



THE EFFECT OF PATULIN ON RABBIT SPERM MOTILITY AND PROGRESSIVE MOTILITY

Monika Schneidgenová^{1*}, Anna Kalafová¹, Jana Emrichová¹, Katarína Zbynovská¹, Peter Petruska¹, Lubomír Ondruska², Rastislav Jurčík², Lubica Chrastinová², Marcela Capcarová¹

Address(es): Ing. Monika Schneidgenová, PhD.,

¹Department of Animal Physiology, Faculty of Biotechnology and Food Sciences, Slovak University of Agriculture in Nitra, Tr. A. Hlinku 2, 949 76 Nitra, Slovak Republic.

²Animal Production Research Centre Nitra, Hlohovecka 2, 949 01 Nitra, Slovak Republic.

*Corresponding author: monika.schneidgenova@uniag.sk

ARTICLE INFO

Received 14. 10. 2013

Revised 20. 11. 2013

Accepted 8. 1. 2014

Published 1. 2. 2014

Regular article

ABSTRACT

The aim of our study was to compare the motility and progressive motility of rabbit sperm after patulin intramuscular administration twice a week for two weeks. Adult male rabbits (n= 30) were used in experiment. Animals were divided into two groups: control group (C) without patulin exposure and experimental group (E) with addition of patulin (10 µg.kg⁻¹ of body weight). Semen collection was performed using an artificial vagina. All samples were analysed using CASA (Computer Assisted Semen Analysis) system and following parameters were evaluated: percentage of motile spermatozoa and percentage of progressive motile spermatozoa. The sperm motility (p<0.05) as well as the progressive motility (p<0.05) was significantly lower in E group when compared to the control.

Keywords: rabbit, motility, progressive motility, patulin



INTRODUCTION

Patulin is a mycotoxin produced by several *Penicillium*, *Aspergillus* and *Byssachlamys* species. Patulin can be produced on different food products including fruits, grains, cheese, cured meats, but in natural situations patulin is exclusively found in apple and apple products (Mortimer *et al.*, 1985; Harrison, 1989; Paster *et al.*, 1995). There are a number of research studies related to reproductive and developmental toxicity, carcinogenicity, mutagenicity and immunotoxicity of patulin (Becci *et al.*, 1981; Choudhary *et al.*, 1992; Smith *et al.*, 1993; Llewellyn *et al.*, 1998; Alves *et al.*, 2000). Patulin has a strong affinity for sulfhydryl group, which explains why it inhibits the activity of many enzymes (Askar, 1999). The seminal characteristics are affected by many factors (breed, feeding, health status, rearing condition, season and collection

The aim of present study was to analyse motility and progressive motility of rabbit's sperm after patulin intramuscular administration twice a week for two weeks.

MATERIAL AND METHODS

Animals

Adult male rabbits (n= 30), maternal albinotic line (crossbreed Newzealand white, Buskat rabbit, French silver) and paternal acromalactic line (crossbreed Nitra's rabbit, Californian rabbit, Big light silver) were used in experiment. Rabbits were healthy and their condition was judged as good at the commencement of the experiment. Water was available at any time from automatic drinking troughs. Groups of adult animals were balanced for age (150 days) and body weight (4 ± 0.5 kg) at the beginning of the experiment. Adult rabbits were fed diet of a 12.35 MJ.kg⁻¹ of metabolizable energy (Table 1) composed of a pelleted concentrate. Animals were divided into two groups: control group (C) and experimental group (E) with addition of patulin (10 µg.kg⁻¹ of body weight). Animals from E group received patulin through intramuscular injection twice a week for two weeks.

In this animal study, institutional and national guidelines for the care and use of animals were followed, and all experimental procedures involving animals were approved by ethical committee.

Semen collection and analysis

Semen collection was performed using an artificial vagina. All samples were analysed using CASA (Computer Assisted Semen Analysis) system – SpermVision (Minitüb, Tiefenbach, Germany) combined with Olympus BX 51 microscope (Olympus, Japan) and following parameters were evaluated: percentage of motile spermatozoa (motility >5 µm/s) and percentage of progressive motile spermatozoa (motility >20µm/s).

Statistical analysis

The data used for statistical analyses represent means of values obtained in blood collection. To compare the results, t-test was applied to calculate basic statistic characteristics and to determine significant differences among the experimental and control groups. Statistical software SIGMA PLOT 12.0 (Jandel, Corte Madera, CA, USA) was used. Differences were compared for statistical significance at the level P< 0.05.

Table 1 The chemical composition (g.kg⁻¹) of experimental diet.

Component	
Dry mater	926.26
Crude protein	192.06
Fat	36.08
Fibre	135.79
Non-nitrogen compounds	483.56
Ash	78.78
Organic mater	847.49
Calcium	9.73
Phosphorus	6.84
Magnesium	2.77
Sodium	1.81
Potassium	10.94
Metabolizable energy	12.35 MJ.kg ⁻¹

RESULTS AND DISCUSSION

The results of effect of patulin on rabbit sperm motility and progressive motility are summarized in Table 2. Motility was significantly lower in E group (59.85 %; $p < 0.05$) as compared with the control group C (82.17 %). Similarly, progressive motility was significantly lower in E group (44.78; $p < 0.05$) as compared with the control group C (73.12%).

Table 2 The percentage of motility and progressive motility of rabbit's spermatozoa after patulin administration.

Group	C	E
MOT (%)		
x	82.17	59.85*
min	74.57	37.58
max	91.20	86.66
SD	5.85	19.22
PROG (%)		
x	73.12	44.78*
min	62.71	19.14
max	80.21	70.93
SD	6.52	20.84

Legend: C – control group, E – experimental group with patulin exposure, MOT (%) – motility spermatozoa, PROG (%) – progressive motility spermatozoa, x – mean, min – minimum value, max – maximum value, SD – standard deviation,

* - means significant difference $p < 0.05$

Selmanoğlu (2006) examined the effects of patulin on the epididymis, seminal vesicle and prostate tissues. While sperm counts increased in patulin-treated rats for 60 days, sperm counts in patulin-treated rats for 90 days decreased compared to the corresponding control group. Patulin affected sperm morphology of growing male rats. In histological examinations of the testes of rats treated with patulin, oedema, fibrosis and local Leydig cell hyperplasia in the interstitial tissue, and disorganization of seminiferous tubule epithelium were observed. The thyroid of rats treated with patulin revealed lymphoid cell infiltration and enlargement of interstitial tissue between follicles, and degenerated colloid. (**Selmanoğlu, Koçkaya, 2004**)

Becci et al. (1981) reported that patulin did not affect some reproductive parameters such as mating success, litter size, fertility, gestation, viability and lactation indices, and pup weight in Wistar rats exposed to 0, 0.1, 0.5, or 1.5 mg/kg bw/day of patulin for 4 weeks before mating, and dosed pregnant females through gestation and lactation.

Meistrich et al. (1985) have reported that a sample containing a high percentage of abnormal sperm is indicative of impaired fertility.

Results of **Selmanoğlu and Koçkaya (2004)** revealed that while patulin caused an increase (66.6%) in testosterone levels and a decrease (17.3%) in T4 levels of rats treated for 60 days, there was no change in the other hormone levels compared to those of the control group. When patulin treatment was extended to 90 days, increased serum testosterone (75%) and LH levels (146%) were observed.

CONCLUSION

In our experiment, we found that patulin administered to animals at a dose of 10 $\mu\text{g}\cdot\text{kg}^{-1}$ significantly decreased motility and progressive motility of rabbit's sperm. Patulin had therefore a negative effect on sperm endpoints of rabbits.

Acknowledgments: This work was financially supported by VEGA scientific grants 1/0084/12, VEGA project No. 1/0532/11, VEGA 1/0022/13, APVV-0304-12 and KEGA 030 SPU-4/2012 and by European Community under project no 26220220180: Building Research Centre „AgroBioTech”.

REFERENCES

ALVES, I., OLIVEIRA, N.G., LAIRES, A., RODRIGUES, A.S., RUEFF, J. 2000. Induction of micronuclei and chromosomal aberrations by the mycotoxin patulin in mammalian cells: role of ascorbic acid as a modulator of patulin clastogenicity. *Mutagenesis*, 15 (3), 229–234.

ASKAR, A. 1999. Patulin in apple juice and children's apple food. *Fruit Processing*, 3, 74–77.

BECCI, P.J., HESS, F.G., JOHNSON, W.D., GALLO, M.A., BABISH, J.G., DAILEY, R.E., PARENT, R.A. 1981. Long-term carcinogenicity and toxicity studies of patulin in the rat. *Journal of Applied Toxicology*, 1, 256–261.

HARRISON, M.A. 1989. Presence and stability of patulin in apple products: a review. *Journal of Food Safety*, 9, 147–153.

CHOUDHARY, D.N., SAHAY, G.R., SINGH, J.N. 1992. Effect of some mycotoxins on reproduction in pregnant albino rats. *Journal of Food Science Technology*, 29 (4), 264–265.

LLEWELLYN, G.C., MCCAY, J.A., BROWN, R.D., MUSGROVE, D.L., BUTTERWORTH, L.F., MUNSON, A.E., WHITE, K.L. 1998. Immunological evaluation of the mycotoxin patulin in female B6C3F₁ mice. *Food and Chemical Toxicology*, 36, 1107–1115.

MEISTRICH, M.L., GOLDSTEIN, L.S., WYROBEK, A.J. 1985. Long-term infertility and dominant lethal mutations in male mice treated with adriamycin. *Mutation Research*, 152, 53–65.

MORTIMER, D.M., PARKER, L., STEPHARD, M.J., GILBERT, J.A. 1985. A limited survey of retail apple and grape juices for the mycotoxin patulin. *Food Additives and Contaminants*, 2, 165–170.

PASTER, N., HUPPERT, D., BARKAI-GOLAN, R. 1995. Production of patulin by different strains of *Penicillium expansum* in pear and apple cultivars stored at different temperatures and modified atmospheres. *Food Additives and Contaminants*, 12, 15–58.

SELMANOĞLU G. 2006. Evaluation of the reproductive toxicity of patulin in growing male rats. *Food and Chemical Toxicology*, 44(12), 2019–24. Epub 2006 Jul 12.

SELMANOĞLU, G., KOÇKAYA, E.A. 2004. Investigation of the effects of patulin on thyroid and testis, and hormone levels in growing male rats. *Food and Chemical Toxicology*, 42(5), 721–727.

SMITH, E.E., DUFFUS, E.A., SMALL, M.H. 1993. Effects of patulin on postimplantation rat embryos. *Archives of Environmental Contamination and Toxicology*, 25, 267–270.