A). One of these identified isozymes was defined as Mn-SOD (which is not affected from KCN and \( \text{H}_2\text{O}_2 \)); two of them are Fe-SOD (unaffected by KCN but inhibited by \( \text{H}_2\text{O}_2 \)) and the other six isozymes were defined as Cu/Zn-SOD according to inhibition by both KCN and \( \text{H}_2\text{O}_2 \). Increased in Mn-SOD and Cu/Zn-SOD isozymes play an important role in increasing SOD enzyme activities.

A total of three different SOD isozymes were determined by electrophoretic separation of SOD enzyme by salt applications from S39 strain. One of these isozymes was defined as Mn-SOD, another one was defined as Fe-SOD and the other isozyme was defined as Cu/Zn-SOD. The 29.9% reduction in bacteria treated with 1200 mM salinity for 12 h was due to the reduction in Fe-SOD and Cu/Zn-SOD isozymes (Figure 4B).

Short and long term salt stress treatments in strain 6E caused different changes in total CAT (tCAT) activity (Figure 5C). tCAT activity was increased when exposed to short-term salinity and decreased in long-term applications. The highest tCAT activity of this strain was determined as 4.3 fold of the control group in 600 mM NaCl for 12 h. In short-term treatments, tCAT activity decreased as opposed to increasing salt concentration in strain S39 (Figure 5D). Both the lowest and highest tCAT activities were detected in bacteria treated with 1200 mM NaCl at both sampling times.

As a result of salt treatments, a total of three different CAT isozymes were determined by electrophoretic separation of CAT enzyme for both strains. Increased CAT2 isozyme plays significantly role in the highest CAT activity of the 6E strain (Figure 5A). Increased in CAT1 and CAT2 isozymes are of great importance in the highest CAT activity of S39 strain (Figure 5B). The lowest CAT activity is due to the decrease in activity of CAT3 isozyme.

Peroxidase (POX) activities of strain 6E increased significantly at both sampling times when compared to the control group (Figure 6A). The highest POX activity under both short and long term salinity was determined to be 3.7 and 2.8 fold of the control in 600 mM NaCl treated bacteria. POX activity of S39 decreased for both sampling times and these reduction rates were determined as 64.9% and 33.6%, respectively (Figure 6B).

Glutathione reductase (GR) activity of 6E group bacteria induced in both short and long term salt applications. GR activity was also increased in parallel with salt treatment time and concentration. The highest GR increase at both sampling times was determined to be 2.1 and 3.1 folds of the control in bacteria treated with 1200 mM NaCl, respectively (Figure 8A). In the S39 group, the GR activity was either unchanged or decreased significantly with salinity compared to the control. The highest reduction rates are 43.3% at 1200 mM in short-term applications and 43.5% in long-term applications (Figure 8B).
In general, NOX activity was increased in 6E strain as a result of short and long term salt applications. The highest increase in the NOX activity was observed at 1200 mM, the highest concentration at both sampling times (Figure 8C). NOX activities of S39 strain differed in short and long term stress treatments. While NOX activity was either unchanged or increased in short-term applications, it was significantly decreased during the 24 h of stress. The lowest NOX activity was determined in bacteria exposed to 1200 mM salinity for 24 h with a decrease of 61.8% compared to the control (Figure 8D).

**DISCUSSION**

Microbiological stress induced by environmental and biological factors, can individually or together, produce significant changes in the physiological events of microorganisms. Various physiological, morphological, biochemical and molecular reactions occur in microorganisms exposed to stress factors such as salinity, drought, pH, acidity, low and high temperatures, nutrient deficiencies or abundance, heavy metals and radiation. Microorganisms can produce a protective or adaptive response when exposed to stress. When adapting to one stress, resistance to another stress is called cross protection (Youssif & Courtney, 2003). Exposure of microorganisms to salinity results in an increase in the production of NOX. Osmotically active molecules keep the positive pressure turgor required for cell division. Because most intracellular macromolecules are sensitive to high levels of inorganic ions, most organisms tend to restrict cytoplasmic ion strength to very low levels. Prokaryotes in high osmolality environments have some strategies for survival and development.

High salt concentrations can lead to major problems for living systems. Osmoadaptation is defined as the physiological and genetic findings of adaptation to low and high water environments (Galinski, 1995). In principle, two strategies have been developed to deal with high osmolarity— conquering stress factors or by de novo synthesis. Osmotically active molecules keep the positive pressure turgor required for cell division. Because most intracellular macromolecules are sensitive to high levels of inorganic ions, most organisms tend to restrict cytoplasmic ion strength to very low levels. Prokaryotes in high osmolality environments have some strategies for survival and development.

**Figure 8** Effects on GR (A-B) and NOX (C-D) activities in *E. coli* strains exposed to 600 and 1200 mM NaCl for 12 and 24 hours. (Data A-C belong to strain 6E; data B-D belong to strain S39). The different letters on the columns indicate statistically significant differences in the values of the same strain (ANOVA, Tukey’s post-test p <0.05).

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exposing to oxidative stress. Our TBARS results in line with the values reported by Aubron et al. (2012).

In the present study, proline levels were measured depending on the salt concentration and reproduction time of the strains exposed to stress. According to the results, 6E strain which is more tolerant to salt treatment was compared to control treatment line parallel with incrementing salt concentration in short and long term salt treatments. The more susceptible strain S39 may not have increased proline in short-term salt treatment. As a result, it could not fully achieve osmotic stabilization. In prolonged applications, although proline level increased at 600 mM, proline level in the medium was calculated as 50% of the maximal NtcI decreased in halophilic and halotolerant bacteria (Nyvssl & Leisola, 2001; Reed et al., 1984; Waditee et al., 2003). In the present study, the increase in osmolyte due to salinity in the 6E strain tend to corroborate to results reported by previous studies.

Superoxide dismutase (SOD), a metalloenzyme, is the key antioxidant enzyme in defense against ROS. SOD is catalytic in oxidative stress in all oxygen-breathing organisms (Gill & Tuteja, 2010). In terms of SOD activity, short-term salt treatments were increased in the 6E strain which was tolerant from the strains exposed to salinity compared with the non-stress group. This increased in SOD activity may reflect an increase in the scavenging of O₂⁻ radicals produced in consequence of electron leakage from the electron transport chain. In S39 strain, SOD activities producing increased more in short term salt treatments. Catalases are enzymes that are absolutely necessary for the detoxification of ROS under stressful conditions. In terms of CAT activities, 6E strain increased CAT activity only in 600 mM concentration in short-term salt treatments when compared to control. S39 showed the lowest activity at 120 mM NaCl concentration in short-term salt treatments. Comparison of SOD and CAT activities, and was also argued that new SOD isozymes will be increased in the 6E strain which was tolerant from the strains exposed to salinity whereas the APX enzyme is interested in H₂O₂, catalase and peroxidase. CAT and POX are susceptible to H₂O₂ radicals produced in consequence of electron leakage from the electron transport chain. Increased 3.5 times of control groups. In the study conducted by Yao et al. (2006), the effect of an insecticide acectamiprid on E. coli K12, Pseudomonas FH2 and B. subtilis was investigated. They identified changes in activity of SOD, CAT, ATPase and SOD isoforms. It was observed that SOD and CAT activities increased at concentrations of acectamiprid depending on the dose. It was emphasized that acectamiprid may cause a certain oxidative stress on the treated bacteria, an increase SOD and CAT activities, and was also argued that new SOD isoforms will be expressed. In a similar study, Lin et al. (2009) bensulifuron-methyl (BSM) was applied to three bacteria (E. coli K12, Bacillus subtilis B19, B. megatrum L1) for investigating the activity of antioxidant enzymes. All three bacteria were found to have salt tolerance. CAT and ATPase activities increased with prolonged 12 h of exposure to BSM, enzyme activities were observed to vary depending on the dose. They found that the activity of all the enzymes began to increase 1-1.5 h after exposure to BSM, followed by a decrease, and an increase again at 9 or 14 h, respectively. Shao et al. (2009) studied the role of antioxidant enzymes in bacterial resistance to nicotine toxicity in their studies and they used nicotine-degrading Pseudomonas sp. HF-1 and two standard strains E. coli and B. subtilis. It was found that SOD and CAT play significant roles in resistance to nicotine stress in E. coli. In Pseudomonas sp. HF-1, they found that too many enzymes play a role in resistance to nicotine stress. In the study of Zhang et al. (2012), the oxidative stress responses of E. coli K12 and B. subtilis B19 exposed to atrazine were examined. It was observed that SOD, CAT, GST and T-AOC enzyme responses were induced by atrazine exposure. In the present study, SOD and CAT activities increased in 6E strain tend to show that these enzymes play an active role in scavenging excessive amounts of stress-induced radicals. However, the constant increase of TBARS and H₂O₂ contents in S39 strain under salinity clearly shows that these enzyme activities are inadequate for adaptation. Peroxisome is one of the important enzymes that play a key role in the scavenging of H₂O₂ produced by SOD scavenging of O₂⁻ radicals (Asada & Takashi, 1987). In this study, 6E strain increased POX activity by 3.7 and 2.8 fold in control to the control at both application times. In S39 strain, POX activity was unchanged at a dose of 1200 mM NaCl but the CAT was increased 3.5 times of control at 24 times of control. Although CAT activity increased 3.2 times of control groups. In the study conducted by El-Rab et al. (2013), POX, CAT and APX antioxidant enzyme activities were induced increasing salt concentration in S39 strain. In general, NOX activity was increased in 6E strain as a result of short and long term salt treatments. NOX activity of strain S39 was either unchanged or increased in short-term applications, but significantly decreased in saline with long-term applications when compared to control. In a study conducted by Waditee et al. (2003), POX, CAT and APX antioxidant enzyme activities were induced increasing salt concentration in S39 strain. In general, NOX activity was increased in 6E strain as a result of short and long term salt treatments. NOX activity of strain S39 was either unchanged or increased in short-term applications, but significantly decreased in saline with long-term applications when compared to control. In a study conducted by Waditee et al. 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