

GENETIC DIVERSITY ANALYSIS OF CASTOR (*RICINUS COMMUNIS* L.) USING SSR MARKERS

Martin Vivodík*, Želmíra Balážová, Zdenka Gálová

Address(es):

Slovak University of Agriculture in Nitra, Faculty of Biotechnology and Food Sciences, Department of Biochemistry and Biotechnology, Tr. A. Hlinku 2, 949 76, Nitra, Slovak Republic.

*Corresponding author: vivodikmartin@gmail.com

doi: 10.15414/jmbfs.2016.6.2.777-780

ARTICLE INFO

Received 23. 2. 2016
Revised 12. 5. 2016
Accepted 12. 5. 2016
Published 3. 10. 2016

Regular article



ABSTRACT

The aim of this study was to assess genetic diversity within the set of 60 ricin genotypes using 5 SSR primers. Ten SSR primers revealed a total of 36 alleles ranging from 5 to 10 alleles per locus with a mean value of 7.20 alleles per locus. The PIC values ranged from 0.758 (Rco30) to 0.879 (Rco29) with an average value of 0.829 and the DI value ranged from 0.774 (Rco30) to 0.881 (Rco29) with an average value of 0.836. Probability of identity (PI) was low ranged from 0.002 (Rco29) to 0.015 (Rco30) with an average of 0.006. A dendrogram was constructed from a genetic distance matrix based on profiles of the 5 SSR loci using the unweighted pair-group method with the arithmetic average (UPGMA). According to analysis, the collection of 60 diverse accessions of castor bean was clustered into five clusters. Cluster 1 contained 14 genotypes, cluster 2 included 7 genotypes of ricin and cluster 3 contained 8 genotypes of ricin. Cluster 4 included 10 genotypes and cluster 5 contained 21 genotypes. We could not distinguish 4 genotypes grouped in cluster 1, RM-103 and RM-104 and genotypes RM-100 and RM-101, which are genetically the closest. Knowledge on the genetic diversity of castor can be used to future breeding programs for increased oil production to meet the ever increasing demand of castor oil for industrial uses as well as for biodiesel production.

Keywords: Castor; Dendrogram; Genetic diversity; Simple sequence repeat (SSR)

INTRODUCTION

Castor (*Ricinus communis* L.) is a cross-pollinated diploid ($2n = 2x = 20$) species belonging to the family *Euphorbiaceae* and genus *Ricinus*. Its seed oil has multifarious applications in production of wide industrial products ranging from medicines to lower molecular weight aviation fuels, fuel additives, biopolymers and biodiesel (Ogunniyi, 2006). Castor seeds contain around 50–55% oil which is rich in an unusual hydroxy fatty acid, ricinoleic acid which constitutes about 80–90% of the total fatty acids (Jeong and Park, 2009).

Knowledge of genetic variability is important for breeding programs to provide the basis for developing desirable genotypes. Genetic variability has been studied using molecular techniques, including amplified fragment length polymorphism (AFLP) (Pecina-Quintero *et al.*, 2013), random amplified polymorphism DNA (RAPD) (Petrovičová *et al.*, 2015; Vivodík *et al.*, 2015), single nucleotide polymorphism (SNP) markers (Foster *et al.*, 2010), simple sequence repeat (SSR) (Tan *et al.*, 2014), start codon targeted polymorphism (SCoT) and inter simple sequence repeat (ISSR) (Kallamadi *et al.*, 2015). Pecina-Quintero *et al.*, (2013) used four different AFLP primer pairs. In total, the four combinations of selective primers amplified 430 products, of which 228 were polymorphic. Vivodík *et al.*, (2014) used 8 RAPD markers to detect genetic variability among the set of 40 castor genotypes. Foster *et al.*, (2010) analyzed the population genetics of *R. communis* in a worldwide collection of plants from germplasm and determined the population genetic structure of 676 samples using single nucleotide polymorphisms (SNPs) at 48 loci. The goal of Tan *et al.*, (2014) was to develop a more complete panel of SSR markers that can be used to construct a genetic map of castor bean and to examine genetic variation in this plant. The present investigation of Kallamadi *et al.*, (2015) has been undertaken to assess the extent of genetic diversity in 31 accessions of castor using ISSR and SCoT primers. These markers are favourable as they exhibit high locus-specificity, high levels of variability, robustness towards genotyping, and a co-dominant mode of inheritance (Woodhead *et al.*, 2005). So far, several investigations on the discrimination between crop genotypes using SSR markers have been carried out by Siripiyasing *et al.*, (2013); Fayyaz *et al.*, (2014); Kanwal *et al.*, (2014); Polat *et al.*, (2015); Yousaf *et al.*, (2015).

This study investigates the genetic diversity among 60 castor genotypes using 5 SSRs markers for the purpose of further breeding ricin.

MATERIAL AND METHODS

Plant material and DNA extraction

A total 60 castor genotypes (called RM-45 – RM-105) obtained from the breeding station Zeainvent Trnava Ltd. (Slovakia), were used in this study. Genotype of castor were grown in a cultivation box at temperature 27 °C and photoperiod 12 hours light and 12 hours dark. DNA of 60 genotypes of castor was extracted from leaves of 10 day old seedlings using the GeneJET Plant Genomic DNA Purification Mini Kit. Each sample was diluted to 20 ng in TE buffer (10 mmol Tris-HCl, pH 8.0 and 0.1 mmol EDTA, pH8.0), stored at -20 °C and resolved on agarose gel with the standard lambda DNA for determining the DNA concentration.

SSR and data analysis

Amplification of SSR fragments was performed according to Bajay *et al.*, (2009, 2011) (Table 1). Polymerase chain reaction (PCR) were performed in 25 µl of a mixture containing 10.5 µl H₂O, 12.0 µl Master Mix (Genei, Bangalore, India), 0.75 µl of each primer (10 pmol) and 1 µl DNA (100 ng). Amplification was performed in a programmed thermocycler (Biometra, Germany) and amplification program consisted of an initial denaturing step at 94 °C for 1 min, followed by 35 cycles of amplification [94 °C (1 min), 1 min at the specific annealing temperature of each primer pair (Table 1), 72 °C (1 min)] and a final elongation step at 72 °C for 10 min. Amplification products were confirmed by electrophoresis in 7% denaturing polyacrylamide gels and silver stained and documented using gel documentation system Grab-It 1D for Windows. Data obtained from SSR analysis were scored as presence (1) or absence (0) of fragments for each castor genotype and entered into a matrix. Based on the similarity matrix, a dendrogram showing the genetic relationships between genotypes was constructed using unweighted pair group method with arithmetic mean (UPGMA) by using the SPSS professional statistics version 17 software package. For the assessment of the polymorphism between castor genotypes and usability of SSR markers in their differentiation diversity index (DI) (Weir, 1990), the probability of identity (PI) (Paetkau *et al.*, 1995) and polymorphic information content (PIC) (Weber, 1990) were used.

Table 1 List of SSR primers (Bajay *et al.*, 2009, 2011)

Marker name	Ta (°C)	Repeat motif	Sequence of the primer (5' - 3')
Rco23	62	(GA) ₁₅ (AG) ₈	F: CATGGATGTAGAGGGTGCAT R: CAGCCAAGCCAAAGATTTTC
Rco26	62	(CT) ₁₉	F: TTGCTTGTCAAAGGGGAGTT R: TCATTTTGAGGGAGAAACCA
Rco29	60	(GA) ₇	F: GGAGAAAAGAAAGGGAGAAGG R: GCCAAAAGCACACTTAATTGA
Rco30	60	(AG) ₁₉	F: TGAAACTTTGGAGCTTGGAGA R: GGTCCCACACATTCATACACA
Rco31	60	(TC) ₁₂ (TCTA) ₄ (AC) ₁₀	F: ACAATGCGTGTGTCTGTGTG R: CCTCAACCCTTTGCTGTTTC

Ta- annealing temperature

RESULTS AND DISCUSSION

Five SSR primers were used for cultivar identification and estimation of the genetic relations among 60 ricin genotypes. All 5 SSR primers generated clear banding patterns with high polymorphism (Figure 1). Five SSR primers revealed a total of 36 alleles ranging from 5 (Rco30) to 10 (Rco29) alleles per locus with a mean value of 7.20 alleles per locus (Table 2). Results indicated the presence of wide genetic variability among different genotypes of castor. Variations in DNA sequences lead to polymorphism. Greater polymorphism is indicative of greater genetic diversity. The PIC values ranged from 0.758 (Rco30) to 0.879 (Rco29) with an average value of 0.829 and the DI value ranged from 0.774 (Rco30) to 0.881 (Rco29) with an average value of 0.836 (Table2). 100% of used SSR markers had PIC and DI values higher than 0.7 that means high polymorphism of chosen markers used for analysis. Probability of identity (PI) was low ranged from 0.002 (Rco29) to 0.015 (Rco30) with an average of 0.006 (Table 2). A dendrogram was constructed from a genetic distance matrix based on profiles of the 5 SSR loci using the unweighted pair-group method with the arithmetic average (UPGMA). According to analysis, the collection of 60 diverse accessions of castor bean was clustered into five clusters. Cluster 1 contained 14 genotypes, cluster 2 included 7 genotypes of ricin and cluster 3 contained 8 genotypes of

ricin. Cluster 4 included 10 genotypes and cluster 5 contained 21 genotypes (Figure 2). We could not distinguish 4 genotypes grouped in cluster 1, RM-103 - RM-104 and RM-100 - RM-101, which are genetically the closest.

Table 2 List of SSR primers, total number of bands and the statistical characteristics of the SSR markers used in castor.

Marker name	Number of alleles	DI	PIC	PI
Rco23	8	0.861	0.856	0.003
Rco26	6	0.822	0.816	0.006
Rco29	10	0.881	0.879	0.002
Rco30	5	0.774	0.758	0.015
Rco31	7	0.843	0.837	0.004
Average	7.20	0.836	0.829	0.006

DI- diversity index, PIC- polymorphic information content, PI- probability of identity

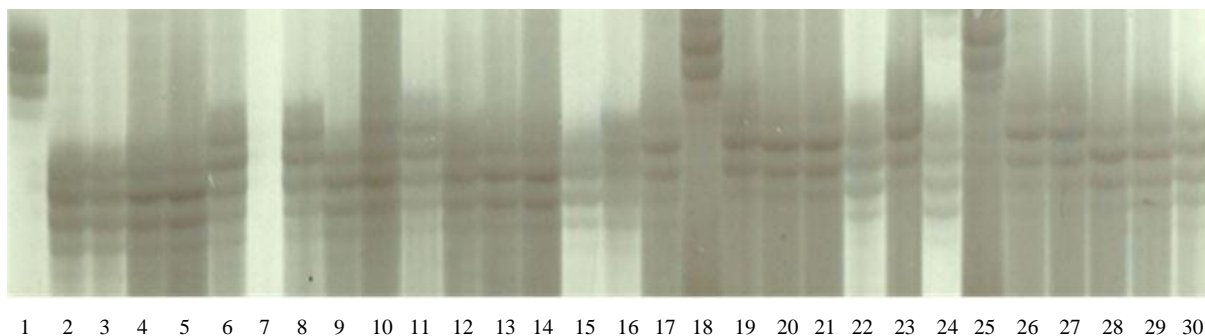


Figure 1 PCR amplification products of 30 genotypes of castor produced by SSR marker Rco29. Lanes 1 - 30 are castor genotypes RM-45 – RM-74.

Similar results detected Pecina-Quintero *et al.*, (2013) who used seven SSR markers and the profiles generated were collectively able to discriminate among 82 *R. communis* accessions and the six controls. Kyoung-In *et al.*, (2011) used 28 SSR loci revealed polymorphisms in a castor bean collection consisting of 72 accessions. A total of 73 alleles were detected, with an average of 3.18 alleles per locus, and the polymorphism information content (PIC) ranged from 0.03 to 0.47 (mean = 0.26). Gedil *et al.*, (2009) used-six primers for analysis of castor. The 6 SSR primers produced amplification products with alleles ranging from 1 to 2 for the parents and the hybrids. The present investigation of Tan *et al.*, (2013) was to assess the genetic diversity in 58 accessions of castor. Seventy alleles were detected among the somaclones and their donors, with an average of 2.1 alleles per locus. Based on the profiles of the SSR loci, a dendrogram was constructed using the unweighted pair-group method with an arithmetic average (UPGMA). Dhingani *et al.*, (2012) used 9 SSR primers for analysis of genetic diversity of castor. SSR analysis yielded 16 fragments, of which 11 were polymorphic, with an average PIC value of 0.87. SSR molecular markers have been used in population genetic studies Yang *et al.*, (2013); Žiarovská *et al.*, (2013); Ahmad *et al.*, (2014); Aslam *et al.*, (2014); Maršálková *et al.*, (2014); Lanciková *et al.*, (2015).

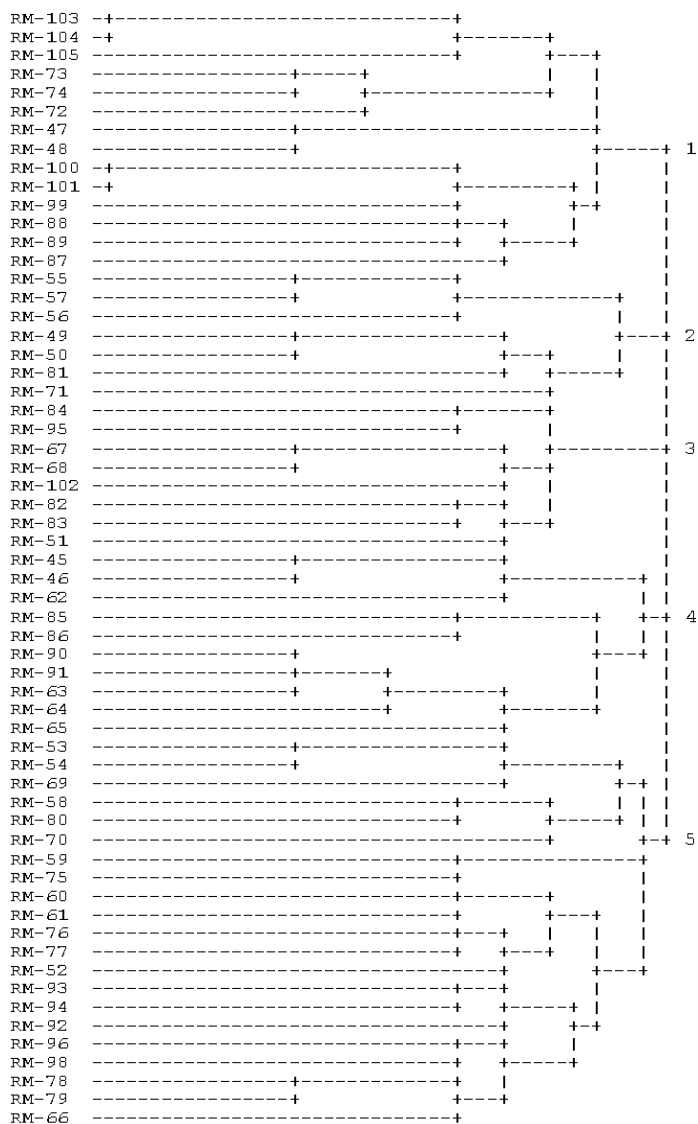


Figure 2 Dendrogram of 60 castor genotypes constructed based on 5 SSR markers.

CONCLUSION

In conclusion, a high level of genetic diversity exists among the castor accessions analyzed. According to analysis, the collection of 60 diverse accessions of castor bean was clustered into five clusters. We could not distinguish 4 genotypes grouped in cluster 1, RM-103 - RM-104 and RM-100 - RM-101, which are genetically the closest. A SSR marker system is a rapid and reliable method for cultivar identification that might also be used in quality control in certified seed production programs, to identify sources of seed contamination, and to maintain pure germplasm collections.

Acknowledgments: This work was co-funded by European Community under project no. 26220220180: Building Research Centre "AgroBioTech" (50%) and KEGA project No. 021SPU-4/2015 (50%).

REFERENCES

Ahmad, Z., Nisar, M., Mumtaz, A. S., Ghafoor, A. and Ali, S. (2014). SSR markers linked to seed size and seed weight in local and exotic chickpea germplasm reported from Pakistan. *Pak. J. Bot.*, 46(6), 2113-2120.
 Aslam, K. and Arif, M. (2014). SSR analysis of chromosomes 3 and 7 of rice (*Oryza sativa* L.) associated with grain length. *Pak. J. Bot.*, 46(4), 1363-1372.
 Bajay, M. M., Pinheiro, J. B., Batista, C. E. A., Nobrega, M. B. M., Zucchi, M. I. (2009). Development and characterization of microsatellite markers for castor (*Ricinus communis* L.), an important oleaginous species for biodiesel production. *Conserv. Genet.*, 1, 237-239. <http://dx.doi.org/10.1007/s12686-009-9058-z>
 Bajay, M. M., Zucchi, M. I., Kiihl, T. A. M., Batista, C. E. A., Monteiro, M., Pinheiro, J. B. (2011). Development of a novel set of microsatellite markers for Castor bean, *Ricinus communis* (Euphorbiaceae). *Am. J. Bot.*, 98, 87-89. <http://dx.doi.org/10.3732/ajb.1000395>

Dhingani, R. M., Tomar, R. S., Parakhia, M. V., Patel, S. V. and Golakiya, B. A. (2012). Analysis of genetic diversity among different *Ricinus communis* genotypes for macrophomina root rot through RAPD and microsatellite markers. *International Journal of Plant Protection*, 5(1), 1-7.
 Fayyaz, L., Farhatullah, Rabbani, M. A., Iqbal, S., Kanwal, M. and Nawaz, I. (2014). Genetic diversity analysis of *Brassica napus/Brassica campestris* progenies using microsatellite markers. *Pak. J. Bot.*, 46(3), 779-787.
 Foster, J. T., Allan, G. J., Chan, A. P., Rabinowicz, P. D., Ravel, J., Jackson, P. J., Keim, P. (2010). Single nucleotide polymorphisms for assessing genetic diversity in castor bean (*Ricinus communis*). *BMC Plant Biol.*, 10, 13-18.
 Gedil, M., Kolade, F., Raji, A., Ingelbrecht, I. and Dixon, A. (2009). Development of molecular genomic tools for verification of intergeneric hybrids between castor bean (*Ricinus communis* L.) and cassava (*Manihot esculenta* Crantz). *Journal of Food, Agriculture & Environment*, 7(2), 534-539.
 Jeong, G. T., Park, D. H. (2009). Optimization of biodiesel production from castor oil using response surface methodology. *Appl. Biochem. Biotech.*, 156, 1-11.
 Kallamadia, P. R., Ganga Rao Nadigatlab, V. P. R., Mulpurib, S. (2015). Molecular diversity in castor (*Ricinus communis* L.). *Industrial Crops and Products*, 66, 271-281. <http://dx.doi.org/10.1016/j.indcrop.2014.12.061>
 Kanwal, M., Farhatullah, Rabbani, M. A., Iqbal, S., Fayyaz, L. and Nawaz, I. (2014). The assessment of genetic diversity between and within brassica species and their wild relative (*Eruca sativa*) using SSR markers. *Pak. J. Bot.*, 46(4), 1515-1520.
 Kyoung-In, S., Gi-An, L., Kyung-Ho, M., Do-Yoon, H., Yong-Jin, P., Jong-Wook, J., Sok-Young, L., Jae-Gyun, G., Chung-Kon, K., Myung-Chul, L. (2011). Isolation and characterization of 28 polymorphic SSR loci from Castor Bean (*Ricinus communis* L.). *J. Crop Sci. Biotech.*, 14(2), 97-103. <http://dx.doi.org/10.1007/s12892-010-0107-7>
 Lanciková, V., Hlavačková, L., Žiarovská, J., Kubíková, H., Bežo, M., Ražná, K., Danchenko, M., Rashydov, N., Hajduch, M. (2015). Analysis of stability of trinucleotide TTC motifs in common flax planted in the Chernobyl area. *J Microbiol Biotech Food Sci.*, 4 (special issue 2), 70-72. <http://dx.doi.org/10.15414/jmbfs.2015.4.special2.70-72>
 Maršálková, L., Židek, R., Pokoradi, J., Golian, J., Belej, L. (2014). Genetic diversity and relatedness among seven red deer (*Cervus Elaphus*) populations. *Potravinarstvo*, 8(1), 15-19. <http://dx.doi.org/10.5219/320>
 Ogunniyi, D. S. (2006). Castor oil: a vital industrial raw material. *Bioresour. Technol.*, 97, 1086-1091.
 Paetkau, D., Calvert, W., Stirling, I., Strobeck, C. (1995). Microsatellite analysis of population structure in Canadian polar bears. *Mol. Ecol.*, 4, 347-354.
 Pecina-Quintero, V., Anaya-López, J. L., Núñez-Colina, C. A., Zamarripa-Colmenero, A., Montes-García, N., Solís-Bonilla, J. L., Aguilar-Rangel, M. R. (2013). Assessing the genetic diversity of castor bean from Chiapas, México using SSR and AFLP markers. *Industrial Crops and Products*, 41, 134-143. <http://dx.doi.org/10.1016/j.indcrop.2012.04.033>
 Petrovičová, L., Gálová, Z., Balážová, Ž., Vivodík, M., Wójcik-Jagla, M., Rapacz, M. (2015). Assessment of RAPD polymorphism in rye (*Secale Cereale* L.) genotypes. *J Microbiol Biotech Food Sci.*, 4 (special issue 2), 94-97. <http://dx.doi.org/10.15414/jmbfs.2015.4.special2.94-97>
 Polat, I., Turgutoglu, E. and Kurt, S. (2015). Determination of genomic diversity within mutant lemon (*Citrus limon* L.) and mandarin (*Citrus reticulata*) using molecular markers. *Pak. J. Bot.*, 47(3), 1095-1102.
 Siripiyasing, P., Kaenratana, K., Mokkaikul, P. and Chaveerach, A. (2013). Molecular assessment for genetic identification and stability of *Cymbidium Sanderae* (Orchidaceae). *Pak. J. Bot.*, 45(2), 519-523.
 Tan, M., Yan, M., Wang, L., Yan, X., Fu, Ch., Wang, L. (2013). Replication of pistillate plants of *Ricinus communis* L. and investigation of the sex stability and genetic variation of the soma clones. *Industrial Crops and Products*, 50, 50-57. <http://dx.doi.org/10.1016/j.indcrop.2013.06.042>
 Tan, M., Wu, K., Wang, L., Yan, M., Zhao, Z., Xu, J., Zeng, Y., Zhang, X., Fu, Ch., Xue, J., Wang, L. X., Yan, X. (2014). Developing and characterising *Ricinus communis* SSR markers by data mining of whole-genome sequences. *Mol Breeding*, 34, 893-904. <http://dx.doi.org/10.1007/s11032-014-0083-6>
 Vivodík, M., Balážová, Ž., Gálová, Z. (2014). Dna analysis of ricin using RAPD technique. *J Microbiol Biotech Food Sci.*, 3 (special issue 2), 181-183.
 Yang, B., Ji, W., Wang, Ch., Xue, F., Ali, M., Wang, Y., Zhang, H. and Liu, X. (2013). Chromosome location and SSR markers of a stripe rust resistance gene from wheat (*Triticum aestivum*) line n9738. *Pak. J. Bot.*, 45(3), 719-724.
 Yousaf, Z., Hu, W., Zhang, Y., Zeng, S. and Wang, Y. (2015). Systematic validation of medicinally important genus epimedium species based on microsatellite markers. *Pak. J. Bot.*, 44(2), 477-484.
 Žiarovská, J., Ražná, K. and Labajová, M. (2013). Using of Inter Microsatellite Polymorphism to evaluate gamma-irradiated *Amaranthus* mutants. *Emir. J. Food Agric.*, 25(9), 673-681. <http://dx.doi.org/10.9755/ejfa.v25i9.15879>
 Weber, J. L. (1990). Informativeness of human (dC-dA)n x (dG-dT)n polymorphism. *Genomics*, 7: 524-530.
 Weir, B. S. (1990). Genetic data analysis. *Methods for discrete population genetic data*. ISBN 0-87893-872-9

Woodhead, M., Russell, J., Squirrell, J., Hollingsworth, P. M., Mackenzie, K., Gibby, M., Powell, W. (2005). Comparative analysis of population genetic structure in *Athyrium distentifolium* (Pteridophyta) using AFLPs and SSRs from anonymous and transcribed gene regions. *Mol Ecol.*, 14, 1681-1695.