

SCREENING OF PLANT EXTRACTS FOR ANTIMICROBIAL ACTIVITY AGAINST BACTERIA

Alexander Vatľák¹, Adriana Kolesárová², Nenad Vukovič³, Katarína Rovná⁴, Jana Petrová¹, Viktória Vimmerová¹, Lukáš Hleba¹, Martin Mellen⁵, Miroslava Kačániová^{*1}

Address(es): prof. Ing. Miroslava Kačániová PhD.,

¹Department of Microbiology, Faculty of Biotechnology and Food Sciences, Slovak University of Agriculture in Nitra, Tr. A. Hlinku 2, 949 76 Nitra, Slovak Republic. ²Department of Animal Physiology, Faculty of Biotechnology and Food Sciences, Slovak University of Agriculture in Nitra, Tr. A. Hlinku 2, 949 76 Nitra, Slovak Republic.

³Department of Chemistry, Faculty of Science, University of Kragujevac, PO Box 12, Serbia.

⁴Department of Animal Products Evaluation and Processing, Faculty of Biotechnology and Food Sciences, Slovak University of Agriculture in Nitra, Slovak Republic. ⁵Hydina Slovensko (Poultry Slovakia), Nová Ľubovňa, 065 11 Nova Ľubovňa.

*Corresponding author: Miroslava.Kacaniova@uniag.sk

ARTICLE INFO	ABSTRACT
Received 8. 10. 2013 Revised 21. 11. 2013 Accepted 8. 1. 2014 Published 1. 2. 2014	The aim of this study was antimicrobial action of the methanolic extracts of <i>Equisetum arvense</i> L. and <i>Urtica dioica</i> L. against gramnegative and grampositive bacteria. The antimicrobial activities of the extracts against gramnegative bacteria: <i>Escherichia coli</i> CCM 3988, <i>Listeria ivanovii</i> CCM 5884, <i>Listeria innocua</i> CCM 4030, <i>Pseudomonas aeruginosa</i> CCM 1960, <i>Serratia rubidaea</i> CCM 4684 and grampositive bacteria: <i>Brochothrix thermosphacta</i> CCM 4769, <i>Enterococcus raffinosus</i> CCM 4216, <i>Lactobacillus rhamnosus</i> CCM 1828, <i>Paenobacillus larvae</i> CCM 4483 and <i>Staphylococcus epidermis</i> CCM 4418 were determined by the disc diffusion method
Regular article	and the microbroth dilution method according to CLSI. Probit analysis was used in this experiment. Of the 2 plant extracts tested, all extracts showed antimicrobial activity against one or more species of microorganisms. The most antimicrobial activity showed methanolic plant extract of <i>E. arvense</i> against <i>S. epidermis</i> with disc diffusion method and with microbroth dilution method against <i>S. rubidaea</i> and plant extract <i>Urtica dioica</i> with disc diffusion method against <i>P. aeruginosa</i> and with microbroth dilution method against <i>S. rubidaea</i> and <i>E. coli</i> .
Regular article	and the microbroth dilution method according to CLSI. Probit analysis was used in this experiment. Of the 2 plant extracts tested, a extracts showed antimicrobial activity against one or more species of microorganisms. The most antimicrobial activity showed methanolic plant extract of <i>E. arvense</i> against <i>S. epidermis</i> with disc diffusion method and with microbroth dilution method against <i>rubidaea</i> and plant extract <i>Urtica dioica</i> with disc diffusion method against <i>P. aeruginosa</i> and with microbroth dilution method against or method against <i>P. aeruginosa</i> and with microbroth dilution method against or method against <i>P. aeruginosa</i> and with microbroth dilution method against or method against <i>P. aeruginosa</i> and with microbroth dilution method against or method against <i>P. aeruginosa</i> and with microbroth dilution method against or method against <i>P. aeruginosa</i> and with microbroth dilution method against or method against <i>P. aeruginosa</i> and with microbroth dilution method against or method against <i>P. aeruginosa</i> and with microbroth dilution method against <i>P. aer</i>

Keywords: Equisetum arvense L., Urtica dioica L., metnanolic extracts, gramnegative and grampositive bacteria

INTRODUCTION

In recent years, the usage of plant materials as food supplement and as alternative medicine has increased due to their phytochemical contents. Among these phytochemicals, alkaloids, carotenoids and phenolics have been widely studied. The most popular area in research is the antioxidant capacities of these substances. Phenolic compounds present in plants exhibit strong antioxidant activities (Guimaraes *et al.*, 2009; Barros *et al.*, 2010).

Equisetum arvense L. (field horsetail) is a fern from the Equisetaceae family, widely spread across the northern hemisphere as a weed in fields and uncultivated land. Multiple healthfulness properties of field horsetail have been known since ancient times and it has been used in the treatment of pulmonary tuberculosis and haemorrhage, anaemia, peptic and other types of ulcers, fistulas and colon polyps, inflammation, bleeding, kidney and bladder tuberculosis (Sandhu et al., 2010; Labun et al., 2013). A large number of papers verify various biological effects of the E. arvense extracts, such as sedative and anticonvulsive, hepatoprotective, antioxidant, antibacterial and antifungal activity (Hyuncheol et al., 2004; Dos Santos et al., 2005; Stajner et al., 2006; Canadanovic-Brunet et al., 2009; Garcia et al., 2011). E. arvense is wellknown for its high content of bioactive components, such as: phenolic compounds, saponins, aconite, oxalic and malic acid, resins, tannins, pectin, flavonic compounds, vitamin C, carotenoids and mineral substances (Radulovic et al., 2006; Uslu et al., 2013). In 2006 researchers investigated the composition and antimicrobial properties of essential oils from equisetum arvense. The twenty five compounds with antimicrobial activities were identified in the essential oil obtained from the aerial parts of the plant (Radulovic et al., 2006). In 2009 a report was published about the antimicrobial and hydroxyl radical scavenging activities of methanol extract of the aerial parts of the plant (Canadanovic-Brunet et al., 2009). Antitumoral activities of the Equisetum arvense peptides were also investigated (Alexandru et al., 2006).

The resistant properties of essences have been known from ancient areas and todays, medicinal plants are very valuable in the industry and scientific researches because of their antimicrobial and antioxidant activities (**Singh** *et al.*, **2006**). *Urtica dioica* which is a member of *Urticaceae* class, its Latin name is

Nettle, has many important functions in traditional treatment because it has a lot of curable effects. There are many reports which show this plant is very effective in the treatment of blood pressure, diabetes, and prostate hyperplasia, rheumatoid arthritis and allergic rhinitis (**Fathi-Azad** *et al.*, 2005). Antimicrobial activities of alcoholic and aqueous extracts of the separate parts of *Urtica* were investigated on the *Staphylococcus aureus, Escherichia coli, Bacillus subtilis, Pseudomonas aeruginosa* and *Candida albicans* in the Islamic Azad University Science-Research Tehran. Its summary illustrated that alcoholic extract of *Urtica* seed had the greatest influence on the gram positive bacteria; leaves extract had the maximum effect on the gram negative bacteria, its blossom oil had the highest impact on the antifungal attribute and aqueous essence had positive effect on the all bacteria except *Pseudomonas* (Majd *et al.*, 2001).

Consumers life is about changes and development. In some causes, it is question of comeback, in another ones the question of futuristic wishes. nevertheless, the only important thing is to satisfy our customer, but nowadays, do not forget sustainability issues in broaden understanding (Horská, 2012).

Both of these plants are well known in the traditional medicine and their history is very long. In this work, antibacterial activity of *Equisetum arvense* L. and *Urtica dioica* L. leaves against 5 different grampositive and gramnegative bacteria was studied.

MATERIAL AND METHODS

Preparation of crude extracts

Leaves and steams samples of *Equisetum arvense* L. and *Urtica dioica* L. were dried and the dried material was ground to a coarse powder. Fifty grams of the sample of dried plant material was extracted extensively in 150 ml methanol for two weeks at room temperature with gentle shaking. The extract was filtered through filter paper (Whatman no. 54) under vacuum followed by drying by rotary evaporation. Detailed information about medical plants show shows tab. 1.

 Table 1 Detail information about plants and plant extracts

Orig. Latin title	Plant parts	Yield	Area	Dissolving time	Extracted by
Urtica dioica	leaf + stem	2007	Nitra	2 weeks at room	Vacuum evaporator from methanol at room
Equisetum arvense	leaf + stem	392.5	Nitra	temperature	temperature at -800 mbar

Tested microorganisms

The following strains of bacteria were used gramnegative bacteria: *Escherichia coli* CCM 3988, *Listeria ivanovii* CCM 5884, *Listeria innocua* CCM 4030, *Pseudomonas aeruginosa* CCM 1960, *Serratia rubidaea* CCM 4684 and grampositive bacteria: *Brochothrix thermosphacta* CCM 4769, *Enterococcus raffinosus* CCM 4216, *Lactobacillus rhamnosus* CCM 1828, *Paenobacillus larvae* CCM 4483 and *Staphylococcus epidermis* CCM 4418. The bacterial strains were purchased from the Czech Collection of Microorganisms (CCM). The microorganisms were grown overnight at 37 °C in Mueller-Hinton Broth (Oxoid, England) at pH 7.4.

Antibacterial activity with disc diffusion method

Antimicrobial activity of each plant extract was determined using a disc diffusion method. Briefly, 100 μ l of the test bacteria were grown in 10 ml of fresh media until they reached a count of approximately 10⁵ cells.ml⁻¹. One hundred microlitres of the microbial suspension was spread onto Mueller Hinton agar plates. The extracts were tested using 6 mm sterilized filter paper discs. The diameters of the inhibition zones were measured in millimeters. All measurements were to the closest whole millimeter. Each antimicrobial assay was performed in at least triplicate. Filter discs impregnated with 10 μ l of distilled water were used as a negative control.

Minimum inhibitory concentration MIC

Minimum inhibitory concentrations (MICs) determination was performed by a serial dilution technique, using 96-well microtitre plates. The bacterial inoculum applied contained approximately 1.0×10^5 cells in a final volume of $100 \ \mu$ l.well⁻¹. The pure plant material tested were dissolved in DMSO (512 to 1 μ g.mL⁻¹) and added to broth medium with bacterial inocula. The microplates were incubated for 16 - 20 hours at 37 °C. The lowest concentrations without visible growth determined as different between start concentration and final concentration of solution by ELISA Reader (Biotek ELx808iU) were defined as concentrations which completely inhibited bacterial growth (MICs). The first row on 96-well microtitrate plate was control of sterility and final row was control of growth without pure compound of plant material.

Statistical analysis

From obtained measured absorbances before and after this experiment we changed differences in absorbance between measuring to set of binary values. These values were assigned exact concentrations. For this experiment we created followed formula: if absorbance values were a lower as 0.05 than numbers for binary system were 1 (inhibitory effect), if absorbance values were a higher as 0.05 than numbers for binary system were 0 (no effect or stimulant effect). For this statistical evaluation Probit analysis in Statgraphic software was used.

RESULTS AND DISCUSSION

Natural products are considered an important source of new antibacterial agents. Medicinal plants continue to be used world-wide for the treatment of various diseases and have a great potential for providing novel drug leads with novel mechanism of action (Singh *et al.*, 2012).

The inhibition zone diameter of selected bacteria compared with *E. arvense* illustrated (tab. 2) that this extract had the highest activity against *S. epidermis* only. Whereas, the maximum diameter of inhibition growth *U. dioica* was related against *P. aeruginosa* and slightly against *E. coli, B. thermospacta, L. rhamnosus* and *P. larvae*.

In this study of Gülçin et al. (2004), nine different microbial and one yeast species were used to screen the possible antimicrobial activity of water exctract

of *U. dioica*. Water extract of *U. dioica* exhibited antimicrobial activity against all tested microorganisms. Of the species used, *Staphylococcus aureus* is one of the most common Gram-positive bacteria causing food poisoning. Its source is not the food itself, but the humans who contaminate food after it has been processed. *Escherichia coli*, belonging to the normal flora of humans, is a Gramnegative bacterium. However, an enterohemmoragic strain of *Escherichia coli* has caused serious cases of food poisoning and preservatives to eliminate its growth are needed.

The results of **Chahardehi** *et al.*, (2012) revealed that ethyl acetate, hexane and chloroform extracts showed higher antimicrobial activity than the other crude extracts, where the ethyl acetate extract showed highest inhibition against *B. cereus*, methicillin resistant *Staphyllococus aureus* and *Vibrio parahaemolyticus*. Terpens and phenols of *U. dioica* are one of the major groups associated with the inhibition of microbial infections and cancer (**Dar** *et al.*, 2012). *U. dioica* is a rich source of phytochemicals such as phenolic compounds and minerals which can be used as a potential source of useful drugs (**Ahmed** *et al.*, 2012).

Table 2 Antibacterial	activity of me	dicinal plants a	gainst bacteria in mm	

Microorganism	Medicinal plant extract	Mean (mm)
-	control	0
E. coli CCM 3988	Equisetum arvense	0
	Urtica dioica	5.00
	control	0
P. aeruginosa CCM 1960	Equisetum arvense	0
-	Urtica dioica	8.00
	control	0
Seratia rubidae CCM 4684	Equisetum arvense	0
	Urtica dioica	0
	control	0
Listeria ivanovii CCM 5884	Equisetum arvense	0
	Urtica dioica	0
	control	0
Listeria innocua CCM 4030	Equisetum arvense	0
	Urtica dioica	0
	control	0
E. raffinosus CCM 4216	Equisetum arvense	0
	Urtica dioica	0
	control	0
B. thermospacta CCM 4769	Equisetum arvense	0
-	Urtica dioica	2.6
	control	0
S. epidermis CCM 4418	Equisetum arvense	3.3
-	Urtica dioica	0
	control	0
L. rhamnosus CCM 1828	Equisetum arvense	0
	Urtica dioica	2.3
	control	0
P. larvae CCM 4483	Equisetum arvense	0
	Urtica dioica	4.6

The determination of the MIC by means of the microbroth dilution method (Table 3) showed that plant extract tested exhibited an antimicrobial effect against some of the ten tested microorganisms. The results of the bioassays showed that extract exhibited moderate to appreciable antibacterial activities against all bacteria. However, this activity varies with the kind of bacteria. The best antimicrobial activity at both medicinal plants were found against four gramnegative bacteria.

Table 3 Determined	MICs	value	for	selected	medical	plants	(MeOH	extracts)	to	gramnegative	and	gram	positive
microorganisms													

		Antimicrobial activity of medicinal plants extract (µg.mL ⁻¹)						
Abr.* N	Microorganisms	Urtica dioid	ca	Equisetum arvense				
		MIC 50	MIC 90	MIC 50	MIC 90			
Gramne	gative microorganisms							
Liv	Listeria ivanovii CCM 5884	1.50	1.63	31.71	55.81			
Sr	Serratia rubidaea CCM 4684	0.75	0.82	24	25.76			
Lin	Listeria innocua CCM 4030	6.0	6.48	766.01	814.26			
Ec	Escherichia coli CCM 3988	0.75	0.82	> 1024	> 1024			
Pa	Pseudomonas aeruginosa CCM 1960	> 1024	> 1024	> 1024	> 1024			
Grampo	sitive microorganisms							
Er	Enterococcus raffinosus CCM 4216	> 1024	> 1024	> 1024	> 1024			
Lr	Lactobacillus rhamnosus CCM 1828	> 1024	> 1024	> 1024	> 1024			
Se	Staphylococcus epidermis CCM 4418	> 1024	> 1024	> 1024	> 1024			
Bt	Brochothrix thermosphacta CCM 4769	> 1024	> 1024	> 1024	> 1024			
Pl	Paenobacillus larvae CCM 4483	> 1024	> 1024	> 1024	> 1024			

*Abreviations, (MICs determined by Probit analysis, p< 0.05)

The essential oil of Equisetum arvense L. in study of Radulovič et al. (2006) was shown to possess a broad spectrum of strong antimicrobial activity against all tested strains. The diameters of growth inhibition zones ranged from 23 to 37 mm (including the diameter of the disk, 12.7 mm) with the highest inhibition zone values observed against Gram-negative S. enteritidis (35 mm) and K. pneumoniae (37 mm). Significant reductions in bacterial growth were obtained with medically important pathogens such as S. aureus (28 mm). The activity was greater or similar to conventional antibiotics even in the case of C. albicans and the fungal filamentous organism A. niger. In the present study, the Gramnegative bacteria K. pneumoniae, P. aeruginosa and S. enteritidis were more susceptible than the Grampositive S. aureus except for the strains of E. coli (Gramnegative) that were the most resistant of the tested bacteria. It has been frequently reported that Gramnegative bacteria are more resistant to the inhibitory effects of essential oils (Smith-Palmer et al., 1998), and this was attributed to the microbial cell impermeability due to the presence of certain lipopolysaccharides in the cell walls. All tested microorganisms were completely non-susceptible to control disks imbued with ether. The antimicrobial nature of the E. arvense essential oil can be attributed to the presence of various substances, mainly the phenolic monoterpene thymol (Tepe et al., 2004; Pattnaik et al., 1997). In addition the combination of thymol and 1,8-cineole may have resulted in a significant synergistic antifungal effect as previously published (Pina-Vaz et al., 2004). Linalool has also been reported as having antibacterial (Onawunmi et al., 1984) and antifungal activity (Reuveni et al., 1984). β-Ionone was shown to possess antimicrobial effect on the strains of S. aureus while in the same study βcaryophyllene had no pronounced activity (Kubo et al., 1992). Very similar patterns of activity of α - and β -ionones compared with the activity of *E. arvense* oil against P. aeruginosa, E. coli, S. aureus and C. albicans were observed previously and their activity correlated with their solubility in water, due to the great importance of the diffusion ability of the compounds in the disk diffusion assay (Griffin et al., 1999). Trans-Phytol was the principal active component responsible for the antimycobacterial activity of the methanol extract of Leucas volkensii (Rajab et al., 1998). The essential oil of E. arvense showed strong antimicrobial activity in vitro and may, despite the small yield, contribute to the medicinal properties of the plant.

CONCLUSION

In conclusion, we can to state that the most antimicrobial activity showed methanolic plant extract of *E. arvense* against *S. epidermis* with disc diffusion method and with microbroth dilution method against *S. rubidaea* and plant extract *Urtica dioica* with disc diffusion method against *P. aeruginosa* and with microbroth dilution method against *S. rubidaea* and *E. coli*.

Acknowledgments: The Paper was supported by the project: Development of International Cooperation for the Purpose of the Transfer and Implementation of Research and Development in Educational Programs conducted by the Operational Program: Education, ITMS code: 26220220525, by grant of KEGA 013SPU-4/2012, VEGA 1/0129/13, APVV grant 0304-12, Food and Agriculture COST Action FA1202 and by European Community under project no 26220220180: Building Research Centre "AgroBioTech".

REFERENCES

AHMED, A.A., ZAIN, U., ABJULUZIZ, M.A., RIUS, U., IUBUL, H., MUHAMMAD, T. 2012. Evaluation of the chemical composition and element analysis of *Urtica dioica*. *African Journal of Pharmacy*, 6(21), 1555-1558.

ALEXANDRU, V., PETRUSCA, D.N., GILLE, E. 2007. Investigation of Proapoptotic Activity of *Equisetum Arvense* 1. Water Extract on Human Leukemia u 937 cells. *Romanian Biotechnological Letters.*, 12(2), 3139-3148.

BARROS, L., OLIVEIRA, S., CARVALHO, A.M., FERREIRA, I.C.F.R. 2010.

In vitro Antioxidant Properties and Characterization in Nutrients And Phytochemicals Of Six Medicinal Plants from The Portuguese Folk Medicine.

Industrial Crops and Products, 32(3), 572-579.

CANADANOVIC'- BRUNET, J.M., CETKOVIC', G.S., DJILAS, S.M., TUMBAS, V.T., SAVATOVIC', S.S., MANDIC', A.I., MARKOV, S.L., CVETKOVIC, D.D. 2009. Radical Scavenging and Antimicrobial Activity of Horsetail (*Equisetum arvense* L.) Extracts. *International Journal of Food Science and Technology.*, 44(2), 269-278.

DAR, S.A., YOUSUF, A.R., GANAI, F.A., SHARMA, P., KUMAR, N., SINGH, R. 2012. Bioassay guided isolation and identification of antiinflammatory and antimicrobial compounds from *Urtica dioica* L. (*Uriticaceae*) leaves. *African Journal of Biotechnology*, 11(65), 12410-12420.

DOS SANTOS, J.G., BLANCO, M.M., DO MONTE, F.H.M., RUSSI, M., LANZIOTTI, V.M.N.B., LEAL, L.K.A.M., CUNHA, G.M. 2005. Sedative and anticonvulsant effects of hydroalcoholic extract of *Equisetum arvense*. *Fitoterapia*, 76(6), 508-513.

FATHI-AZAD, F., GARJANI, A., MALEKI, N., RANJDOST, S. 2005. Study of the hypoglycemic activity of the hydro alcoholic extract of *Urtica dioica* in normal and diabetic rats. *Pharmaceutical Sciences*, 94(2), 65-69.

GARCÍA, D., RAMOS, A.J., SANCHIS, V., MARÍN, S. 2012. Effect of *Equisetum arvense* and *Stevia rebaudiana* extracts on growth and mycotoxin production by *Aspergillus flavus* and *Fusarium verticillioides* in maize seeds as affected by water activity. *International Journal of Food Microbiology*, 153(1-2), 21-27.

GRIFFIN, S.G., WYLLIE, S.G., MARKHAM, J.L., LEACH, D.N. 1999. The role of structure and molecular properties of terpenoids in determining their antimicrobial activity. *Flavour and Fragrance Journal*, 14(5), 322–332.

GUIMARÃES, R., BARROS, R., CARVALHO, A., SOUSA, M., MORAIS, J., FERREIRA, I.C.F.R. 2009. Aromatic Plants as A Source of Important Phytochemicals: Vitamins, Sugars and Fatty Acids in *Cistus Ladanifer*, *Cupressus Lusitanica* and *Eucalyptus Gunnii* Leaves. *Industrial Crops and Products*, 30(3), 427-430.

GÜLÇIN, I., KÜFREVIOGLU, O.I., OKTAY, M., BÜYÜKOKUROGLU, M.E. 2004. Antioxidant, antimicrobial, antiulcer and analgesic activities of nettle (*Urtica dioica* L.). *Journal of Ethnopharmacology*, 90(2-3), 205–215.

HORSKÁ, E. 2012. New Consumerism and Trends at the Food Market: within and Beyond Visegrad Borders. In Horská, E. Food Sciences & Business Studies. Nitra : SPU, 2012, 410 p. ISBN 978-80-552-0815-2.

HYUNCHEOL, O., DO-HOON, K., JUNG-HEE, CH, YOUN-CHUL, K. 2004. Hepatoprotective and free radical scavenging activities of phenolic petrosins and flavonoids isolated from *Equisetum arvense*. *Journal of Ethnopharmacology*, 95(2-3), 421-424.

CHAHARDEHI, A.M., IBRONIM, D., SULAIMANI, S.F., MOUSAVI, L. 2012. Screening antimicrobial activity of various extracts of *Urtica dioica*. *International Journal of Tropical Biology*, 60 (4), 1567-1576.

KUBO, I., MUROI, H., HIMEJIMA, M. 1992. Antimicrobial activity of green tea flavour components and their combination effects. *Journal of Agricultural Food Chemistry*, 40, 245-248.

LABUN, P., GRULOVA, D., SALAMON, D., ŠERŠEŇ, F. 2013. Calculating the Silicon in Horsetail (*Equisetum arvense* L.) during the Vegetation Season, *Food and Nutrition Sciences*, 4, 510-514

MAJD, A., MEHRABIAN, S., JAFARI, Z. 2001. Tissue culture of some species of Artemisia and studing the antimicrobial effects of these species. *Iran Journal of Medicinal Aromatic Plants*, 19(3), 287.

ONAWUNMI, G.O., YISAK, W.A., OGUNLANA, E.O. 1984. Antibacterial constituents in the essential oil of *Cymbopogon citratus* (DC) Stapf. *Journal of Ethnopharmacology*, 12(3), 279–286.

PATTNAIK, S., SUBRAMANYAM, V.R., BAPAJI, M., KOLE, C.R. 1997. Antibacterial and antifungal activity of aromatic constituents of essential oils. *Microbios*, 89, 39-46. PINA-VAZ, C., RODRIGUES, A.G., PINTO, E., COSTA-DE-OLIVEIRA, S., TAVARES, C., SALGUEIRO, L., CAVALEIRO, C., GONÇALVES, M.J., MARTINEZ-DE-OLIVEIRA, J. 2004. Antifungal activity of thymus oils and their major compounds. *Journal of European Academy of Dermatology and Venereology*, 18(1), 73-78.

RAJAB, M.S., CANTRELL, C.L., FRANZBLAU, S.G., FISHER, N.H. 1998. Antimycobacterial activity of (E)-phytol and derivatives. A preliminary structureactivity study. *Planta Medicine*, 64(1), 2–4.

REUVENI, R., FLEISCHER, A., PUTIEVSK, E. 1984. Fungistatic activity of essential oils from *Ocimum basilicum* chemotypes. *Phytopathology*, 110(1), 20–22.

SANDHU, N.S., KAUR, S. CHOPRA, D. 2010. "Equisetum Aervens: Pharmacology and Phytochemistry—A Review," Asian Journal of Pharmaceutical and Clinical Research, 3(3), 146-150.

SINGH, G., MAURYA, S., LAMPASONA, M.P., CATALAN, A.N. 2006. Studies on essential oils, Part 41. Chemical composition, antifungal, antioxidant and sprout suppressant activities of coriander (*Coriandru sativum*) essential oil and its oleoresin. *Flavor and Fragrance Journal*, 21(3), 472-479.

SINGH, R., DAR, S.A., SHARMA, P. 2012. Antibacterial activity and toxicological evaluation of semipurified hexane extract of *Urtica dioica* leaves. *Research Journal of Medicinal Plants*, 6(2), 123-135.

SMITH-PALMER, A., STEWART, J., FYFE, L. 1998. Antimicrobial properties of plant essential oils and essences against five important food-borne pathogens. *Letters of Applied Microbiology*, 26(1), 118–122.

ŠTAJNER, D., POPOVIĆ, B.M., ČANADANOVIĆ-BRUNET, J., BOŽA, P. 2006. Free radical scavenging activity of three *Equisetum* species from Fruška gora mountain. *Fitoterapia*, 77(7-8), 601 - 604.

RADULOVIĆ, N., STOJANOVIĆ, G., PALIĆ, R. 2006. Composition and Antimicrobial Activity of *Equisetum arvense* L. Essential Oil. *Phytotherapy Research*, 20(1), 85-88.

TEPE, B., DAFERERA, D., SOKMEN, M., POLISSIOU, M., SOKMEN, A. 2004. *In vitro* antimicrobial and antioxidant activities of the essential oils and various extracts of Thymus eigii M Zohary et P.H. Davis. *Journal of Agricultural and Food Chemistry*, 52(5), 1132–1137.

TEPKEEVA, I. I., MOISEEVA, E.V., CHAADAEVA, A.V., ZHAVORONKOVA, E.V., KESSLER, Y.V., SEMUSHINA, S.G., DEMUSHKIN, V.P. 2008. Evaluation of Antitumor Activity of Peptide Extracts from Medicinal Plants on the Model of Transplanted Breast Cancer in CBRB-Rb(8.17)11em Mice. *Bulletin of Experimental Biology and Medicine*. 145(4), 464-466.

USLU, M.E., ERDOGAN, I., BAYRAKTAR, O., ATES, M. 2013. Optimization of extraction conditions for active components in *Equisetum arvense* extract. *Romanian Biotechnological Letters*, 18(2), 8115-8131.