

REVIEW

BIOFLAVONOID QUERCETIN-FOOD SOURCES, BIOAVAILABILITY, ABSORBTION AND EFFECT ON ANIMAL CELLS

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ABSTRACT

Bioflavonoid quercetin is found in the edible portion of the majority of dietary plants. The absorption and metabolism of quercetin is still poorly understood. It is known that aglycone form of quercetin, which is absorbed better than quercetin administered in nonglucosidic forms Absorbed quercetin is probably extensively modified before being excreted by kidneys. It has a broad range of activities within animal cells and occurs to be able to prevent or reduce the development of different type of diseases.

Keywords: bioflavonoid, quercetin, sources, bioavailability, absorbtion, animal cells

INTRODUCTION

Bioflavonoid quercetin belongs to a group of natural substances with variable phenolic structures and is found in fruit, vegetables, grains, bark, roots, stems, flowers, tea, and wine (Middleton, 1998).

The absorption and metabolism of quercetin is still poorly understood. For many years, it was believed that quercetin was not absorbed at all, since no unchanged compound could be measured in the plasma after oral administration. In the last few years, it has become clear that theis bioflavonoid is indeed absorbed but heavily metabolized prior to reaching the

plasma. Most of the metabolism is in the form of glucuronidation or the formation of glucuronide conjugates (Drewa et al., 2001).

Quercetin has a broad range of activities within cells (Bjeldanes and Chang, 1977). Scientific studies suggest its antioxidative (Song et al., 2001), antiproliferative (Yoshida et al., 1992), antiinflammatory (Comalada et al., 2005; Dias et al., 2005), anticarcinogenic (Soleas et al., 2006), antihypertensive (Duarte et al., 2001), antidiabetic (Vessal et al., 2003) effect and is able to protect different types of cells against various diseases such as osteoporosis, certain forms of cancer, pulmonary and cardiovascular diseases but also against aging (Boots and Guido, 2008).

Quercetin

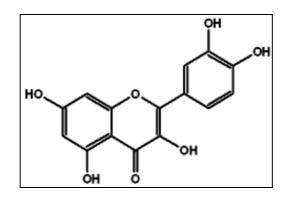


Figure 1 Molecular structure of quercetin (Moskuag, 2004)

Food sources

Bioflavonoid quercetin found in the edible portion of the majority of dietary plants (e.g., citrus, berries, leafy vegetables, roots, tubers and bulbs, herbs and spices, legumes, cereal grains, tea, and cocoa) (Singleton, 1981). Quercetin is ingested as a major constituent in the diet (Bjeldanes and Chang, 1977; Holman and Katan, 1997). Content of aglycone form of quercetin, which is absorbed better than quercetin administered in nonglucosidic forms, varies among different food sources (see Table 1).

Table 1 Content	of aglycone for	orm of querce	etin in differe	nt food sources	(pursuant to Erlund,
2004)					

Forms of quercetin	Source	Content of aglycone (mg/kg) and reference	
Quercetin-3,4'-glucoside	Onion	284-486 (Hertog et al., 1992)	
Quercetin-3-glucoside			
Quercetin-3-	Black tea	10-25 (Hertog et al., 1993)	
rhamnoglucoside (rutin)	Diack tea		
Quercetin-3-galactoside			
Quercetin-3-rhamnoside	Annlo	21-72 (Hertog et al., 1992)	
Quercetin-3-arabinoside	Apple		
Quercetin-3-glucoside			
Quercetin-3-			
rhamnoglucoside			
Quercetin-3-rhamnoside	Black currant	44 (Häkkinen et al., 1999)	
Quercetin-3-galactoside			
Myrisetin-3-glucoside			

Bioavailabity and absorbtion

Like many other compounds absorbed quercetin is probably extensively modified before being excreted by kidneys (Ueno et al., 1982).

Quercetin aglycone and its glucosides are absorbed better than quercetin administered in nonglucosidic forms. The bioavailability of 2 quercetin glucosides, 3-glucoside and 4'-glucoside, do not differ When quercetin and its derivatives are provided for consumption along with their natural sources in which these compounds are dispersed in the matrix, quercetin aglycone is more bioavailable than its glucosides. This finding suggests that in some cases, bioavailability of isolated food components consumed as food supplements could be less than when they are consumed with the food matrix (Olthof et al., 2000). Autors Graefe et al. (2001) in theirs scientific study observed that the bioavailability of quercetin from onion, in which a variety of quercetin glucosides is present, is comparable to the bioavailability of isolated quercetin 4-glucoside. The lipophilic character of quercetin suggests that it can cross enterocyte membranes via simple diffusion (Wiczkowski et al.,

2008). Urinary excretion of quercetin seemed to be a small but constant function of quercetin intake (**Young et al., 1999**). Nowadays is known that humans absorb appreciable amounts of quercetin and that absorption is enhanced by conjugation with glucose (Hollman et al., 1995).

Effect on animal cells

Quercetin has a broad range of activities within cells (**Bjeldanes and Chang, 1977**). As an antioxidant, it prevents oxidation of low-density lipoproteins and the expression of metalloproteinase 1, thus inhibiting the disruption of atherosclerotic plaques and contributing to plaque stabilization (**Song et al., 2001**) and also brings about the regeneration of the pancreatic islets and probably increases insulin release in streptozocin-induced diabetic rats; thus exerting its beneficial antidiabetic effects (**Vessal et al., 2003**).

It has been suggested as a potent anticarcinogenic flavonol. In 9,10-dimethyl-1,2benzanthracene-initiated and TPA-promoted two-stage mouse skin cancer models, quercetin exerted the anticarcinogenic effects (Soleas et al., 2006). In tumor cells, it exerts antiproliferative effects, arrests human leukemic T cells in late G1 phase of the cell cycle (Yoshida et al., 1992) and markedly inhibited the growth of human gastric cancer cells (Yoshida et al., 1990). Quercetin decreased expression of metalloproteinase-2 and metalloproteinase -9 in a dose-dependent manner in prostate cancer cells PC-3 *in vitro* (Vijayababu et al., 2006).

Quercetin has also antiinflammatory effects, regulating nitric oxide, interleukin-6, and tumor necrosis factor- α release (Comalada et al., 2005; Dias et al., 2005; Liu et al., 2005a; Manjeet and Glosh, 1999; Kim et al., 2007; Kumayawa et al., 2006), thereby alleviating oxidative damage in the tissue (Dias et al., 2005) and inhibiting the lipopolysaccharide-induced delay in spontaneous apoptosis and activation of neutrophils (Liu et al., 2005b). It is likely that quercetin may be inhibiting the synthesis of the hormone or interfering with hormone-receptor binding. It could also be blocking some primary events stimulated by the hormone-receptor interaction. Quercetin does appear to inhibit tyrosine protein kinase activity (Levy et al., 1984; Sharoni et al., 1986) what have been implicated as necessary for normal mammary growth and development (Levy et al., 1984; Sharoni et al., 1984; Sharo

It is examined that a single oral daily dose of the bioflavonoid quercetin reduced blood pressure and heart rate, the cardiac and renal hypertrophy, the endothelial dysfunction and the oxidant status in a rat model of spontaneous hypertension, but had no effect on normotensive rats. This report showed the chronic antihypertensive effect of a quercetin (Duarte et al., 2001).

Protective effect of quercetin against various diseases such as osteoporosis, certain forms of cancer, pulmonary and cardiovascular diseases but also against aging was also observed (**Boots et al., 2008**). Short-term, high intake of black currant and apple juices had a prooxidant effect on plasma proteins and increased glutathione peroxidase activity, whereas lipid oxidation in plasma seemed to decrease (**Young et al., 1999**). Quercetin (12,5-50 mg/kg) reduced the area of gastric ulcer but not the number. It is suggested that α 2-adrenergic receptors mediate the effect of quercetin on intestinal motility and secretion (**Carlo et al., 1994**).

SUMMARY, CONCLUSIONS, AND FUTURE PERSPECTIVES

Data from several studies suggest that quercetin has a broad range of activities within animal cells and occurs to be able to prevent or reduce the development of different type of diseases. The absorption and metabolism of quercetin is still poorly understood, but future studies can detect more facts connected with these processes.

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REFERENCES

BJELDANES, L.F. - CHANG, G.W. 1977. Mutagenic activity of quercetin and related compounds. In *Science*, vol. 197, 1977, p. 577-578.

BOOTS, A.W. - GUIDO, R.M.M. 2008. Haenen, Aalt Bast. Health effects of quercetin: From antioxidant to nutraceutical. In *European Journal of Pharmacology*, vol. 585, 2008, no. 2-3, p. 325-337.

CARLO, G.D. - MASCOLO, N. - IZZO, A.A. - CAPASSO, F. - AUTORE, G. 1994. Effects of quercetin on the gastrointestinal tract in rats and mice. In *Phytotherapy Research*, vol. 8, 1994, no. 1, p. 42-45.

COMALADA, M. - CAMUESCO, D. - SIERRA, S. - BALLESTER, I. - XAUS, J. -

GALVEZ, J. - ZARZUELO, A. 2005. *In vivo* quercitrin anti-inflammatory effect involves release of quercetin, which inhibits inflammation through down-regulation of the NF-kappaB pathway. In *European Journal of Immunology*, vol. 35, 2005, no. 2, p. 584-592.

DIAS, A.S. - PORAWSKI, M. - ALONSO, M. - MARRONI, N. - COLLADO, P.S. - GONZALEZ-GALLEGO, J. 2005. Quercetin decreases oxidative stress, NF-kappaB activation, and iNOS overexpression in liver of streptozotocin-induced diabetic rats. In *Journal of Nutrition*, vol. 135, 2005, no. 10, p. 2299-2304.

DREWA, G. - WOZQAK, A. - PALGAN, K. - SCHACHTSCHSBEL, D.O. - GRZANKA, A. - SUJIKOWSKA, R. 2001. Influence of quercetin on B16 melanotic melanoma growth in C57BL/6 mice and on activity of some acid hydrolases in melanoma tissue. In *Neoplasma*, vol. 48, 2001, no. 12-18.

DUARTE, J. - PÉREZ-PALENCIA, R. - VARGAS, F. - OCETE, M.A. - PÉREZ-VIZCAINO, F. - ZARZUELO, A. - TAMARGO, J. 2001. Antihypertensive effects of the flavonoid quercetin in spontaneously hypertensive rats. In *British Journal of Pharmacology*, vol. 133, 2001, no. 1, p. 117-124.

ERLUND, I. 2004. Review of the flavonoids quercetin, hesperetin, and naringenin: dietary sources, bioactivities, bioavailability, and epidemiology. In *Nutrition Research*, vol. 24, 2004, no. 10, p. 851-874.

GRAEFE, E.U. - WITTING, J. - MUELLER, S. - RIERHLING, A.K. - UEHLEKE, B. - DREWELOW, B. - PFORTE, H. - JACOBASCH, G. - DERENDORF, H. 2001. Pharmacokinetics and bioavailability of quercetin glycosides in humans. In *Journal of Clinical Pharmacology*, vol. 41, 2001, no. 5, p. 492-499.

HÄKKINEN, S.H. - KÄRENLAMPI, S.O. - HEINONEN, I.M. - MYKKÄNEN, H.M. - TÖRRÖNNEN, A.R. 1999. Content of the flavonols quercetin, myricetin, and kaempferol in 25 edible berries. In *Journal of Agricultural Food Chemistry*, vol. 47, 1999, no. 6, p. 2274-2279.

HERTOG, M.G.L. - HOLLMAN, P.C.H. - PUTTE, B. 1993. Flavonol and flavone content of beverages. In *Journal of Agricultural Food Chemistry*, vol. 41, 1993, p. 1242-1246.

HERTOG, M.G.L. - HOLLMAN, P.C.H. - KATAN, M.B. 1992. Content of potentially anticarcinogenic flavonoids of 28 vegetables and 9 fruits commonly used in the Netherlands. In *Journal of Agricultural Food Chemistry*, vol. 40, 1992, no. 12, p. 2379-2383.

HOLLMAN, P.C. - KATAN, M.B. 1997. Absorption, metabolism and health effects of dietary flavonoids in man. In *Biomedical Pharmacotheraphy*, vol. 51, 1997, no. 8, p. 305-310. HOLLMAN, P.C. - VRIES, J.H. - LEEUWEN, S.D. - MENGELERS, M.J. - KATAN, M.B.

1995. Absorption of dietary quercetin glycosides and quercetin in healthy ileostomy volunteers. In *Americal Journal of Clinical Nutrition*, vol. 62, 1995, no. 6, p. 1276-1282.

LEVY, J. - TEUERSTEIN, I. - MARBACH, M. - RADIAN, S. - SHARONI, Y. 1984. Tyrosine protein kinase activity in the DMBA-induced rat mammary tumor: inhibition by quercetin. In *Biochemical and Biophysical Research Communications*, vol. 123, 1984, no. 3, p. 1227-1233.

LIU, J. - LI, X. - YUE, Y. - LI, J. - He, T. - He, Y. 2005. The inhibitory effect of quercetin on IL-6 production by LPS-stimulated neutrophils. In *Cellular & Molecular Immunol*ogy, vol. 2, 2005, no. 6, p. 455-460.

LIU, J.J. - SONG, C.W. - YUE, Y. - DUAN, C.G. - YANG, J. - HE, T. - HE, Y.Z. 2005. Quercetin inhibits LPS-induced delay in spontaneous apoptosis and activation of neutrophils. In *Inflammation Research*, vol. 54, 2005, no. 12, p. 500-507.

MIDDLETON, E.J. 1998. Effect of plant flavonoids on immune and inflammatory cell function. In *Advances in Experimental Medicine and Biology*, vol. 439, 1998, p. 175-182.

MOSKUAG, J.O. 2004. Molecular Imaging of the biological effects of Quercetin and Quercetin-rich foods. In *Mechanism of Ageing and Development*, vol. 125, 2004, no. 4, p. 315-324.

OLTHOF, M.R. - HOLLMAN, P.C.H. - VREE, T.B. - KATAN, M.B. 2000. Bioavailabilities of quercetin-3-glucoside and quercetin-4-glucoside do not differ in humans. In *Journal of Nutrition*, vol. 130, 2000, no. 5, p. 200-203.

SHARONI, Y. - TEUERSTEIN, I. - LEVY, J. 1986. Phosphoinositide phosphorylation precedes growth in rat mammary tumors. Biochem. In *Biophys. Res. Commun.*, vol. 134, 1986, no. 2, p. 876-882.

SINGLETON, V. L. 1981. Naturally occurring food toxicants: phenolic substances of plant origin common in risk foods. In *Advances in Food Research*, vol. 27, 1981, p. 149-242.

SOLEAS, G.J. - GRASS, L. - JOSEPHY, P.D. - GOLDBERG, D.M. - DIAMANDIS, E.P. 2006. A comparison of the anticarcinogenic properties of four red wine polyphenols. In *Clinical Biochemistry*, vol. 35, 2006, no. 2, p. 492-497.

SONG, L. - XU, M. - LOPES-VIRELLA, M.F. - HUANG, Y. 2001. Quercetin inhibits matrix metalloproteinase-1 expression in human vascular endothelial cells through extracellular signal-regulated kinase. In *Arch Biochem Biophys.*, vol. 391, 2001, no. 1, p. 72-78.

UENO, I. - NAKANO, N. - HIRONO, I. 1982. Metabolic fate of [14C] quercetin in the ACI rat. In *The Japanese journal of experimental medicine*, vol. 53, 1982, no. 1, p. 41-50.

VESSAL, M. - HEMMATI, M. - VASEI, M. 2003. Antidiabetic effects of quercetin in

streptozocin-induced diabetic rats. In *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*, vol. 135, 2003, no. 3, p. 357-364.

VIJAYABABU, M.R. - ARUNKUMAR, A. - KANAGARAJ, P. - VENKATAMARAN, P. - KRISHAMOORTHY, G. – ARUNAKARAN, J. 2006. Quercetin downregulates matrix metalloproteinases 2 and 9 proteins expression in prostate cancer cells (PC-3). In *Molecular and Cellular Biochemistry*, vol. 287, 2006, no. 1-2, p. 109-116.

WICZKOWSKI, W. - ROMASZKO, J. - BUCINSKI, A. - SZAWARA-NOWAK, D. -HONKE, J. - ZIELINSKY, H. - PISKULA, M.K. 2008. Quercetin from Shallots (Allium cepa L. var. aggregatum) Is More Bioavailable Than Its Glucosides 1–3. In *The Journal of Nutrition Nutrient Physiology, Metabolism, and Nutrient-Nutrient Interactions*, vol. 138, 2008, no. 5, p. 885-888.

YOSHIDA, M. - YAMAMOTO, M. - NIKAIDO, T. 1992. Quercetin arrests human leukemic T-cells in late G1 phase of the cell cycle. In *Cancer Research*, vol. 52, 1992, no. 23, p. 6676-6681.

YOSHIDA, M. - TOSHIYUKI, S. - NOBUKO, H. - NOBUYUKI, M. - MATSUMOTO, K. - FUJIOKA, A. - NISHINO, H. - AOIKE, A. 1990. The effect of quercetin on cell cycle progression and growth of human gastric cancer cells. In *FEBS Letters*, vol. 260, 1990, no.1, p. 10-13.

YOUNG, J.F. - NIELSEN, S.E. - HARALDSDÓTTIR, J. - DANESHVAR, B. - LAURIDSEN, S.T. - KNUTHSEN, P. - CROZIER, A. - SANDSTRÖM, B. - DRAGSTED, L.O. 1999. Effect of fruit juice intake on urinary quercetin excretion and biomarkers of antioxidative status. In *American Journal of Clinical Nutrition*, vol. 69, 1999, no. 1, p. 87-94.