



## CYTOTOXIC AND ANTIOXIDANT ACTIVITY OF BUCKWHEAT HULL EXTRACTS

*Martina Danihelová\*, Soňa Jantová, Ernest Šturdík*

**Address:** Mgr. Martina Danihelová, Slovak University of Technology in Bratislava, Faculty of Chemical and Food Technology, Institute of biochemistry, nutrition and health protection, Radlinského 9, 812 37 Bratislava, Slovak Republic.

\*Corresponding author: [martina.danihelova@stuba.sk](mailto:martina.danihelova@stuba.sk)

---

### ABSTRACT

Buckwheat contains many prophylactic compounds that are concentrated mainly in outer layers of buckwheat grain. The aim of this study was to prepare buckwheat hull extracts. Ten buckwheat cultivars were screened for their antioxidant and anticancer properties. Total polyphenol content was determined using Folin-Ciocalteu's reagent. Antioxidant activity was established by the method of binding free radical DPPH. Cytotoxic properties were measured on human cervical cancer cells HeLa using mitochondrial cytotoxic test (MTT). Total polyphenol content ranged from 166.67 to 635.31 mg GAE/100 g DW. The highest content displayed tartary buckwheat cultivar *Madawaska* (0.64% of hulls weight). Among common buckwheat the richest in polyphenols were cultivars *Bamby* and *KASHO-2*. The best free radical binding antioxidant activity was found for cultivars with highest polyphenol content. This relationship was not observed for cytotoxic action on human cervical cancer cells. The best growth inhibitory activity on HeLa cancer cells displayed common buckwheat cultivars *Bamby* and *KASHO-2* (up to 50%, extract concentration 100 µg/ml). This was not found for tartary buckwheat cultivar *Madawaska*.

**Keywords:** antioxidant activity, buckwheat hull, cytotoxic effect, polyphenol content

---

## INTRODUCTION

Buckwheat represents a raw material interesting in term of its nutritional and health beneficial suitability. Buckwheat grain is a source of valuable proteins, starch with low glycemic index or high amount of unsaturated fatty acids. It contains compounds with prophylactic value, too. Buckwheat is one of the richest sources of polyphenols and flavonoids. These are concentrated mainly in outer layers of buckwheat seed (**Krkošková and Mrázová, 2005**). Among them the most abundant is rutin with its content 0.02% to 2% (**Jiang et al., 2007**).

Data obtained from laboratory and epidemiological experiments indicate that buckwheat can play an important role in cancer prevention. Buckwheat extracts effectively inhibited the growth of lung, hepatic, colon, breast, gastric or cervical cancer cells (**Chan, 2003; Kim et al., 2007**). Buckwheat flavonoids were able to induce apoptosis in human leukemic cells HL-60 (**Ren et al., 2003**). Common buckwheat extracts exhibited antimutagenic action in Ames testing (**Brindzová et al., 2009**). Among other mechanisms of action antioxidant and antiproliferative properties are important (**Cao et al., 2008; Guo et al., 2010**).

In our study nine common buckwheat cultivars and one tartary buckwheat cultivar were screened for their anticancer properties. There was determined total polyphenol content, the ability to scavenge free radicals and cytotoxic effect on human cervical cancer cells HeLa. Correlation between polyphenol content and observed beneficial properties was evaluated.

## MATERIAL AND METHODS

Nine common buckwheat cultivars and one tartary buckwheat cultivar were kindly provided from Plant production research center in Piešťany. Overview of tested cultivars is outlined in table 1. Buckwheat grains were mechanically dehulled. Obtained hulls were extracted using methanol (p. a.) for 24 hours at room temperature, filtered and used for polyphenol content determination and antioxidant activity testing. For purposes of cytotoxicity evaluation extracts were evaporated and dissolved in dimethylsulfoxide.

**Table 1** Overview of tested buckwheat cultivars

<b>Buckwheat cultivar</b>	<b>Buckwheat variety</b>	<b>Crop year</b>
<i>Pyra</i>	Common buckwheat	2011
<i>Špačinská 1</i>	Common buckwheat	2011
<i>Siva</i>	Common buckwheat	2011
<i>Emka</i>	Common buckwheat	2011
<i>Bamby</i>	Common buckwheat	2011
<i>Aiva</i>	Common buckwheat	2011
<i>Madawaska</i>	Tartary buckwheat	2011
<i>KASHO-2</i>	Common buckwheat	2011
<i>JANA CI</i>	Common buckwheat	2011
<i>Hrusowska</i>	Common buckwheat	2011

### Total polyphenol content

The content of polyphenols was determined using Folin-Ciocalteu's colorimetric method (**Singleton and Rossi, 1965**). 500 µl of Folin-Ciocalteu's reagent was added to 100 µl of sample. After 3 minutes, 1.5 ml of 20% Na<sub>2</sub>CO<sub>3</sub> was pipetted to the reaction mixture and allowed to react for 2 hours at room temperature. The absorbance was measured at 765 nm using UV-visible spectrophotometer (UV-1700, Shimadzu, China). Samples were measured in three replicates. Standard curve of gallic acid was prepared using the similar procedure. Results were expressed in gallic acid equivalents (mg GAE/g dry sample).

### Free radical scavenging activity

For determination of the ability to scavenge free radicals there was used simple spectrophotometric method according **Yen and Chen (1995)**. In this method 12 mg of 2,2'-diphenyl-1-picrylhydrazyl (DPPH) were dissolved in 100 ml of methanol. 50 µl of sample, 100 µl of methanol and 100 µl of DPPH solution were pipetted into microplate well. The reaction of samples with DPPH solution lasted for 10 min at room temperature in dark. After that, the absorbance changes were measured using microplate reader (Multiskan FC, Thermo Scientific, China). Samples were measured in five replicates. Trolox was used as standard antioxidant control. Results were expressed in trolox equivalents (TE/g dry sample).

### Cytotoxic activity (MTT test)

Cytotoxic effect of extracts to cancer cells was detected *in vitro* using mitochondrial cytotoxic test according **Theiszová et al. (2005)** with modifications. Cell viability was evaluated using thiazolyl blue tetrazolium bromide (MTT) which measures the metabolic activity of cells. Experiment was performed in 96-well microplates. The cells were seeded at a density of  $3.5 \times 10^3$  HeLa cells per well. Samples were diluted in medium to the final concentration of 12.5 µg/ml to 100 µg/ml and after 24 h added to cells. Microplates were cultivated for 72 hours in thermostat at 37 °C and 5% CO<sub>2</sub>. After incubation 30 µl of thiazolyl blue tetrazolium bromide (3.33 mg/ml phosphate buffered saline, pH = 7.4) was pipetted to each well and left to incubate for another two hours. After that, medium with MTT solution was removed. Formazan crystals in viable cells were dissolved in 150 µl of lysis solution (4 mM HCl and 0.1% Nonidet P40 in ethanol). Microplates were shaken 15 min at 1500 rpm. Absorbance was measured at 540 nm and reference wavelength 740 nm. Samples were measured in six replicates. Inhibition activity was expressed as percentages of control with DMSO.

## RESULTS AND DISCUSSION

### Total polyphenol content and antioxidant activity determination

Buckwheat is nutritionally valuable food source. Additional value lies in the presence of many prophylactic components. Phytosterols, group B vitamins or minerals have a part in managing of many diseases (**Danihelová and Šturdík, 2012**). Among them the most abundant are polyphenols, especially the group of flavonoids. The main and content-dominant flavonoid is rutin. Except of rutin there are present other flavonoids (quercetin, kaempferol, luteolin, catechin,...) and phenolic acids (caffeic acid, syringic acid, chlorogenic acid,...) (**Alvarez-Jubete et al., 2010**).

One of the best described properties of polyphenols is their ability to act as antioxidants. Beneficial effects for organisms are attributed to inhibition of lipid peroxidation, binding metals, reactive oxygen and nitrogen species or free radicals. Through their positive action they can lower oxidative damage of DNA, proteins, lipids and other important biomolecules (**Rice-Evans et al., 1997**).

Because present antioxidant polyphenols are mainly concentrated in outer layers of buckwheat seed and buckwheat hulls represent waste material that has no important commercial utilization, buckwheat hulls from ten cultivars were used for their screening of beneficial compounds and activities. In prepared methanolic hull extracts there was assessed total polyphenol content using simple spectrophotometric assay. Results are expressed in mg GAE/100 g of dry sample. Along with this the antioxidant capacity was evaluated through monitoring the ability of samples to scavenge free radical DPPH. Inhibition activities were compared to standard antioxidant trolox. Obtained data are presented in table 2.

**Table 2** Total polyphenol content in ten buckwheat cultivars as determined using Folin-Ciocalteau's reagent and their antioxidant activity assessed via binding free radical DPPH

Buckwheat cultivar	TP (mg GAE/100 g DW)	DPPH ( $\mu$ M TE/ g DW)
<i>Pyra</i>	168.75 $\pm$ 8.38	20.24 $\pm$ 0.67
<i>Špačinská 1</i>	217.39 $\pm$ 10.09	25.34 $\pm$ 0.91
<i>Siva</i>	187.64 $\pm$ 2.45	19.32 $\pm$ 0.28
<i>Emka</i>	193.69 $\pm$ 9.03	22.94 $\pm$ 1.11
<i>Bamby</i>	406.03 $\pm$ 18.64	471.35 $\pm$ 16.12
<i>Aiva</i>	177.47 $\pm$ 8.28	20.69 $\pm$ 0.44
<i>Madawaska</i>	635.31 $\pm$ 23.13	1038.67 $\pm$ 4.23
<i>KASHO-2</i>	361.39 $\pm$ 18.26	452.38 $\pm$ 5.79
<i>JANA C1</i>	166.67 $\pm$ 4.36	21.08 $\pm$ 0.75
<i>Hrusowska</i>	177.44 $\pm$ 8.78	21.22 $\pm$ 0.78

Detected polyphenol content in buckwheat hull samples was in the range from 166.67 to 635.31 mg GAE/100 g of dry hulls weight. The highest content of polyphenols was found in the only tested cultivar of tartary buckwheat *Madawaska* (about 0.64% of hulls weight). Among common buckwheat the richest in polyphenols were cultivars *Bamby* (0.41%) and *KASHO-2* (0.36%). Most of common buckwheat samples achieved polyphenol content values about 0.2% of hulls weight, what represents one third of polyphenols present in examined tartary buckwheat sample.

Our findings are in accordance with informations from the literature. **Sharma et al. (2012)** published that tartary buckwheat samples expressed higher total phenolic content compared to the common buckwheat. Tartary buckwheat groats have been shown to contain two times higher amount of total phenolics than common buckwheat (**Cao et al., 2008**). Differences in polyphenol content between buckwheat varieties can be much higher because of varietal and environmental influences.

Buckwheat samples with high content of polyphenols achieved the highest ability to scavenge free radicals. We have found a good correlation between polyphenol content and

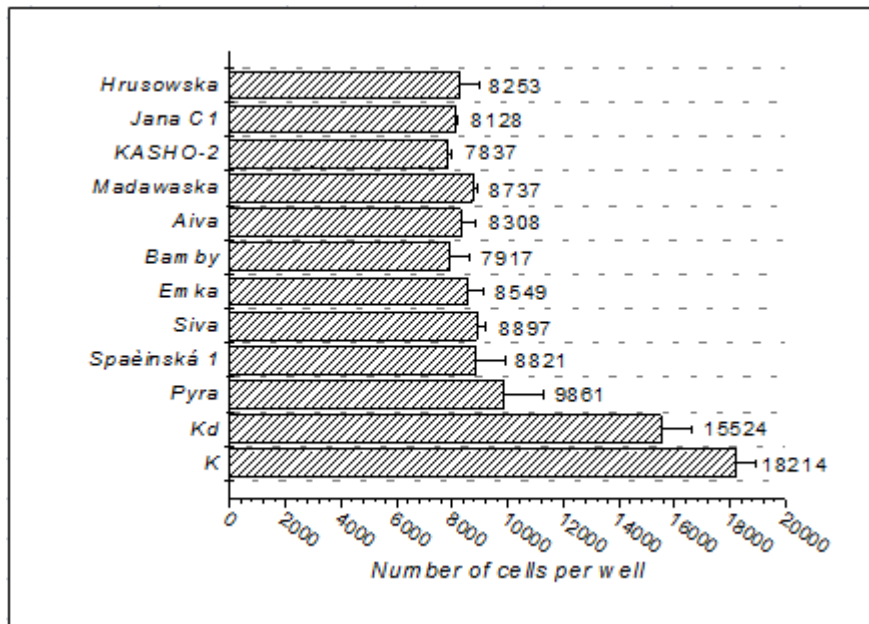
antioxidant activity in tested buckwheat cultivars ( $R^2 = 0.9889$ ). Although tartary buckwheat was three times higher in phenolics than common buckwheat, its antioxidant activity as determined by binding free radical DPPH exhibited even fifty times better value. The best antioxidant effect in common buckwheat was determined for cultivars *Bamby* and *KASHO-2*. Their activity was about twenty-five times higher in comparison with other common buckwheat cultivars.

Positive relationship between the present polyphenolic compounds and their antioxidant activity has been described by many authors (**Zieliński and Kozłowska, 2000; Gorinstein et al., 2007; Kishore et al., 2010**). Rutin appears to be the major antioxidant in buckwheat (**Morishita et al., 2007**). Tartary buckwheat cultivars display several times higher antioxidant activity than the common one (**Morishita et al., 2007; Cao et al., 2008**).

### **Cytotoxicity evaluation**

Performed studies indicate that buckwheat possess anticancer properties. However this literature is quite scarce. Available informations document that common and tartary buckwheat extracts induce apoptosis (**Ren et al., 2003**) and have antimutagenic action (**Brindzová et al., 2009**). They are effective in growth inhibition of various cancer cells (**Chan, 2003; Zheng et al., 2012**).

Ten buckwheat cultivars were tested for their ability to inhibit growth of human cervical cancer cell line HeLa. Cell viability was detected *in vitro* using mitochondrial cytotoxic test. Viable cells convert yellow tetrazolium salt to purple formazan. Colour changes are measured spectrophotometrically. Samples were tested at the concentration of 100 µg extract/ml. Results are shown in figure 1.



**Figure 1** Antiproliferative activity of ten buckwheat cultivars on HeLa cancer cell line as measured by MTT test (100 µg extract/ml, 72 h)

Obtained results indicate that buckwheat hull extracts at the concentration of 100 µg/ml are able to inhibit the growth of human cervical cancer cells. After 72 hours of exposure there were observed inhibitory activities in the range from 36.5% to 49.5%. Majority of samples displayed similar inhibitory effect. The best results have reached common buckwheat cultivars *Bamby* and *KASHO-2*. Among common buckwheat these two have shown to contain the highest polyphenol content. In contrary tartary buckwheat cultivar *Madawaska* containing from tested samples the most polyphenols didn't exhibit high antiproliferative effect.

Data from the literature describe for buckwheat extracts potent cytotoxic effect against various cancer cells. **Kim et al. (2007)** published that buckwheat hulls effectively inhibited the growth of breast, hepatic, lung, gastric and cervical cancer cells. They observed inhibitory action up to 93%, but they used higher extract concentrations (250 µg/ml to 1000 µg/ml). Tartary buckwheat flavonoids induced apoptosis in human leukemic cells (**Ren et al., 2003**). Tartary buckwheat was active against skin cancer cells (**Park and Park, 2004**). Authors observed slight cytotoxic activity.

## CONCLUSION

Buckwheat material provides nutritional and health benefits. Many prophylactic compounds such as polyphenols and flavonoids are concentrated in outer layers of buckwheat

grain. In our study there were screened 9 common buckwheat cultivars and 1 tartary buckwheat cultivar. Tartary buckwheat hulls were more abundant in polyphenols than common buckwheat. Cultivars rich in polyphenols displayed higher antioxidant activity. The highest cytotoxic action on human cervical cancer cell line HeLa have shown hulls of common buckwheat cultivars *Bamby* and *KASHO-2*. Obtained results indicate that buckwheat has potential in prevention of cancer diseases.

**Acknowledgments:** This work was financially supported by The Agency of the Ministry of Education, Science, Research and Sport of the Slovak Republic for the Structural Funds of EU (ITMS 26240220040). We are grateful to Plant Production Research Center Piešťany for providing buckwheat samples.

## REFERENCES

- ALVAREZ-JUBETE, L. - WIJNGAARD, H. - ARENDT, E. K. - GALLAGHER, E. 2010. Polyphenol composition and in vitro antioxidant activity of amaranth, quinoa, buckwheat and wheat as affected by sprouting and baking. In *Food chemistry*, vol. 119, 2010, no. 2, p. 770-778.
- BRINDZOVÁ, L. – MIKULÁŠOVÁ, M. – TAKÁCSOVÁ, M. – MOŠOVSKÁ, S. – OPATTOVÁ, A. 2009. Evaluation of the mutagenicity and antimutagenicity of extracts from oat, buckwheat and wheat bran in the *Salmonella*/microsome assay. In *Journal of food composition and analysis*, vol. 22, 2009, no. 1, p. 87-90.
- CAO, W. – CHEN, W.-J. – SUO, Z.-R. – YAO, Y.-P. 2008. Protective effects of ethanolic extracts of buckwheat groats on DNA damage caused by hydroxyl radicals. In *Food research international*, vol. 41, 2008, no. 9, p. 924-929.
- CHAN, P.-K. 2003. Inhibition of tumor growth *in vitro* by the extract of *fagopyrum cymosum* (fago-c). In *Life sciences*, vol. 72, 2003, no. 16, p. 1851-1858.
- DANIHELOVÁ, M. - ŠTURDÍK, E. 2012. Nutritional and health benefits of buckwheat. In *Potravinárstvo*, vol. 6, 2012, no. 3, p. 1-9.
- GORINSTEIN, S. – VARGAS, O. J. M. – JARAMILLO, N. O. – SALAS, I. A. – AYALA, A. L. M. – ARANCIBIA-AVILA, P. – TOLEDO, F. – KATRICH, E. – TRAKHTENBERG, S. 2007. The total polyphenols and the antioxidant potentials of some selected cereals and pseudocereals. In *European food research and technology*, vol. 225, 2007, no. 3-4, p. 321-328.



- GUO, X. – ZHU, K. – ZHANG, H. – YAO, H. 2010. Antitumor activity of a novel protein from tartary buckwheat. In *International journal of molecular sciences*, vol. 11, 2010, no. 12, p. 5201-5211.
- JIANG, P. – BURCZYNSKI, F. – CAMPBELL, C. – PIERCE, G. – AUSTRIA, J. A. – BRIGGS, C. J. 2007. Rutin and flavonoid contents in three buckwheat species *Fagopyrum esculentum*, *F. tataricum*, and *F. homotropicum* and their protective effects against lipid peroxidation. In *Food research international*, vol. 40, 2007, no. 3, p. 356-364.
- KIM, S. H. – CUI, C. B. – KANG, I. J. – KIM, S. Y. – HAM, S. S. 2007. Cytotoxic effect of buckwheat (*Fagopyrum esculentum* Moench) hull against cancer cells. In *Journal of medicinal food*, vol. 10, 2007, no. 2, p. 232-238.
- KISHORE, G. – RANJAN, S. – PANDEY, A. – GUPTA, S. 2010. Influence of altitudinal variation on the antioxidant potential of tartar buckwheat of Western Himalaya. In *Food science and biotechnology*, vol. 19, 2010, no. 5, p. 1355-1363.
- KRKOŠKOVÁ, B. – MRÁZOVÁ, Z. 2005. Prophylactic components of buckwheat. In *Food research international*, vol. 38, 2005, no. 5, p. 561-568.
- MORISHITA, T. – YAMAGUCHI, H. – DEGI, K. 2007. The contribution of polyphenols to antioxidative activity in common buckwheat and tartary buckwheat grain. In *Plant production science*, vol. 10, 2007, no. 1, p. 99-104.
- PARK, B. J. – PARK, C. H. 2004. Cytotoxic activities of tartary buckwheat against human cancer cells. In *Proceedings of the 9th International symposium on buckwheat*, Praha, 2004, p. 665-668.
- REN, W. – QIAO, Z. – WANG, H. – ZHU, L. – ZHANG, L. – LU, Y. – ZHANG, Z. – WANG, Z. 2003. Molecular basis of Fas and cytochrome c pathways of apoptosis induced by tartary buckwheat flavonoid in HL-60 cells. In *Methods and findings in experimental and clinical pharmacology*, vol. 25, 2003, no. 6, p. 431-436.
- RICE-EVANS, C. - MILLER, N. - PAGANGA, G. 1997. Antioxidant properties of phenolic compounds. In *Trends in plant science*, vol. 2, 1997, no. 4, p. 152-159.
- SHARMA, P. – GHIMERAY, A. K. – GURUNG, A. – JIN, C. W. – RHO, H. S. – CHO, D. H. 2012. Phenolic contents, antioxidant and  $\alpha$ -glucosidase inhibition properties of Nepalese strain buckwheat vegetables. In *African journal of biotechnology*, vol. 11, 2012, no. 1, p. 184-190.
- SINGLETON, V. L. – ROSSI, J. A. 1965. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. In *American journal of enology and viticulture*, vol. 16, 1965, no. 3, p. 144-158.

- THEISZOVÁ, M. - JANTOVÁ, S. - DRAGÚŇOVÁ, J. - GRZNÁROVÁ, P. - PALOU, M. 2005. Comparison the cytotoxicity of hydroxyapatite measured by direct cell counting and MTT test in murine fibroblast NIH-3T3 cells. In *Biomedical papers*, vol. 149, 2005, no. 2, p. 393-396.
- YEN, G. C. – CHEN, H. Y. 1995. Antioxidant activity of various tea extracts in relation to their antimutagenicity. In *Journal of agricultural and food chemistry*, vol. 43, 1995, no. 1, p. 27-32.
- ZHENG, C. – HU, C. – MA, X. – PENG, C. – ZHANG, H. – QIN, L. 2012. Cytotoxic phenylpropanoid glycosides from *Fagopyrum tataricum* (L.) Gaertn. In *Food chemistry*, vol. 132, 2012, no. 1, p. 433-438.
- ZIELIŃSKI, H. – KOZŁOWSKA, H. 2000. Antioxidant activity and total phenolics in selected cereal grains and their different morfological fractions. In *Journal of agricultural and food chemistry*, vol. 48, 2000, no. 6, p. 2008-2016.