



THE TOXIC EFFECT OF 4-NONYLPHENOL ON MALE REPRODUCTIVE SYSTEM

*Jana Lukáčová**, Zuzana Kňazická, Eva Tvrdá, Peter Massányi, Norbert Lukáč

Address: Slovak University of Agriculture, Faculty of Biotechnology and Food Sciences, Department of Animal Physiology, Trieda A. Hlinku 2, 949 76 Nitra, Slovak Republic, phone number: +421-37-6414288

*Corresponding author: jana312@gmail.com

ABSTRACT

Reproductive system influences many environmental toxicants that present the serious risk to the reproduction of human and animals, including hormone-like compounds called endocrine disrupting chemicals (EDCs). The interest about EDCs increased during the last 50 years due to their interference with the development and functioning of the endocrine system in aquatic animals. One of these toxic contaminants is nonylphenol that has the negative impact on the reproductive parameters not only on fish, but also on amphibians and mammals. Nonylphenol can decrease male fertility following a disruption of testicular development, a damage of Sertoli and Leydig cells and a reduction of a count and motility of spermatozoa.

Keywords: endocrine disruptors, nonylphenol, Sertoli cells, Leydig cells, spermatozoa, toxicity

INTRODUCTION

The endocrine system is one of the body's major homeostatic control system whose aim is to maintain normal functions and development in the face of constantly changing environment. Working in tandem with the nervous system, which mainly is responsible for rapid and immediate responses, the approximately 30 different glands comprising the

endocrine system tend to act in a slower and more sustained manner to regulate processes as diverse as the female reproductive cycle, bone growth, cell proliferation, and psychosocial behavior. Like all homeostatic control systems, the capacity to maintain physiological parameters within normal bounds is finite, and when this capacity is exceeded by chemical exposures, drugs, or environmental stressors, adverse consequences can ensue (**Marty et al., 2011**).

Endocrine disrupting chemicals (EDCs) or endocrine disruptors are compounds that can interfere with and alter the homeostasis of the endocrine system, by modulating its response, resulting in long-term adverse effects on human and animal health or their progeny. The effects also extend to the thyroid, nervous, immune system and metabolism in general (**Hotchkiss et al., 2008**). EDCs can be divided on the natural hormones normally occurring in humans or animals and plant derived metabolites. On the contrary, the second group includes various exogenous man-made substances such as synthetic hormones and drugs, industrial chemicals (organochloride pesticides, dioxins, polychloro biphenils (PCBs), polybromo biphenils (PBBs), and alkylphenols, parabens found in cosmetics and other personal care products), manufactured industrial products such as plastic additives (bisphenol A, phthalates), antifouling paints, and chemicals used in farm animal production (**Pinto et al., 2004; Diamanti-Kandarakis et al., 2009**).

The endocrine reproductive effects of these chemicals are believed to be due to their ability to disrupt the synthesis and metabolism of endogenous hormones, disrupt the synthesis of hormone receptors, mimic the effect of endogenous hormones or antagonize the effect of endogenous hormones, by altering the hormonal and homeostatic systems (**Mueller, 2004**).

The biological actions of hormones, including estrogens, androgens, progesterone, thyroxine and the neurosteroids pregnenolone and dehydroepiandrosterone (DHEA), are mediated via high affinity protein receptors within the target cells. All steroids are fat-soluble and readily cross the cell membrane, interacting with dimeric receptor proteins; in the case of estrogens these are ER- α and ER- β , although recent evidence suggests a third class, putative-ER or ER- γ , is involved in fish and possibly mammals (**Dogde et al., 1996; McLachlan et al., 2001**).

The affinity of steroid for its receptor is so great that it is the equivalent of being able to taste a teaspoonful of sugar dissolved in the water of a swimming pool. The steroid-receptor complex binds to target regions of DNA termed “response elements”. This activates the cascade of reactions, which are the response to the presence of steroids (**Laws et al., 1995; Gray et al., 1999; Stroheker et al., 2004**). Impairment of endocrine functioning by

exogenous compounds affects predominantly the estrogen, androgen and thyroid receptors (Mueller, 2004).

Estrogenic effects

The principal mechanism for the effect of EDs on the reproductive system may be mediated through binding the estrogen receptor or androgen receptor, and mimicking the effects of estrogen and antagonizing the effects of androgen, which disrupts the normal effects of the sex hormones (Kelce *et al.*, 1998; Sonnenschein and Soto, 1998).

Estrogen is a hormone that influences the development and maintenance of female sex characteristics, and the maturation and function of accessory sex organs. Estrogen is also produced in males and affects reproduction (Alberts *et al.*, 1983; Hess *et al.*, 1997). In males, it is present low concentration in blood but is present at extraordinarily high levels in semen. Estrogenic effects are mediated by 2 distinct nuclear receptors, estrogen receptor α (ER α) and ER β . The receptors are encoded by 2 different genes and are expressed in germ cells (O'Donnell *et al.*, 2001). ER α is present in postacrosomal, midpiece and tail regions and ER β in midpiece and tail regions and mitochondria of ejaculated human sperm (Durke *et al.*, 1998; Aquila *et al.*, 2004; Solakidi *et al.*, 2005). Incubation of sperm with 17- β -estradiol affects capacitation, acrosomal exocytosis, and fertilizing ability (Adeoya-Osigawa and Fraser, 2003).

Alkylphenols

A well known class of environmental endocrine disruptors is the group of compounds known as alkylphenolpolyethoxylates (APEs), which are non-ionic surfactants, consisting of branched-chain alkylphenol which has been reacted with ethylene oxide, producing an ethoxylate chain (White *et al.*, 1994). They are used in variety of products, including detergents, emulsifiers, pesticides, herbicides, paints, cosmetics, plastic ware (Nimrod and Benson, 1996) and intra-vaginal spermicides (Bolt *et al.*, 2001). APEs undergo a biodegradation process to produce short side chain derivatives such as nonylphenol, octylphenol and butylphenol in anaerobic conditions in water (Montgomery-Brown and Reinhard, 2003).

The estrogenicity of alkylphenols has been attributed to ability these chemicals have to mimic natural estrogen 17 β -estradiol (E2) in binding to and activating the estrogen receptor

(Routledge and Sumpter, 1997). Most alkylphenols are able to bind to the estrogen receptors in a concentration-dependent manner and thus potentially act as activators (agonists) or inhibitors (antagonists) of cellular responses mediated through the estrogen receptor. The binding activity of alkylphenols is associated with the presence of and the size (length) of substituted alkyl group **(Tabira et al., 1999)**. The hydrophobicity of organic compounds increases with alkylation **(Hansch et al., 1973)** and the increase in binding affinity of the alkylphenols is proposed to be partially caused by stronger hydrophobic interactions with the ligand-binding domain of the ER **(Routledge and Sumpter, 1997; Tabira et al., 1999; Hu and Azaiwa, 2003)**.

Nonylphenol

Nonylphenol (NP) is the major degradation product of alkylphenol polyethoxylates **(Junk et al., 1974)** and is one of the most common chemical contaminants. NP is present in ambient air, water, soil, sediments and biota **(Ying et al., 2002)**. It is widely used as detergents, emulsifiers and surfactants in industrial and household products such as paints, plastics, cosmetics, construction materials, vulcanized rubber and paper **(USEPA, 2005)**. NP can stay biologically active for a longer period of time in the body than endogenous estrogens **(Nimrod and Benson, 1996)** and can interfere with reproduction in fish, reptiles and mammals, and induces the cell death in gonads and changes to other reproductive parameters **(Nagao et al., 2001; Weber et al., 2002; Cardinali et al., 2004)**.

Effect of nonylphenol on male reproductive system

There is an increasing concern that environmental contaminants affect the male reproduction of humans and wildlife. Male reproductive abnormalities may be due to the increased exposure to environmental contaminants such as organochlorine pesticides, polychlorinated biphenyls, dioxins, phytoestrogens, alkylphenol ethoxyxylates and other xenoestrogens that enter humans through food, drinking water, air and skin contact **(Toppari et al., 1996)**.

Effect of nonylphenol on testis

The mammalian testis is a complex organ that serves two important functions: synthesis of steroids and production of spermatozoa that controlled by gonadotrophins and numerous locally synthesized factors (Saez, 1994) and among them oestrogens (Carreau et al., 1999).

NP profoundly impairs testicular function as evidenced by reduced testis size (Chitra et al., 2002), low circulating testosterone, disturbed testicular structure and suppressed spermatogenesis (Nagao et al., 2001; Tan et al., 2003; Cardinali et al., 2004). NP influences an increasing incidence of testicular cancer, an increase in occurrence of cryptorchidism and hypospadias during the past decades (Carlsen et al., 1995; Toppari et al., 1996). Various environmental contaminants can induce oxidative stress by generating reactive oxygen species (ROS) such as hydrogen peroxide (H_2O_2) and superoxide anion (O_2^-) (Chitra et al., 2003; Wang et al., 2003). Low micromolar concentrations of NP showed to induce testicular oxidative stress and cytotoxicity *in vitro* (Gong and Han, 2006). NP administration increased ROS level and lipid peroxidation and depressed the activity of antioxidant enzymes such as superoxide dismutase and glutathione reductase in rat testis (Chitra and Mathur, 2004).

Effect of nonylphenol on epididymis

The epididymis is known to play an important role in providing the microenvironment for sperm maturation and storage of sperm (Cornwall, 2009; Raymond et al., 2010). Numerous proteins synthesized and secreted by the epididymal epithelia cells are thought to be considerable members of epididymal microenvironment and be involved in male reproductive activities including the initiation of sperm maturation, sperm-oocyte recognition, the acrosome reaction directly or indirectly (Yenuqu et al., 2006), interaction with *zona pellucida* and binding to and fuse with plasma membrane of the oocyte (Cuasnicu et al., 2002). All these modifications necessitate a minimum period in which they can occur, a time in which the spermatozoa must stay in the epididymal caput and corpus (Franca et al., 2005).

A study of Uguz et al. (2009) proved that NP in high concentrations negatively influences motility, mitochondrial membrane potencial and chromatin integrity of epididymal rat spermatozoa.

Effect of nonylphenol on Leydig cells and steroidogenesis

The testicular Leydig cells are the predominant source of the male sex steroid hormone testosterone, express estrogen receptors (ERs) and are subject to regulation by estrogen (Sherrill *et al.*, 2010). In mammalian species, Leydig cell development is regulated by the pituitary gonadotropin luteinizing hormone (LH) and by steroid hormones (i.e. androgen and estrogen) (Abney, 1999).

NP have the effect on testosterone biosynthesis (Gong and Han, 2006) and can inhibit hCG-induced testosterone release in rat Leydig cells (Wu *et al.*, 2010).

Effect of nonylphenol on Sertoli cells

The normal onset of spermatogenesis and the eventual production of a sufficient sperm number to insure fertility depends at least in part on Sertoli cells. They are the somatic cells that provide physical reinforcement to germ cells and mediate the movement of growth factors, hormones and signals into seminiferous tubules where spermatogenesis commences. Sertoli cells contribute to the consumption of excess byproducts after sperm cells have fully developed (Martincic *et al.*, 2001; Sinha and Swerdloff, 1999). Specific impairment of Sertoli cells will produce a parallel dysfunction in sperm production (Weber *et al.*, 2002).

The exposure to NP can induce hypertrophy of Sertoli cells (Miura *et al.*, 2005) and increase the levels of intracellular ROS and lipid peroxidation markers (Gong and Han, 2006), which may play an important role in the induction of apoptosis (Gong *et al.*, 2009).

Apoptosis occurs under the direction of an active, intracellular death program that can be stimulated or inhibited by environmental agents (Jacobson *et al.*, 1997). It is a form of cell death by a characteristic set of morphological and biochemical changes. Apoptosis is recognized as an early cellular inductor of toxicity (Weber *et al.*, 2002).

The endoplasmic reticulum, which is the site for folding and assembly of proteins, lipid biosynthesis, vesicular traffic, and cellular Ca²⁺ storage, is sensitive to alterations in homeostasis. Several stimuli, including alterations of Ca²⁺ homeostasis and accumulation of unfolded proteins in the endoplasmic reticulum, can cause oxidative stress of the endoplasmic reticulum and prolonged oxidative stress of the endoplasmic reticulum will lead to apoptosis (Kadowaki *et al.*, 2004; Gong and Han, 2006). NP can induce apoptosis in rat testicular cells, particularly in Sertoli cells by inhibiting the activity Ca²⁺ pump in the endoplasmic reticulum (Hughes *et al.*, 2000).

Effect of nonylphenol on spermatozoa

The mammalian spermatozoon is a deceptively simple, and terminally differentiated cell. A functional spermatozoon is composed of three main regions: the head, the midpiece and the tail. The sperm midpiece consists of a variable number of mitochondria wrapped helically around the anterior portion of the flagellum (**Ramalho-Santos et al., 2002**).

Mitochondria are important organelles in sperm homeostasis and the activity of sperm mitochondria is correlated with sperm motility, and thus with fertilization potential (**Gravance et al., 2000**). The role of these mitochondria might be provide ATP for sperm movement through oxidative phosphorylation. The necessary ATP is produced mostly by glycolytic pathways throughout the flagellum, with mitochondrial ATP required only in specific circumstances (**Ramalho-Santos et al., 2002**).

Mitochondria are the referential target of many toxic compounds (**Higgins and Rogers, 1974; Bragadin and Dell'Antone, 1996; Bragadin and Marton, 1997; Bragadin and Viola, 1997; Bragadin et al., 1998, 1999**) since a damage to mitochondria which synthesize ATP gives rise to a corresponding cell damage. The exposure to low-level doses of NP inhibits ATP synthesis in mitochondria (**Bragadin et al., 1999**).

CONCLUSION

The variety of man-made chemicals and their degradation products, on which the modern human life is dependent, can act as endocrine disruptors, enter humans and animals through food, drinking water, air and skin contact and so affect the health and reproduction. One of these chemicals is nonylphenol that can cause the abnormalities in male reproduction. Many authors confirm toxic ability of NP to disrupt a structure and development of testis and epididymis, decrease male fertility, including a decline of spermatozoa count, decreased spermatozoa motility parameters, production of oxidative stress and Sertoli cells apoptosis. Therefore it is very important the presence and the effects of nonylphenol on living organisms in the environment further monitor and investigate.

Acknowledgments: This work was supported by the Scientific Agency of the Slovak Republic VEGA No. 1/0532/11, KEGA No. 013SPU-4/2012 and KEGA No. 030SPU-4/2012.

REFERENCES

- ABNEY, T. O. 1999. The potential roles of estrogens in regulating Leydig cell development and function: A review. In *Steroids*, vol. 64, 1999, p. 610-617.
- ADEOYA-OSIGAWA, S. – FRASER, L. 2003. Calcitonin acts as a first messenger to regulate adenylyl cyclase/cAMP and mammalian sperm function. In *Molecular Reproduction and Development*, vol. 65, 2003, p. 228-236.
- ALBERTS, B. – BRAY, D. – LEWIS, J. – RAFF, M. – ROBERTS, K. – WATSON, J. D. 1983. *Molecular Biology of the Cell*. Garland Publishing Inc., New York.
- AQUILA, S. – SISI, D. – GENTILE, M. – MIDDEA, E. – CATALANO, S. – CARPINO, A. – RAGO, V. – ANDO, S. 2004. Estrogen receptor (ER) alpha and ER beta are both expressed in human ejaculated spermatozoa: evidence of their direct interaction with phosphatidylinositol-3-OH kinase/Akt pathway. In *The Journal of Clinical Endocrinology and Metabolism*, vol. 89, 2004, p. 1443-1451.
- BOLT, H. M. – JANNING, P. – MICHNA, H. 2001. Comparative assessment of endocrine modulators with oestrogenic activity: I. Definition of a hygiene-based margin of safety (HB MOS) for xeno-oestrogens against the background of European developments. In *Archives of Toxicology*, vol. 74, 2001, p. 649-662.
- BRAGADIN, M. – DELL'ANTONE, P. 1996. Mitochondrial bioenergetics as affected by cationic detergents. In *Archives of Environmental Contamination and Toxicology*, vol. 30, 1996, p. 280-284.
- BRAGADIN, M. – MARTON, D. 1997. A proposal for a new mechanism of interaction of trialkyltin (TAT) compounds with mitochondria. In *The Journal of Inorganic Biochemistry*, vol. 68, 1997, p. 75-78.
- BRAGADIN, M. – VIOLA, E. R. 1997. Ni⁺⁺ as a competitive inhibitor of calcium transport in mitochondria. In *The Journal of Inorganic Biochemistry*, vol. 66, 1997, p. 227-229.
- BRAGADIN, M. – MARTON, D. – MURGIA, M. – RIZZOLLI, V. – SCUTARI, G. – DEANA, R. 1998. Interactions of trialkyllead compounds with rat liver mitochondria. In *The Journal of Inorganic Biochemistry*, vol. 69, 1998, p. 259-262.
- BRAGADIN, M. – PERIN, G. – IERO, A. – MANENTE, S. – RIZZOLI, V. – SCUTARI, G. 1999. An in vitro study on the toxic effects of nonylphenols (NP) in mitochondria. In *Chemosphere*, vol. 38, 1999, p. 1997-2001.
- CARDINALI, M. – MARADONNA, F. – OLIVOTTO, I. – BORTOLUZZI, G. – MOSCONI, G. – POLZONETTI-MAGNI, A. M. – CARNEVALI, O. 2004. Temporary

impairment of reproduction in freshwater teleost exposed to nonylphenol. In *Reproductive toxicology*, vol. 18, 2004, p. 597-604.

CARLSEN, E. – GIWERCMAN, A. – KEIDING, N. – SKAKKEBAEK, N. E. 1995. Declining semen quality and increasing incidence of testicular cancer: is there a common cause? In *Environmental Health Sciences*, vol. 7, 1995, p. 137-139.

CARREAU, S. – GENISSEL, C. – BILINSKA, B. – LEVALLET, J. 1999. Sources of oestrogen in the testis and reproductive tract of the male. In *International Journal of Andrology*, vol. 22, 1999, p. 211-223.

CORNWALL, G. A. 2009. New insights into epididymal biology and function. In *Human Reproduction Update*, vol. 15, 2009, 213-227.

CUASNICU, P. S. – COHEN, D. J. – ELLERMAN, D. A. – BUSSO, D. – Da ROS, V. G. – MORGENFELD, M. M. 2002. *Changes in specific sperm proteins during epididymal maturation*. In Robaire B, Hinton BT (eds): *The Epididymis: From Molecules to Clinical Practice*. Kluwer Academic/Plenum Publisher, New York, p. 389-403.

DIAMANTI-KANDARAKIS, E. – BOURGUIGNON, J. P. – GIUDICE, L. C. – HAUSER, R. – PRINS, G. S. – SOTO, A. M. – ZOELLER, R. T. – GORE, A. C. 2009. Endocrine-disrupting chemicals: an Endocrine Society scientific statement. In *Endocrine reviews*, vol. 30, 2009, p. 293-342.

DODGE, J. A. – GLASEBROOK, A. L. – MAGEE, D. E. – PHILLIPS, D. L. – SATO, M. – SHORT, L. L. – BRYANT, H. U. 1996. Environmental estrogens: effects on cholesterol lowering and bone in the ovariectomized rat. In *The Journal of Steroid Biochemistry and Molecular Biology*, vol. 59, 1996, p. 155-161.

DURKE, T. – MUELLER, M. – ZINAMAN, M. 1998. Identification of estrogen receptor protein and messenger ribonucleic acid in human spermatozoa. In *American journal of obstetrics and gynecology*, vol. 178, 1998, p. 1288-1297.

EDDY, E. – O'BRIEN, D. 1994. *The spermatozoon*. In Knobil E, & Neill J, eds. *Physiology of Reproduction*. New York : Raven Press, 1994. p. 29-77.

FRANCA, L. R. – AVELAR, G. F. – ALMEIDA, F. F. L. 2005. Spermatogenesis and sperm transit through the epididymis in mammals with emphasis on pigs. In *Theriogenology*, vol. 63, 2005, p. 300-318.

GONG, Y. – HAN, X.D. 2006. Nonylphenol-induced oxidative stress and cytotoxicity in testicular Sertoli cells. In *Reproductive toxicology*, vol. 22, 2006, p. 623-630.

- GONG, Y. – WU, J. – HUANG, Y. – SHEN, S. – HAN, X. D. 2009. Nonylphenol induces apoptosis in rat testicular Sertoli cells via endoplasmic reticulum stress. In *Toxicology Letters*, vol. 186, 2009, p. 84-95.
- GRAVANCE, C. G. – GARNER, D. L. – BAUMBER, J. – BALL, B. A. 2000. Assessment of equine sperm mitochondrial function using JC-1. In *Theriogenology*, vol. 53, 2000, p. 1691-1703.
- GRAY, L. E. Jr – OSTBY, J. – MONOSSON, E. – KELCE, W. R. 1999. Environmental antiandrogens: low doses of the fungicide vinclozolin alter sexual differentiation of the male rat. In *Toxicology and Industrial Health*, vol. 15, 1999, p. 48-64.
- HANSCH, M. B. – LEO, A. – UNGER, S. H. – KIM, K. H. – NIKAITANI, D. – LIEN, E. J. 1973. "Aromatic" substituent constants for structure-activity correlations. In *Journal of medicinal chemistry*, vol. 16, 1973, p. 1207-1216.
- HOTCHKISS, A. K. – RIDER, C. V. – BLYSTONE, C. R. – WILSON, V. S. – HARTIG, P. C. – ANKLEY, G. T. – FOSTER, P. M. – GRAY, C. L. – GRAY, L. E. 2008. Fifteen years after "Wingspread"--environmental endocrine disrupters and human and wildlife health: where we are today and where we need to go. In *Toxicological Sciences*, vol. 105, 2008, p. 235-259.
- HESS, R. – BUNICK, D. – LEE, K. – BAHR, J. – TAYLOR, J. – KORACH, K. – LUBAHN, D. 1997. A role for oestrogens in the male reproductive system. In *Nature*, vol. 390, 1997, p. 509-512.
- HIGGINS, E. S. – ROGERS, K. S. 1974. Effects of cetyltrimethylammonium and dodecylnicotinamide halides on functional states of mitochondria. In *Proceedings of the Society for Experimental Biology and Medicine*, vol. 146, 1974, p. 85-91.
- HU, J.Y. – AIZAWA, T. 2003. Quantitative structure-activity relationships for estrogen receptor binding affinity of phenolic chemicals. In *Water research*, vol. 37, 2003, p. 1213-1222.
- HUGHES, P. J. – McLELLAN, H. – LOWES, D. A. – KHAN, S. Z. – BILMEN, J.G. – TOVEY, S. C. – GODFREY, R. E. – MICHELL, R. H. – KIRK, C. J. – MICHELANGELI, F. 2000. Estrogenic alkylphenols induce cell death by inhibiting testis endoplasmic reticulum Ca²⁺ pumps. In *Biochemical and biophysical research communications*, vol. 277, 2000, p. 568-574.
- CHITRA, K. C. – LATCHOUMYCANDANE, C. – MATHUR, P. P. 2002. Effect of nonylphenol on the antioxidant system in epididymal sperm of rats. In *Archives of toxicology*, vol. 76, 2002, p. 545-551.

- CHITRA, K. C. – LATCHOUMYCANDANE, C. – MATHUR, P. P. 2003. Induction of oxidative stress by bisphenol A in the epididymal sperm of rats. In *Toxicology*, vol. 185, 2003, p. 119-27.
- CHITRA, K. C. – MATHUR, P. P. 2004. Vitamin E prevents nonylphenol-induced oxidative stress in testis of rats. In *Indian Journal of Experimental Biology*, vol. 42, 2004, p. 220-223.
- JACOBSON, M. D. – WEIL, M. – RAFF, M. C. 1997. Programmed cell death in animal development. In *Cell*, vol. 88, 1997, p. 347-354.
- JUNK, G. A. – SVEC, H. J. – VICK, R. D. – AVERY, M. J. 1974. Contamination of water by synthetic polymer tubes. In *Environmental Science and Technology*, vol. 8, 1974, p. 1100–1106.
- KADOWAKI, H. – NISHITOH, H. – ICHIJO, H. 2004. Survival and apoptosis signals in ER stress: the role of protein kinases. In *Journal of chemical neuroanatomy*, vol. 28, 2004, p. 93-100.
- KELCE, W. R. – GRAY, L. E. – WILSON, E. M. 1998. Antiandrogens as environmental endocrine disruptors. In *Reproduction, Fertility and Development*, vol. 10, 1998, p. 105-111.
- LAWS, S. C. – CAREY, S. A. – KELCE, W. R. 1995. Differential effects of environmental toxicants on steroid receptor binding [Abstract]. In *Toxicologist*, vol. 15, 1995, p. 294.
- MARTINCIC, D. S. – VIRANT, K. I. – ZORN, B. – VRTOVEC, H. M. 2001. Germ cell apoptosis in the human testis. In *Pflügers Archiv : European journal of physiology*, vol. 442, 2001, p. 159-160.
- MARTY, M. S. – CARNEY, E. W. – ROWLANDS, J. C. 2011. Endocrine Disruption: Historical Perspectives and Its Impact on the Future of Toxicology Testing. In *Toxicological Sciences*, vol. 120, 2011, p. 93-108.
- MATHUR, P. P. – SARADHA, B. – VAITHINATHAN, S. 2008. Impact of Environmental Toxicants on Testicular Function. In *Immunology, Endocrine & Metabolic Agents - Medicinal Chemistry*, vol. 8, 2008, p. 79-90.
- McLACHLAN, J. A. – NEWBOLT, R. R. – BUROW, M. E. – LI, S.F. 2001. From malformations to molecular mechanisms in the male: three decades of research on endocrine disrupters. In *Acta Pathologica, Microbiologica et Immunologica Scandinavica*, vol. 109, 2001, p. 263-272.
- MIURA, C. – TAKAHASHI, N. – MICHINO, F. – MIURA, T. 2005. The effect of par-nonylphenol on Japanese eel (*Anguilla japonica*) spermatogenesis in vitro. In *Aquatic Toxicology*, vol. 71, 2005, p. 133-141.

- MONTGOMERY-BROWN, J. – REINHARD, M. 2003. Occurrence and behavior of alkylphenol polyethoxylates in the environment. In *Environmental Engineering Science*, vol. 20, 2003, p. 471-478.
- MUELLER, S. O. 2004. Xenoestrogens: mechanisms of action and detection methods. In *Analytical and Bioanalytical Chemistry*, vol. 378, 2004, p. 582-587.
- NAGAO, T. – WADA, K. – MARUMO, H. – YOSHIMURA, S. – ONO, H. 2001. Reproductive effects of nonylphenol in rats after gavage administration: a two-generation study. In *Reproductive toxicology*, vol. 5, 2001, p. 293-315.
- NAIR, R. – SHAHA, C. 2003. Diethylstilbestrol induces rat spermatogenic cell apoptosis in vivo through increased expression of spermatogenic cell Fas/FasL system. In *The Journal of Biological Chemistry*, vol. 278, 2003, p. 6470-6481.
- NIMROD, A. C. – BENSON, W. H. 1996. Environmental Estrogenic Effects of Alkylphenol Ethoxylates. In *Critical Reviews in Toxicology*, vol. 26, 1996, p. 335-364.
- O'DONNELL, L. – ROBERTSON, K. M. – JONES, M. E. – SIMPSON, E. R. 2001. Estrogen and spermatogenesis. In *Endocrine reviews*, vol. 22, 2001, p. 289-318.
- PINTO, B. – PICARD, D. – REALI, D. 2004. A recombinant yeast strain as a short term bioassay to assess estrogen-like activity of xenobiotics. In *Annali di igiene : medicina preventiva e di comunità*, vol. 16, 2004, p. 579-585.
- RAMALHO-SANTOS, J. – SCHATTEEN, G. – MORENO, R. D. 2002. Control of membrane fusion during spermiogenesis and the acrosome reaction. In *Biology of Reproduction*, vol. 67, 2002, p. 1043-1051.
- RAYMOND, A. S. – EILDER, B. – ENSSLIN, M. – SHUR, B. D. 2010. Loss of SED1/MFG-E8 results in altered luminal physiology in the epididymis. In *Molecular reproduction and development*, vol. 77, 2010, p. 550-563.
- ROUTLEDGE, E. J. – SUMPTER, J. P. 1997. *Structural* features of alkylphenolic chemicals associated with estrogenic activity. In *The Journal of biological chemistry*, vol. 272, 1997, p. 3280-3288.
- SAEZ, J. M. 1994. Leydig cells: endocrine, paracrine, and autocrine regulation. In *Endocrine Reviews*, vol. 15, 1994, p. 574-626.
- SOLAKIDI, S. – PSARRA, A. M. – NIKOLAROPOULOS, S. – SEKERIS, C. E. 2005. Estrogen receptors alpha and beta (ERalpha and ERbeta) and androgen receptor (AR) in human sperm: localization of ERbeta and AR in mitochondria of the midpiece. In *Human reproduction*, vol. 20, 2005, p. 3481-3487.

- SHERRILL, J. D. – SPARKS, M. – DENNIS, J. – MANSOUR, M. – KEMPPAINEN, B. W. – BARTOL, F. F. – MORRISON, E. E. – AKINGBEMI, B. T. 2010. Developmental exposures of male rats to soy isoflavones impact Leydig cell differentiation. In *Biology of Reproduction*, vol. 83, 2010, p. 488-501.
- SINHA HIKIM, A. P. – SWERDLOFF, R.S. 1999. Hormonal and genetic control of germ cell apoptosis in the testis. In *Reviews of reproduction*, vol. 4, 1999, p. 38-47.
- SONNENSCHNEIN, C. – SOTO, A. M. 1998. An updated review of environmental estrogen and androgen mimics and antagonists. In *The Journal of Steroid Biochemistry and Molecular Biology*, vol. 65, 1998, p. 143-150.
- STROHECKER, T. – PICARD, K. – LHUGUENOT, J. C. – CANIVENC-LAVIER, M. C. – CHAGNON, M. C. 2004. Steroid activities comparison of natural and food wrap compounds in human breast cancer cell lines. In *Food and Chemical Toxicology*, vol. 42, 2004, p. 887-97.
- TABIRA, Y. – NAKAI, M. – ASAI, D. – YAKABE, Y. – TAHARA-SHINMYOZU, T. – NOGUCHI, M. – TAKATSUKI, M. – SHIMOHIGASHI, Y. 1999. Structural requirements of para-alkylphenols to bind to estrogen receptor. In *European journal of biochemistry / FEBS*, vol. 262, 1999, p. 240-245.
- TAN, B. L. – KASSIM, N. M. – MOHD, M. A. 2003. Assessment of pubertal development in juvenile male rats after sub-acute exposure to bisphenol A and nonylphenol. In *Toxicology letters*, vol. 143, 2003, p. 261-270.
- TOPPARI, J. – LARSEN, J. C. – CHRISTIANSEN, P. – GIWERCMAN, A. – GRANDJEAN, P. – GUILLETTE, L. J. Jr. – JEGOU, B. – JENSEN, T. K. – JOUANNET, P. – KEIDING, N. – LEFFERS, H. – McLACHLAN, J. A. – MEYER, O. – MULLER, J. – RAJPERT-De MEYTS, E. – SCHEIKE, T. – SHARPE, R. M. – SUMPTER, J. S. – SKAKKEBAEK, N. E. 1996. Male reproductive health and environmental xenoestrogens. In *Environmental health perspectives*, vol. 104, 1996, p. 741-803.
- UGUZ, C. – VARISLI, O. – AGCA, C. – AGCA, Y. 2009. Effects of nonylphenol on motility and subcellular elements of epididymal rat sperm. In *Reproductive toxicology*, vol. 28, 2009, p. 542-549.
- USEPA. 2005. Aquatic life ambient water quality criteria – Nonylphenol (Edited by Water O.o.)
- WANG, H. – YANG, X. – ZHANG, Z. – XU, H. 2003. Both calcium and ROS as common signals mediate Na₂SeO₃-induced apoptosis in SW480 human colonic carcinoma cells. In *The Journal of Inorganic Biochemistry*, vol. 97, 2003, p. 221-230.

- WEBER, L. P. – YIANNIS, K. – HWANG, G. S. – NIIMI, A. J. – JANZ, D. M. – METCALFE, C. D. 2002. Increased cellular apoptosis after chronic aqueous exposure to nonylphenol and quercetin in adult medaka (*Oryzias latipes*). In *Comparative Biochemistry and Physiology Part C: Pharmacology, Toxicology and Endocrinology*, vol. 131, 2002, p. 51-59.
- WHITE, R. – JOBLING, S. – HOARE, S. A. – SUMPTER, J. P. – PARKER, M. G. 1994. Environmentally persistent alkylphenolic compounds are estrogenic. In *Endocrinology*, vol. 135, 1994, p. 175-182.
- WU, J. J. – WANG, K. L. – WANG, S. W. – HWANG, G. S. – MAO, I. F. – CHEN, M. L. – WANG, P. S. 2010. Differential effects of nonylphenol on testosterone secretion in rat Leydig cells. In *Toxicology*, vol. 268, 2010, p. 1-7.
- YENUGU, S. – HAMIL, K. G. – FRENCH, F. S. – HALL, S. H. 2006. Antimicrobial actions of human and macaque sperm associated antigen (SPAG) 11 isoforms: influence of the N-terminal peptide. In *Molecular and cellular biochemistry*, vol. 284, 2006, p. 25-37.
- YING, G. G. – WILLIAMS, B. – KOOKANA, R. 2002. Environmental fate of alkylphenols and alkylphenol ethoxylates--a review. In *Environment international*, vol. 28, 2002, p. 215-226.