

THE HEXOKINASE 1 GENE OF BLOODSTREAM FORM *Trypanosoma brucei brucei* (FEDERE ISOLATE) CONSERVES AMINO ACIDS IN DOMAINS AND MOTIFS PECULIAR TO THE HEXOKINASE 2 SUPERFAMILY

Rotimi Johnson Ojo^{1,3}, Ishaya Yohanna Longdet¹, Richard J. Kutshik¹, Titilayo Omolara Johnson¹ and Yakubu Bitrus²

Address(es):

¹ Department of Biochemistry, University of Jos, Plateau state, Nigeria.

² Department of Biotechnology, NVRI, Vom, Jos, Plateau state, Nigeria.

³ Department of Biochemistry, Faculty of Science and Technology, Bingham University Karu, Nasarawa state, Nigeria.

*Corresponding author: saintlevites@yahoo.com

doi: 10.15414/jmbfs.2019/20.9.3.578-584

ARTICLE INFO

Received 25. 4. 2018

Revised 5. 6. 2019

Accepted 6. 6. 2019

Published 1. 12. 2019

Regular article



ABSTRACT

Trypanosoma brucei brucei hexokinase is the first key regulatory enzyme of the glycolytic pathway which serves as the only source of ATP and nucleic acid precursors for the parasite in the infective stage. This enzyme is attracting much interest because during the infective stage of the parasites, the bloodstream form (BSF) of this parasite depends solely on glycolysis because of its poorly developed mitochondrion. This research, therefore, attempted to characterize the hexokinase 1 gene of the *T. b. brucei* (Federe isolate) which is prevalent in Nigeria. The parasites were grown in rats and purified by diethyl aminoethyl (DEAE) cellulose chromatography. The genomic DNA was isolated, and the parasites hexokinase 1 gene amplified using consensus primers. The amplicon was purified and sequenced. The sequences were studied using software from the National Centre for Biotechnology Information (NCBI) and CLAWSTAL W server. The nucleotide sequence (Accession number MH198230) and the translated amino acids revealed high similarity with sequences of *Trypanosoma brucei brucei* TREU 927, *T. brucei gambiense* and *Trypanosoma cruzi* but low similarity with Human Glucokinase (hexokinase IV). The translated peptide sequence contains the N-terminal peroxisome-targeting signal (PTS-2) that is peculiar to glycosome containing organisms; ATP-binding and the hexokinase binding sites, and other amino acids in the domain peculiar to hexokinase 2 super-family were also conserved. These showed that *T. brucei brucei* hexokinase 1 conserves the unique features peculiar to the hexokinase 2 super-family and its uniquely from its mammalian forms make it a potential target for drug or vaccine against trypanosomiasis.

Keywords: *Trypanosoma brucei brucei*, hexokinase 2 superfamily, hexokinase 1, glycolytic pathway, African trypanosomiasis

INTRODUCTION

African trypanosomiasis is a deadly neglected tropical disease that affects both humans and animals. It is prevalent in African communities where people depend mostly on hunting, livestock rearing, farming and fishing for their livelihood and among those that reside and work around riverine area (Matthews et al., 2015; Shaw et al., 2014). The disease is mainly caused by strains of *Trypanosoma brucei* and is estimated to affect around 70 million people in Africa (Franco et al., 2014; Tesfaye et al., 2012; Simarro et al., 2012). This disease results in an annual economic loss of about \$ 4.5 billion from animal fatality and disease control (Franco et al., 2014; Tesfaye et al., 2012; Simarro et al., 2012; Simarro et al., 2010; Steverding, 2008).

African trypanosomiasis results in death if it is untreated. This makes chemotherapy or vaccination compulsory (Franco et al., 2014). Unfortunately, investment in the research, development and production of new drugs are not on the priority list of pharmaceutical industry since the disease affects only nations who cannot afford the drugs (Bouteille and Buguet, 2012). Therefore there is need for vaccine development that may be attractive to both the pharmaceutical industries and the patients.

One of such targets that is now being considered is *T. brucei brucei* hexokinase, the first key regulatory enzyme of the glycolytic pathway which serves as the only source of ATP and nucleic acid precursors for the parasite in the infective stage and differs from the mammalian host enzyme (Besterio et al., 2002, Willson et al., 2002). This enzyme is attracting attention because during the infective stage of the parasites in the mammalian bloodstream, the parasites live in glucose rich environment in the mammalian bloodstream and depend solely on glycolysis because of its poorly developed mitochondrion which lacks the active components of the citric acid cycle and the electron transport chain. This dependence on glycolysis for ATP in addition to the parasite's poorly developed mitochondrial limits the metabolic options available to the bloodstream form (BSF) of this parasites (Joice et al., 2013; Bringaud et al., 2006; Besterio et al.,

2002). Good understanding of this enzyme can guide in its selective inhibition at different stages of synthesis without interference with the mammalian host hexokinase. This will deprive the parasite of the only source of ATP and nucleic acid precursors. The enzyme is therefore a possible potential target for drugs development against the African trypanosomiasis (Willson et al., 2002). This research, therefore, attempted to characterize the hexokinase 1 gene of the *T.b.brucei* (Federe isolate) which has been the subject of several screening campaigns for small molecule inhibitors in some isolates (Joice et al., 2013).

MATERIALS AND METHODS

Reagents

All reagents used were of analytical grade and purchased from Sigma, BDH POOLE, England, Whatman, USA; Pharmacia Fine Chemicals and Amresco Life Science, Fountain Parkway, Solon. DNA extraction kit was purchased from ZYMO RESEARCH. *Taq* polymerase and High-Fidelity Polymerase Enzyme Mix were bought from Promega, USA and Fermentas, respectively. Molecular size marker was purchased from Roche, Mannheim Germany. Oligonucleotide primers were synthesized by Inqaba biotec Industry, Pretoria South Africa. The GeneAmp PCR System 9700 for gene amplification was obtained from Applied Biosystems, Indonesia and the Gel Documentation System was obtained from SynGene® Inc. Indonesia. Sequencing analyses were performed by Macrogen Corp. in the Netherlands.

Parasite Isolate

Trypanosoma brucei brucei (Federe isolate) was obtained from the Veterinary and Livestock Studies Department of the Nigerian Institute for Trypanosomiasis Research, Vom, Plateau State, Nigeria.

Cultivations of Trypanosomes in Rats

Healthy rats were infected by intraperitoneal inoculation of 1x10⁴ parasites/rat in 0.3ml of Puck saline glucose (PSG) (pH 7.8). Parasitemia was monitored daily from the second day after the infection by bleeding the tail of each animal and the blood observed under a light microscope with fields examined and the mobile parasites counted by using the rapid matching method described by Herbert and Lumsden, (1976), in this method, 20µl of blood from the tail vein of rat was placed on a glass slide and covered with coverslip. The number of parasites was determined microscopically at 40× magnification. The microscopic field was compared with a range of standard logarithmic values and the logarithmic values which matched the microscopic observation were therefore converted to antilogarithm where the absolute number of trypanosome per milliliter (ml) of blood was obtained.

Purification of Bloodstream form *T. brucei brucei* from Rat

After 72 hours from the time of inoculation and at peak parasitemia, the circulating blood was withdrawn by cardiac puncture using a syringe loaded with a 0.2ml of PSG containing heparin (5:1) (see the supplementary material). The whole blood was centrifuged at 1,250 × g for 15 minutes at 4°C resulting in separation of the sample into three different layers. The Buffy layer was carefully removed using a Pasteur pipette and 10⁷*T.bruceibrucei* cells per ml were re-suspended in phosphate buffered saline supplemented with 0.1% glucose (pH = 7.8) and thereafter purified using diethyl aminoethyl cellulose (DEAE) anion exchanger column (DE-52, Whatman) as described by Lanham and Godfrey (1970). 5ml of the eluate containing the trypanosomes in centrifuge tubes were centrifuged for 10mins at 1,600 x g in a bench top centrifuge, and the supernatant was discarded. The trypanosome pellets were then washed by gentle re-suspension in 1ml PBS (pH 7.4) and transferred to micro-centrifuge tubes and centrifuged for 5 minutes at 1,600 x g, the supernatant was removed, the trypanosomes were pooled, and the pellet was re-suspended in PBS (pH 7.4) at 1 x 10⁸cells/ml for subsequent analysis.

GENOMIC DNA ISOLATION

Genomic DNA (gDNA) was extracted from the resulting trypanosome cell pellets using the ZR Fungal/Bacterial DNA MiniPrep (ZYMO RESEARCH) following the manufacturer’s instruction.

Gene Searches and Primer Design

Nucleotide sequence for the *T. brucei brucei* hexokinase 1 gene was located through text searches for the gene name in GeneDB (www.genedb.org) and a BLAST (Basic Local Alignment Search Tool) search against the *T. brucei brucei* genome was performed in NCBI databases (www.ncbi.nlm.nih.gov). Primers based on the sequence obtained from GeneDB were designed as shown below to amplify the gene of *T. brucei brucei* hexokinase 1 (Accession Number:

XM841212.1) and subsequently sent to Inqaba Biotech Industry, Pretoria, South Africa for synthesis.

FORWARD 5’-ATGTCTAGACGCCTAAACAATATCC-3’
 REVERSE 5’-TTACTTGTGCTTACCACCATTGCG-3’

POLYMERASE CHAIN REACTION (PCR)

T. brucei brucei hexokinase 1 (TbHK1) sequence from the total genomic DNA of *T. brucei brucei* was amplified by Polymerase Chain Reaction (PCR) using the designed primers. The PCR reaction was performed in a 50µl reaction mixture which contained 27µl of nuclease free water, 10 µl of 5 X phusion buffers (Promega), 1 µl of dNTPS, 1.5µl of DMSO, 2.5 µl of forward primer, 2.5 µl of reverse primer, 0.5 µl of *Taq*DNA polymerase (Promega) and 5.0 µl of the genomic DNA of *T.brucei brucei*. The tubes containing the mixture will be subjected to 30 cycles of amplification in a thermocycler using the following Amplification conditions: Polymerase activation 94°C for 5 min; Denaturation 94°C for 30 s; Annealing 48°C for 30 s; Extension 72°C for 1 min. Step 2-4 were repeated for 30 cycles and final extension was carried out at 72°C for 5 minutes. The PCR product was then subjected to 1.5% agarose gel electrophoresis. The results were documented using GelDocumentation System (SynGene® Inc. Indonesia).

DNA sequencing and Bioinformatics Analysis

The *T. brucei brucei* (Federe isolate) hexokinase 1 gene amplified was sequenced by MacroGen Corp. in the Netherlands. The sequence obtained was analysed using BLAST tools (BLASTN, BLASTX and BLASTP) provided by NCBI (<http://www.ncbi.nlm.nih.gov/>). The sequence similarity of Tbhk1 and other organism studied were aligning using CLAWSTAL W.

RESULTS

The gene sequence gave 1401bp (Accession number: MH198230.1) and the analysis of the sequence in National Centre for Biotechnology Information(NCBI) BASTN revealed its similarity to *Trypanosoma brucei brucei* TREU927 ([AJ437577.1](#)); *Trypanosoma brucei Gambiense* ([FN554973.1](#)); *Trypanosoma cruzi* ([XM841212.1](#)) hexokinase sequences. The analysis of the sequence gave 87% similarity to those of *T. brucei brucei* (TREU 927) and *Trypanosoma brucei gambiense* and 74% to that of *Trypanosoma cruzi*, the causative agent of American Trypanosomiasis. There was 100% coverage for both *T. brucei brucei* (TREU 927) and *Trypanosoma brucei gambiense* hexokinase sequences with no gap in between while only 38% coverage was observed for *Trypanosoma cruzi* due to the presence of gaps. The result of the comparison is shown in Figure 1 and summarised in Table 1

HK1

ATGTCTAGACGCCTAAACAATATCCTCGAACACATCTCGATCCAGGGAAATGATGGTGAG
 Tcru ATGTCCGCTCGCTGAACAACCTGCTGCAGCACATTGCTGTGAAAGACAAAGACAGCGAC
 Tbg ATGTCTAGACGCCTAAACAATATCCTCGAACACATCTCGATCCAGGGAAATGATGGTGAG
 Tb927 ATGTCTAGACGCCTAAACAATATCCTCGAACACATCTCGATCCAGGGAAATGATGGTGAG

HK1 ACTGTGCGTTCTTTTA-----TGTTACGTTGCACGCCCTACCAACCAACCTTTTCTG
 Tcru ACAATGCGGCACCTGAAACAGCGCATGGCGTGGCGTCCCTCGCGAACCAATTCACGGTG
 Tbg ACTGTGCGTGCCGTTAAGCGTGATGTTGCAATGGCAGCGCTGACCAACCAATTCACAATG
 Tb927 ACTGTGCGTGCCGTTAAGCGTGATGTTGCAATGGCAGCGCTGACCAACCAATTCACAATG

HK1 AGTGTGAGTCTATGCGACAGATCATGACATACCTCCTGTACGAGATGGTGGAGGGTCTT
 Tcru GGCAAGGACCACCTCAAGCAGCTGATGTTGTACATGGTACACCAGATGATTGAGGGACTG
 Tbg AGTGTGGAGTCTATGCGACAGATCATGACATACCTCCTGTACGAGATGGTGGAGGGTCTT
 Tb927 AGTGTGGAGTCTATGCGACAGATCATGACATACCTCCTGTACGAGATGGTGGAGGGTCTT

HK1 GAGGGTCTTACACCACCTCTCCATTTTACC-----ATGTACCCGACGCGGACCCTA--
 Tcru GAGGGACGCGAGAGCACTTTGCGTATGCTGCCTTCTTACGTGTACAAAACCGATCCACG
 Tbg GAGGGTCTGTAAAGCACCCTCCGATGTTACCATCTTATGTGTACAAGGCGGACCCTAAG
 Tb927 GAGGGTCTGTAAAGCACCCTCCGATGTTACCATCTTATGTATACAAGGCGGACCCTAAG

HK1 CGCCCTGGCCTTCTCTCCCTTGGACCTCTGTGTTCCATTTTCTGTGTGTGCGT
 Tcru AAGGCGACAGGCGTCTTCTACGCACTTGACCTCGGGGACGAACTTTCGAGTGCTGCGT
 Tbg CGTGCTACTGGCGTCTTCTACGCACTTGACCTCGGTGGTACCAACTCCGTGTGTGCGT
 Tb927 CGTACTGGCGTCTTCTACGCACTTGACCTCGGTGGTACCAACTCCGTGTGTGCGT

HK1 GGC-CATGCAAGGAGGGTCCCGTGGTGTTCCTCTCCTTCTGCATTCAAAATCCCCAAA
 Tcru GTAACGTGCAAGGAAGGAAGAGTAGCGGACCGTGGACGCCAAGTTTGTGATTCCGCAG
 Tbg GTTGCATGCAAGGAGGGTCCCGTGGTGTTCCTCTACTTCTGCATTCAAGATTCCCCAAA
 Tb927 GTTGCATGCAAGGAGGGTCCCGTGGTGTTCCTCTACTTCTGCATTCAAGATTCCCCAAA

HK1 TATGCCCTTGAGGGTAACGCCACCGATCTGTTGGCTTCATTGCATCCAATGTGAAAAA
Teru CAGGCACTGCAAGGAACGGCAGAGGATTTGTTGGCTTTATTGCGCAGAGTGTGAAGAAA
Tbg TATGCCCTTGAGGGTAACGCCACCGATCTGTTGACTTCATTGCATCCAATGTGAAGAAA
Tb927 TATGCCCTTGAGGGTAACGCCACCGATCTGTTGACTTCATTGCATCCAATGTGAAGAAA
* * * * *

HK1 CCCCTGGAACCTCTCGTACCATGGAAAACCTCGTGACCTGAGGACCTCAATCGCACAGTTCC
Teru -----ATGATGGAGCAGAAAAGCACCGGAGGACTTGAACCGCACCGTGCC
Tbg -----ACCATGGAACCTCGTGACCTGAGGACCTCAATCGCACAGTTCC
Tb927 -----ACCATGGAACCTCGTGACCTGAGGACCTCAATCGCACAGTTCC
* * * * *

HK1 TCTTGGGTTTACCTTCAGTTTCCCCGTGGAGCAGACGAAGGTTAACCGTGGTGTGCTTAT
Teru GCTTGGCTTACATTTAGCTTTCCAACAGAGCAGAAAAGGCGTCGATCACGGTTTCTGTAT
Tbg TCTTGGGTTTACCTTCAGTTTCCCCGTGGAGCAGACGAAGGTTAACCGTGGTGTGCTTAT
Tb927 TCTTGGGTTTACCTTCAGTTTCCCCGTGGAGCAGACGAAGGTTAACCGTGGTGTGCTTAT
* * * * *

HK1 CCGGTGGACGAAGGGCTTCAGC-----GGATGCTATGTGATTGCCCTTCT
Teru AAAATGGACGAAGGGCTTCTCGACACGGGGTGTGGAGGGGAAGGACGTGGTGGAACTTCT
Tbg CCGGTGGACGAAGGGCTTCAGCAGAAAAGGCGTTCAAGGAAACGATGTGATTGCCCTTCT
Tb927 CCGGTGGACGAAGGGCTTCAGCAGAAAAGGCGTTCAAGGAAATGATGTGATTGCCCTTCT
* * * * *

HK1 TCGCGCTGCTTTGGGAAATTTAGCCTAAGTGTCAATGTTGTGGCTTTGTGCAACGACCC
Teru GCAGAAGGCATTGAAACGCATGGAAGTGAAGGTGAAAGTGGTGGCACTCTGCAACGACAC
Tbg TCAGGCTGCTTTGGGCGAGTGAAGTGAAGTGAAGTGGCGTTGTGCAACGACAC
Tb927 TCAGGCTGCTTTGGGCGAGTGAAGTGAAGTGAAGTGGCGTTGTGCAACGACAC
* * * * *

HK1 CTTTGAACCTTAATTTTCGCATTACTTTAAGGACCCTGAGGTACAGGTTGGTGTGATTAT
Teru TGTCGGTACGCTTATCAGCACTACTTTTACCCCGACACGACAGGTTGGCGTAATCAT
Tbg CGTTGGAACATTAATTTTCGCATTACTTTAAGGACCCTGAGGTACAGGTTGGCGTGATTAT
Tb927 TGTTGCCACGATGATTTTCGCATTACTTTAAGGACCCTGAGGTACAGGTTGGTGTGATTAT
* * * * *

HK1 CGGCACCTGGTCCAGATGCGTGCTACTTTGACAGGGCCTCTGCTGTGACAAAAGACCGTGC
Teru CGGCACCGGCTCCAACGCGTGCTACTTTCGAGGATGCCTACGCCGTGACGAAGGAGCCCTC
Tbg CGGCACCTGGTCCAAATGCGTGCTACTTTGAGACGGCGTCTGCTGTGACGAAGGACCTGC
Tb927 CGGCACCTGGTCCAAATGCGTGCTACTTTGAGACGGCGTCTGCTGTGACGAAGGACCTGC
* * * * *

HK1 CTTTGTGCTCGTGAGTCAGCCTTACTCCCATCAATATACAAAGCCTCCATTTGACTC
Teru AGTGGCCGCGCGCGGTACGACACAGACCAATCAACATGGAGTGGGGAACCTTTGACTC
Tbg CGTTGTGCTCGTGGGTCAGCACTTACTCCCATCAATATGAAAGCGGCAACTTTGACTC
Tb927 CGTTGTGCTCGTGGGTCAGCACTTACTCCCATCAATATGAAAGCGGCAACTTTGACTC
* * * * *

HK1 CGAGTACCGGTATGTCCT—CTACAACAAATTTCAACTTGTATATCGACGATCCGTCGTT
Teru CAAGTACAAGTTTGTGCTGCCGTCACCGCGTACGACGAGGCGATGGATGCCGTTACGCC
Tbg CAAGTACCGGTTTGTCTCCCTACGACGAAGTTCGACTTGGATATTGACGATGCGTCGTT
Tb927 CAAGTACCGGTTTGTCTCCCTACGACGAAGTTCGACTTGGATATTGACGATGCGTCGTT
* * * * *

HK1 AAACAAATGTCAACATGCTCTAGAGAAGAT-ATATCCCGCATGTATCGCGGTAAAATCTC
Teru GAACCGCAACTTCCAGACGCAAGAGAAGATGGTCTCCGGCATGTATCTGGGGAAAATCAG
Tbg GAACAAGGGTCAACAGGCGCTGGAGAAGATGATATCCGGCATGTACCTCGGTGAAAATCGC
Tb927 GAACAAGGGTCAACAGGCGCTGGAGAAGATGATATCCGGCATGTACCTCGGCGAAAATCGC
* * * * *

HK1 CCGCCGCGTTATGTGCACCTTTCGTCTATAAGCCGCTTC-----CTGCGGCATCCA
Teru TCGCCGATGATAGCGCACCTTGGCGAGTTGCATTGCCTCCCAAGTGTCTTGCATCCAA
Tbg CCGCCGCGTTATGTGCACCTTTCGTCTATAAAGTGCCTTC-----CTGCGGCATCCA
Tb927 CCGCCGCGTTATGTGCACCTTTCGTCTATAAAGTGCCTTC-----CTGCGGCATCCA
* * * * *

HK1 GACGGCTTTTGGACAACCGGGTGTGCTTTTAAATCCCATTTCCCGGAATCACCAGTGT
Teru GATGGC-----CAAGCCGTGGAGCTTTGAGACGAAATTT-ATGGGCATGATATCCGCA
Tbg GACTGC-TTTGGGCAACCGGGGTCGTTTGTGAGTCCCGATTT-GCCGGGATGATCAGTGT
Tb927 GACTGC-TTTGGGCAACCGGGGTCGTTTGTGAGTCCCGATTT-GCCGGGATGATCAGTGT
* * * * *

HK1 TCACCTAAGCCCCGACTTCAGTTCACTCGCAGCAGATCCAGAAGGTGTGTGGTGTGA
Teru G-ACCGCATGCCGGGTCTGCAGTTACGCGGCAAGTGTTCGAAGAGCTTTTCCAAGTCCA
Tbg G-ACCGTATGCCGGGACTTCAGTTCACTCGCAGCAGATCCAGAAGGTGTGTGGTGTGA
Tb927 G-ACCGTATGCCGGGACTTCAGTTCACTCGCAGCAGATCCAGAAGGTGTGTGGTGTGA
* * * * *

HK1 CGTGCAGTCAATTGAAGACCTTCGCATCATTCCGATGTGTGCCGCTTGTCCGTGGGAG
Teru CGTGACGGATGTTGCAGACCTGCACGTGATCCGTGATGTGTGCTGCCTGGTGCAGCGGCCG
Tbg CGTGCAGTCAATTGAAGACCTTCGCATCATTCCGATGTGTGCCGCTTGTCCGTGGGAG
Tb927 CGTGCAGTCAATTGAAGACCTTCGCATCATTCCGATGTGTGCCGCTTGTCCGTGGGAG
* * * * *

HK1 GGCTGCGCAACTCTCTGCTTCTTCTGCTGCGCTCCACTGGTTAAGACTCAAACACAGGG
Teru CGCTGCGCAGATCAGTGCCATGTTCTGCACTGCGCCACTTGTGAAGACAAGGAAAGAGGG
Tbg GGCTGCGCAACTCTCTGCTTCTTCTGCTGCGCTCCACTGGTTAAGACTCAAACACAGGG
Tb927 GGCTGCGCAACTCTCTGCTTCTTCTGCTGCGCTCCACTGGTTAAGACTCAAACACAGGG
* * * * *

HK1 CCGTGCAACTATTGCAATTGACGGCTCCGTGTTTGAAGATTCCGTCATTCCGCCGCGT
Teru CCGTGCAACTATTGCAATTGACGGCTCCGTGTTTGAAGACACCCCTCATTCCGCCGCGT
Tbg CCGTGCAACTATTGCAATTGACGGCTCCGTGTTTGAAGATTCCGTCATTCCGCCGCGT
Tb927 CCGTGCAACTATTGCAATTGACGGCTCCGTGTTTGAAGATTCCGTCATTCCGCCGCGT

```

*****
HK1 CTTCGAGGACAACATCAACCGTATCCTTGGCCCTGAGTGGCGATGTCAGGGCCGTTCTCGC
Tcru GCTGCAGCAAAACATGAACGCCATTCTCGGTCTGGGTGTGATGTCACGACGGCACTCGC
Tbg CTTGCAGGACAACATCAACCGTATCCTTGGCCCTGAGTGGCGATGTCAGGGCCGTTCTCGC
Tb927 CTTCGAGGACAACATCAACCGTATCCTTGGCCCTGAGTGGCGATGTCAGGGCCGTTCTCGC
*****
HK1 AAAGGATGGCAGTGGAAATTGGTGCTGATTTATTTCCGCAATGGTGGTGAACGACAAGTAA
Tcru AAGGGACGGCAGCGGCATTGGCGCCGCTTTTATCTCCGCGCTGGTGTGAACGACAAATAA
Tbg AAAGGATGGCAGTGGAAATTGGTGCTGATTTATTTCCGCAATGGTGGTGAACGACAAGTAA
Tb927 AAAGGGTGGCAGTGGTGTGCGTGGCAGCTTATCTCCGCTATCGTTGCTACGGGAAGTGA
*****

```

Figure1 Alignment of the gene sequences of *Trypanosoma brucei* hexokinase with other hexokinase sequences. Tcru: *Trypanosoma cruzi* (AJ437577.1); Tbg: *Trypanosoma brucei gambiense* (FN554973.1); *Trypanosoma brucei brucei* (TREU927) (XM841212.1). Asterisks indicate where all the organisms share identical or highly conserved sequence.

Table 1 Summary of the Comparison of hexokinase 1 Sequence with some trypanosomes Sequences in the NCBI Data Base (DB) using NCBI BLASTN

Species	Query Cover	Identity	NCBI Accession Number
<i>T. brucei Gambiense</i>	100%	87%	FN554973.1
<i>T. brucei brucei</i> TREU927	100%	87%	XM841212.1
<i>Trypanosoma cruzi</i>	38%	74%	AJ437577.1

The NCBI TBLASTP analysis gave 464 translated amino acids, which is close to 471 amino acids of *T. brucei brucei* hexokinase 1 (XP_822456.1) in the NCBI data bank and 474 amino acids sequence earlier reported (Willson *et al.*, 2002). Further analysis of the sequence and aligned using CLUSTAL W software, the result of comparison is shown in Table 2 and Figure 2. It revealed that translated amino acids from the hexokinase 1 gene is 68% similar to that of *T. brucei gambiense* and *T. brucei brucei* TREU927. On the other hand, there is only 48% and 26% similarity between it and that of *Trypanosoma cruzi* and Human Glucokinase (Hexokinase IV) (Figure 2 and Table 2).

```

      10  20  30  40  50  60  70  80  90  100
...|...|...|...|...|...|...|...|...|...|...|
HK1  MSRRLNNLEHISIQGNDGETVRAVKRDVAMAALTNQFTMSVESMRQIMTYLLYEMVEGLEGR---
ESTVRMLPSYVYKADPKRATGVFYALDLGGTNER
Gluk  -----
MLDDRARMEAAKKEKVEQILAEFQLQEEEDLKKVMRRMQKEMDRGLRLETHEEASVKMLPTYVVRSTPEGSEVGDFLSLDLGGTNER
Tcru  MSARLNNLLQHIAVKDKDSDTMRHLKQRMALASLANQFTVGGDKHLKQLMLYMHQMIIEGLEGR---
ESTLRMLPSYVYKIDPSKATGVFYALDLGGTNER
Tbg   MSRRLNNLEHISIQGNDGETVRAVKRDVAMAALTNQFTMSVESMRQIMTYLLYEMVEGLEGR---
ESTVRMLPSYVYKADPKRATGVFYALDLGGTNER
TbU927 MSRRLNNLEHISIQGNDGETVRAVKRDVAMAALTNQFTMSVESMRQIMTYLLYEMVEGLEGR---
ESTVRMLPSYVYKADPKRATGVFYALDLGGTNER

      110 120 130 140 150 160 170 180 190 200
...|...|...|...|...|...|...|...|...|...|...|
HK1  VLRVACKEGA----
VVDSSSTSAFKIPKYALEGNATDLFGFIASNVKRTMETRAPEDLNRTVPLGFTFSFPVEQTKVNRGVLIRWTKGFSTKGVQGNVDIA
Gluk  VMLVKVGEEGEQWSVKTKHQMYSIPEDAMTGAEMLFDYISECISDFLDKHKQMK--
HKKLLPLGFTFSFPVRHEDIDKILLNWTGKFKASGAEGNNVVG
Tcru  VLRVTCKEGR----
VADRVDKAFVIPQALQGTAEEDLFGFIAQSVKMMEQKAPEDLNRTVPLGFTFSFPTEQKGVVDHGFLLIKWTKGFSTRGVEGKDVVE
Tbg   VLRVACKEGA----
VVDSSSTSAFKIPKYALEGNATDLFGFIASNVKKTMETRAPEDLNRTVPLGFTFSFPVEQTKVNRGVLIRWTKGFSTKGVQGNVDIA
TbU927 VLRVACKEGA----
VVDSSSTSAFKIPKYALEGNATDLFGFIASNVKKTMETRAPEDLNRTVPLGFTFSFPVEQTKVNRGVLIRWTKGFSTKGVQGNVDIA

      210 220 230 240 250 260 270 280 290 300
...|...|...|...|...|...|...|...|...|...|...|
HK1  LLQAAFQ-
RVSLKVNVALCNDTVGTLISHYFKDPEVQVGVHIGTGPDAICYFDRASAVTKDRAFAARESALTPINIQSLHFDFSEYRYVL---KFQL-VYR
Gluk  LLRDAIKRRGDFEMDVAMVNDTVATMISCYEDHQCEVGMIVGTGCNACYMEEMQNVELVEGDEGR----
MCVNTEWGAFGDSGELDEFLELYDRLVDE
Tcru  LLQKALK-
RMEVKVVALCNDTVGTLITNYFFDPDTQVGVHIGTGSNACYFEDAYAVTKEPSVAARGTTQTPINMECGNFDSKYKFLVPTAYDEAMD
A
Tbg   LLQAAFQ-
RVSLKVNVALCNDTVGTLISHYFKDPEVQVGVHIGTGSNACYFETASAVTKDPAVAARGSAALTPINMESGNFDSKYRFLVPTTKFDLDIDD
TbU927 LLQAAFQ-
RVSLKVNVALCNDTVGTLISHYFKDPEVQVGVHIGTGSNACYFETASAVTKDPAVAARGSAALTPINMESGNFDSKYRFLVPTTKFDLDIDD

      310 320 330 340 350 360 370 380 390 400
...|...|...|...|...|...|...|...|...|...|...|
HK1  RSVVKGQQALEKMGISMYLGEIARRVIVHLSSINCLPA-ALQALGNRGSFESRFAGMIK---
PRLQFTRSTIQKVCVGVVQSIEDLRIIRDVCRLVRG
Gluk  SSANPGQQLYEKLLIGGKYMGEVRLVLLRLVDENLLFHGEASEQLRTRGAFETRFVSQVESDTGDRKQIYN--ILSTLGLRPS-
TTDCDIVRACESVST
Tcru  VTPNRFQTQEKMVSGMYLGEISRRMIAHLAELHCLPS-
ALASKMAKPWSFETKFMGMISADRMPLQFTRQVFQELFQVDVTDVADLHVIRDVCCLVRG
Tbg   ASLNKGQQALEKMGISMYLGEIARRVIVHLSSINCLPA-
ALQALGNRGSFESRFAGMISADRMPLQFTRSTIQKVCVGVVQSIEDLRIIRDVCRLVRG
TbU927 ASLNKGQQALEKMGISMYLGEIARRVIVHLSSINCLPA-
ALQALGNRGSFESRFAGMISADRMPLQFTRSTIQKVCVGVVQSIEDLRIIRDVCRLVRG

```

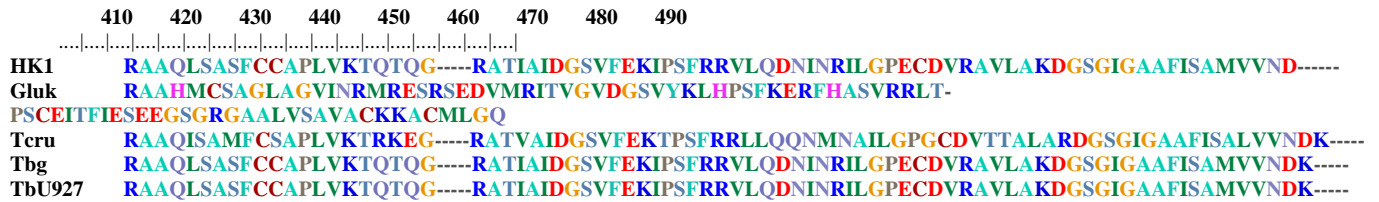



Figure 2 Comparison of the Amino Acid Sequences of the *Trypanosoma brucei brucei* hexokinase (*Federe isolate*) with the hexokinases of *T. brucei gambiense* (XP_01177423.1) - Tbg; *T. brucei brucei* TREU927 (XP_822456.1) - TbU927; *T. cruzi* (AAL93565.1) -Tcr; Human Glucokinase (NP_000153.1) - Gluk

Table 2 Summary of the Comparison of hexokinase 1 Gene Product (Protein) and hexokinase of some common trypanosomes and Human Glucokinase NCBI BLASTP

Organisms	Query Cover	Identity	NCBI Accession Number
<i>T. brucei Gambiense</i>	99%	68%	XP_01177423.1
<i>T. brucei brucei</i> TREU927	99%	68%	XP_822456.1
<i>Trypanosoma cruzi</i>	99%	48%	AL93565.1
Human Glucokinase	91%	26%	NP_000153.1

The NCBI Conserved Domains analysis of *T. b. brucei* hexokinase 1 protein aligned to members of hexokinase 2 super family that have Accession Number c127242 as shown in Figure 3. The actual alignment results was detected with superfamily member (Accession Number [PTZ00107](#)). The conserved domains for glucose were represented by the amino acids in blue colour, ATP in purple and glucose 6-phosphate in red respectively. The plus sign indicate the binding sites of the sugar moiety of G6P. The letter “a” indicates the binding sites of the phosphate moiety of G6P. The amino acids residues marked “C” are present in the conserved hydrophobic channel. The putative, linear B-cell epitopes with higher conservations are marked with green over-line. The putative connecting the two domains are respectively marked with blue over-line

L.b	MASRVNNLMNHLAIRSDSEEMRYIKQRLA-LASLSTQFTMASEKMKQLTMYMVYEMVEG	60
L.i	MATRVNNLLSHIAIRSDSEEMRYIKQRLA-LASLATQFTMSSEKMKQLTMYMIHEMVEG	60
L.m	MAARVNNLLSHIAIRSDSEEMRYIKQRLA-LASLATQFTMSSEKMKQLTMYMIHEMVEG	60
HK1	MSRRLNILEHISIQGNDGETVRSFY--VT-LHALTNQPFSLVESMRQIMTYLLYEMVEG	60
T.bb	MSRRLNILEHISIQGNDGETVRAVKRDVA-MAALTNQFTMSVESMRQIMTYLLYEMVEG	60
C.p	-----MEEENQAKRFLFDLYEYFFLSNLKLELVDLDFHKSLEDG	60
C.m	-----MEVRKYRSSDEWIELYRPFYFCLSKAKLQDLVEDFLNSLICG	60
c+c a		
L.b	LEGRP-----STVRMLPSYVYTS DPAKATGVYYALDLGGTNFRVLRVSLRSGK	120
L.i	LEGRP-----STVRMLPSFVYTS DPAKATGVYYALDLGGTNFRVLRVSLRGGK	120
L.m	LEGRP-----STVRMLPSFVYTS DPAKATGVYYALDLGGTNFRVLRVSLRGGK	120
HK1	LEGLH-----TTL SILPCTRRG--PYALAFFSPFDLCSIFCVLRGHARRVP	120
T.bb	LEGRE-----STVRMLPSYVYKADPKRATGVFYALDLGGTNFRVLRVACKEGA	120
C.p	LENHANRLKIDKYHSNYKPFKMLDSCVDR LPTGKEKGVYYAIDMGGTNLRCVVRNLLGNG	120
C.m	LESHNGITSAN--ICIQKPLKVL DSCIFNLPSGKEQGIHYAIDMGGTNLRCVVRVELRGNG	120
c+c a		
L.b	VDDRIDSKFVIPKSAL TGN-----SANLDFDIA	180
L.i	VDDRTDSKFVIPKSALVGD-----ATDLDFDIA	180
L.m	VDDRTDSKFVIPKSALVGD-----ATDLDFDIA	180
HK1	WCFPLLLHKS SPNMLRVT-----PPICLASLHPMKN	180
T.bb	VVDSSTSAFKIPKYALEGN-----ATDLFGFIA	180
C.p	QSETKFKKVKLSEMKVSTGKVVNSNKISK-----VDQEVNIFDKTVSSETMFNSIA	180
C.m	ESFVTTYKMMALNDLRISKHKAIENNINIETGDNLFFNSDIQSFSILDKLASATDMFDAIS	180
c c		
L.b	QSVKKMM-----SE-----NAPE	240
L.i	QSVKKMM-----SE-----NAPD	240
L.m	QSVKKMM-----SE-----NAPD	240
HK1	PWNSRTM-----ET-----RAPE	240
T.bb	SNVKKTM-----ET-----RAPE	240
C.p	VFFNEFLDECGLDNLDTNDN-----N	240
C.m	NFFYDFL VSCGDINERILEFRDNNEAYTFNSKIITKPLSTNLNKNENQLQKYHEKDLK	240
c c		
L.b	DLEKRVPLGFTFSFPVDQKAVNKGLLIKWTKGFST----KNVEGNDVV ELLQGS LRRMH	300
L.i	DLEKRVPLGFTFSFPVDQKAVNKGLLIKWTKGFST----KNVEGNDVV ELLQAS LRRVR	300
L.m	DLEKRVPLGFTFSFPVDQKAVNKGLLIKWTKGFST----KNVEGNDVV ELLQAS LRRVR	300
HK1	DLNRTVPLGFTFSFPVEQTKVNRGVLRWTKGFS-----GCYVIALLR AAFGKFS	300
T.bb	DLNRTVPLGFTFSFPVEQTKVNRGVLRWTKGFS-----KGVQGN DVIALLQAAFGRVS	300
C.p	VGNLPLEV GFTFSFPVQSKIASAKLVIWTKEIETGRLTDDPVEGKDIGDLLN LAFKRNG	300
C.m	YQSNYFSV AFTFSFPPTQLSIANAHLISWTKGIETGRATLEPVEGYDVG NLLNSAFNRNK	300
C c c c c		
L.b	INVNVVALCNDTVGTLVARYFV-----DTNAQVGV IIGTGSNA	360
L.i	VNVNVVALCNDTVGTLVARYFV-----DTDVQVGV IIGTGSNA	360
L.m	VNVNVVALCNDTVGTLVARYFV-----DTDVQVGV IIGTGSNA	360
HK1	LSNVVALCNDPF-TLISHYFK-----DPEVQVGV IIGTGPDA	360
T.bb	LKVNVALCNDTVGTLISHYFK-----DPEVQVGV IIGTGSNA	360
C.p	VPAQCKCVLNDTVGTLISAMYDLNINNYCDNNHSLKVNFPNNISENQPLIGI VVGTVGNA	360
C.m	IPAHCCKIINDTVGTLISAMYDLNSNMVGNMSSYSNNSTPRQLTNSNPVIGI VIGTGINA	360
+ a		

L.b	CYFERASAVTKDPAVCARGNAVTPINMECGNFDSKYKYALPTTVYDDEMDAITPNRDHQR	420
L.i	CYFERASAVTKDPAVSARGNAVTPINMECGNFDSKYKYALPITVYDDEMDAITPNRENQR	420
L.m	CYFERASAVTKDPAVSARGNAVTPINMECGNFDSKYKYALPITVYDDEMDAITPNRENQR	420
HK1	CYFDRASAVTKDRAFAARESALTPINIQLSHFDSEYRYVLYN-KFQLVYRRSVVKQMSTC	420
T.bb	CYFETASAVTKDPAVAARGSAALTPINMESGNFDSKYRFVLPPTTKFDLIDDDASLNKGQQA	420
C.p	CYLEPNSSNF-----GYKGVIIINTECGDFYS---TKLPITDCDYSDWFSNDRGEQI	420
C.m	CYIEPMSLFY-----GYKGVIIINTECGDFTC---SNLPITDCDLVLDWFSNDRGDQQ	420
L.b	QEKLVSVMYLGEISRRMIVHLAQLGCLPRDLVDGLGKWPWAFESKHMGM-VAADQMP----	480
L.i	QEKLVSVMYLGEISRRLLIVHLAQLGCLPRGLVDGLCRPWAFESKHMGM-IAADQMP----	480
L.m	QEKLVSVMYLGEISRRLLIVHLAQLGCLPRGLVDGLCRPWAFESKHMGM-IAADQMP----	480
HK1	SREDISRMRYRGKISRRLVIVHLSSISRLPAALQTAFGQPGVVLIPISPESVVLHLKP----	480
T.bb	LEKMISGMYLGEIARRVIVHLSSINCLPAALQTAALGNRGSFESRFAGM-ISADRMP----	480
C.p	FEKMISGTYLGEISRLLIINFLKKNKTPEIFFQK-----NSLKTEHIAKIIISHFNNDNHQKY	480
C.m	FEKMISGTYLGEISRLLIINFLKKNKTPEIFFQK-----KIITTEDIANIASFDEDN----	480
L.b	GLQFTRELIRKRVAGVN---VTDVADLHTIREICLVRNRAAQQAAVLSAAPMLK-TRTQG	540
L.i	GLQFTRELIRKRIAGVD---VTDMSDLHTIREACCLVRNRAAQQGAVFTAAPMLK-TRTQG	540
L.m	GLQFTRELIRKRIAGVD---MADISDLHTIRETCCLVRNRAAQQGAVFTAAPMLK-TRTQG	540
HK1	RLQFTRSTIQKVCQVD---VQSIEDLRIIRDVCRVLRGAAQLSASFCCAPLVK-TQTQG	540
T.bb	GLQFTRSTIQKVCQVD---VQSIEDLRIIRDVCRVLRGAAQLSASFCCAPLVK-TQTQG	540
C.p	NQNHDLKSNIENYLKETFSSNLDHNSTYIIAKISQMVLMRAASLVSAIIAFAFFKRFNKPRN	540
C.m	-TSINIDKIQRFIKSKYINVILDTSSCNTIKTMAIFVLKRAAGLVATVIAALIKKIENFQN	540
L.b	LATVAVDGVSVEKVPSPFQRLYQECITGILGPTSN---AKVVVLQKDGSGVGAAMICALAAN	600
L.i	LATVAVDGVSVEKTPSFQRLYQECITSILGTSN---VKVVVLQRDGSVGAAMICALAAN	600
L.m	LATVAVDGVSVEKTPSFQRLYQECITSILGTSN---VKVVVLQRDGSVGAAMICALAAN	600
HK1	RATIAIDGVSVEKIPSFRRVLDNINRILGPECD---VRAVLAKDGSVGAAFISAMVVN	600
T.bb	RATIAIDGVSVEKIPSFRRVLDNINRILGPECD---VRAVLAKDGSVGAAFISAMVVN	600
C.p	QITIAIDGVSVEKIPKFKQYKVDLSLSSLIQESGYLGSIHFYESDDGSGRGAAILASTTVN	600
C.m	GITVAVDGVSVMTRVVKFQNYTKENLNILGE-QVSQFIQFYEADDGSGKGAAILAATMD-	600
	+ a	a
L.b	QK	602
L.i	KK	602
L.m	TK	602
HK1	DK	602
T.bb	DK	602
C.p	TH	602
C.m	--	602

Figure 3 Amino acids in the conserved domains of *T.b.brucei* hexokinase 1(Hk1) as aligned to some members of hexokinase 2 super family (Accession Number [c127242](#)). *Leishmania braziliensis*(L.b, XP_001564741.1);*Leishmania infantum*(L.i,XP_001465335.1);*Leishmania major* (L.m,XP_001682957.1);*Trypanosoma brucei brucei* (T.bb,XP_822456.1);*Cryptosporidium parvum* (C.p,XP_627719.1); *Cryptosporidium muris* (C.m,XP_002140303.1)

DISCUSSION

The open reading frame (ORF) of *T. b. brucei* hexokinase 1 (*TbbHK-1*) consisted of 1401 bp encoding 464 translated amino acids. This gene size was larger than the 1350 base pairs obtained for hexokinase from *Clonorchis sinensis* (Chen et al., 2014), but closer to that of *T. cruzi* (1416 bp) (Cáceres et al., 2003). BLASTx showed that the deduced amino acid sequence of *TbbHK 1*, shared 68%, 68%, 48% and 26% identity with *T. brucei gambiense* (XP 01177423.1), *T.bruceibrucei* (TREU927: XP822456.1), *T. cruzi* (AAL93565.1) and Human Glucokinase(NP 000153.1) respectively. These 464 translated amino acid sequence was slightly shorter than the number of the translated amino acids of *T. brucei brucei* (TREU927: XP822456.1) which is the reference standard for *T. brucei brucei* hexokinase in the NCBI data bank (471 amino acids) and (474 amino acids) obtained by Willson et al.,(2002).

A sequence alignment of the amino acids sequence also demonstrated that *T. brucei brucei* hexokinase 1 shared common evolutionary relationships (Bork et al., 1993). As observed in hexokinase from other organisms, the polypeptide also contained the amino acid sequence for ATP-binding site ⁸⁸ALDLGGTNR⁹⁶, ²³²VGVIIGTGSNAC²⁴³, and ⁴¹⁷AIDGSV⁴²², with all the conserved amino acid preserved as described by Cáceres et al. (2003) and Bork et al. (1992) except one in position 92 where Glycine to Cysteine (transition or substitution) occurs. The amino acids sequence that constitute hexokinase prosite signature¹⁵⁶LGFTFSFPTEQKG VDHGFLIKWTKGF¹⁸¹ were also conserved (Cáceres et al., 2003; Schirch & Wilson, 1987). There were multiple binding sites for glucose, sugar and phosphate constituents of Glucose 6- phosphate in the amino acid sequence of *T. brucei brucei* hexokinase 1 (Mulichak et al.,1998). The translated amino acids from the hexokinase 1 gene carry some unique features which include peroxisomal targeting signals of type 2 (PTS-2) sequences (⁴RLxLxH¹²) in the N-terminal which is important for the translocation of the enzyme to glycosomes (Blattner et al., 1995). All the hexokinase sequences aligned contain the PST-2 sequences except Human Glucokinase (Hexokinase

IV). This shows that all the enzymes are glycosome resident enzymes except Human Glucokinase which lacks the PST sequence. In addition to the PST-2 sequence, the result also showed that *T. brucei brucei* (Federe strain) hexokinase 1 also contained “NTD25” sequence which is the next 25 amino acids after the peroxisome targeting sequence-2 sequence in *T. brucei brucei* hexokinase (Flynn et al., 1998). This sequence has been shown to direct *T. brucei brucei* hexokinase to the mitochondria/basal bodies in Procyclic forms and the flagellum in Blood Stream Forms where it serves alternative or additional functions in this extra-glycosomal locations before the NTD25 is finally removed (Harris, 2015; Joice et al., 2013). Harris (2015) in his work showed that misprocessing of the PTS2 of *T. brucei brucei* hexokinase leading to its premature removal can divert this glycosome bound enzyme to an alternative destination, that is, removal of PST-2 can mis-localize the enzyme which is harmful to the parasites. He also observed that mutation of serine 13 which is the first amino acid in NTD 25 to alanine had no effect on either processing or cellular localisation but mutation of conserved arginine 23 to have a deadly effect on cell morphology in both BSF and PF cells. In BSF parasites, it resulted in the failure of the cell to properly divide while PF cells had a flagella defect, with multiple flagella emanating from each cell and concluded that apart from its function *T. brucei brucei* hexokinase has other critical role in the parasite apart from glycolysis.

The NCBI Conserved Domains analysis of *T.b. brucei* Hexokinase 1 protein aligned to members of Hexokinase 2 super family that have Accession Number [c127242](#) as shown in Figure 3 showed that the *TbbHK1* protein sequence depicted that it has a close relationship with the members of the family and the amino acids peculiar to this family well preserved (Marchler-Bauer et al., 2017). The two structurally similar domains in this family are also preserved (Marchler-Bauer et al., 2015).

CONCLUSION

From this study it can be seen clearly that *T. brucei brucei* hexokinase 1 is uniquely different from its mammalian forms but conserves the amino acids in domains and motifs peculiar to the Hexokinase 2 super family (Accession Number c127242 and EC:2.7.1.1). The unique difference between *T. brucei brucei* (Federe isolate) hexokinase 1 and its mammalian forms revealed potential for its selective inhibition without interference with the host hexokinase. This may be a possible avenue for immunisation or drug development against this disease.

ETHICAL APPROVAL

Animal experiments were carried out in accordance with the instructions for the care and use provided by the University of Jos, Nigeria.

REFERENCE

- Bakker, B.M., Mensonides, F.I., Teusink, B., van Hoek, P., Michels, P.A., & Westerhoff, H.V. (2000). Compartmentalisation protects trypanosomes from the dangerous design of glycolysis. *Proceedings of National Academic of Science*, 97(5), 2087–2092. <https://doi.org/10.1073/pnas.030539197>
- Besteiro, S., Biran, M., Biteau, N., Coustou, V., Baltz, T., Canioni, P., & Bringaud, F. (2002). Succinate secreted by *Trypanosoma brucei* is produced by a novel and unique glycosomal enzyme, NADH-fumarate reductase. *The Journal of Biological Chemistry*, 277 (41), 38001-12. <http://www.jbc.org/lookup/doi/10.1074/jbc.M201759200>
- Blattner, J., Dörsam, H., & Clayton, C.E. (1995). Functions of N-terminal import signals in Trypanosome microbodies. *Federation of European Biochemical Societies Letter*, 360, 310–314. [https://doi.org/10.1016/0014-5793\(95\)00128-V](https://doi.org/10.1016/0014-5793(95)00128-V)
- Bork, P., Sander, C., & Valencia, A. (1992). An ATPase domain common to prokaryotic cell cycle proteins, sugar kinases, actin, and hsp70 heat shock proteins. *Proceedings of National Academic of Science*, 89, 7290–4. <https://doi.org/10.1073/pnas.89.16.7290>
- Bouteille, B., & Buguet, A. (2012). The detection and treatment of human African trypanosomiasis. *Research and Reports in Tropical Medicine*, 3, 35-45. <https://doi.org/10.2147/RRTM.S24751>
- Bringaud, F., Riviere, L., & Coustou, V. (2006). Energy Metabolism of Trypanosomatids: Adaptation to available carbon sources. *Molecular and Biochemical Parasitology*, 149, 1-9. <https://doi.org/10.1016/j.molbiopara.2006.03.017>
- Cáceres, A. J., Ramon, P., Acosta, H., David, R., Wilfredo, Q., Luisana, ... Concepción, L. (2003). Molecular and biochemical characterization of hexokinase from *Trypanosoma cruzi*. *Molecular & Biochemical Parasitology*, 126, 251–262. [https://doi.org/10.1016/S0166-6851\(02\)00294-3](https://doi.org/10.1016/S0166-6851(02)00294-3)
- Chen, T., Ning, D. Sun, H., Li, R., Shang, M., Li, X., & Yu, X. (2014). Sequence Analysis and Molecular Characterization of Clonorchissinensis Hexokinase, an Unusual Trimeric 50-kDa Glucose-6-Phosphate-Sensitive Allosteric Enzyme. *PLoS ONE*, 9 (9), e107940. <https://doi.org/10.1371/journal.pone.0107940>
- Franco, J.R., Simarro, P.P., Diarra, A., & Jannin, J.G. (2014). Epidemiology of human African trypanosomiasis. *Clinical Epidemiology*, 6, 257–275. <https://doi.org/10.2147/CEP.S39728>
- Flynn, C.R., Mullen, R.T., & Trelease, R.N. (1998). Mutational analyses of a type 2 peroxisomal targeting signal that is capable of directing oligomeric protein import into tobacco BY-2 glyoxysomes. *Plant Journal*, 16, 709-720. <https://doi.org/10.1046/j.1365-313x.1998.00344.x>
- Gasteiger, E., Hoogland, C., Gattiker, A., Duvaud, S., Wilkins, M.R., Appel, R.D., & Bairoch, A. (2000). *Protein Identification and Analysis Tools on the ExPASy Server*; (In) John M. Walker (ed): *The Proteomics Protocols Handbook*, Humana Press
- Harris, M. T. (2015). "A Survey of Protozoan Parasite Hexokinases: Characterization studies and potential for therapeutic interventions". *All Dissertations*. Paper 1496. https://tigerprints.clemson.edu/all_dissertations/1496?utm_source=tigerprints.clemson.edu%2Fall_dissertations%2F1496&utm_medium=PDF&utm_campaign=PDFCoverPages
- Haanstra, J.R., van Tuijl, A., Kessler, P., Reijnders, W., Michels, P. A., Westerhoff, H. V., & Bakker, B. M. (2008). Compartmentation prevents a lethal turbo-explosion of glycolysis in trypanosomes. *Proceedings of National Academic of Science*, 105, 17718-17723. <https://dx.doi.org/10.1073/pnas.0806664105>
- Herbert, W.J., & Lumsden, W.H. (1976). *Trypanosoma brucei*: A rapid matching method for estimating the host's parasitaemia. *Experimental Parasitology*, 40, 427-31. [https://doi.org/10.1016/0014-4894\(76\)90110-7](https://doi.org/10.1016/0014-4894(76)90110-7)
- Joice, A. C., Harris, M.T., Kahney, E. W., Dodson, H.C., Maselli A.G., Whitehead, D.C. & Morris, J.C. (2013). Exploring the mode of action of ebsele in *Trypanosoma brucei* hexokinase inhibition. *International Journal for Parasitology, Drugs and Drug Resistance*, 3, 154-160. <https://doi.org/10.1016/j.ijpddr.2013.08.002>

- Lanham, S.M., & Godfrey, D.G. (1970). Isolation of salivarian trypanosomes from man and other mammals using DEAE-cellulose. *Experimental Parasitology*, 521-534. [https://doi.org/10.1016/0014-4894\(70\)90120-7](https://doi.org/10.1016/0014-4894(70)90120-7)
- Matthews, K.R., McCulloch, R., & Morrison, L.J. (2015). The within-host dynamics of African trypanosome infections. *Philosophical Transactions of The Royal Society B Biological Sciences*, 370, 20140288. <https://doi.org/10.1098/rstb.2014.0288>
- Marchler-Bauer, A., Bo Y, Han, L., He, J., Lanczycki C.J., Lu, S., ... Bryant, S.H. (2017). "CDD/SPARCLE: functional classification of proteins via subfamily domain architectures.", *Nucleic Acids Res.* 45(D)200-3. <https://dx.doi.org/10.1093/nar/45D200-3>
- Marchler-Bauer, A., Derbyshire, M.K., Gonzales, N.R., Lu, S., Chitsaz, F., Geer, L.Y., ... Bryant, S.H. (2015). "CDD: NCBI's conserved domain database.", *Nucleic Acids Res.* 43(D)222-6. <https://doi.org/10.1093/nar/gku1221>
- Mulichak, A.M, Wilson, J.E., Padmanabhan, K., & Garavito, R.M. (1998). The structure of mammalian hexokinase-1. *Nature Structural Biology*, 5, 555–560. <https://doi.org/10.1038/811>
- Schirch, D.M. & Wilson, J.E. (1987). Rat brain hexokinase: location of the substrate hexose binding site in a structural domain at C-terminus of the enzyme. *Archives of Biochemistry and Biophysics*, 254, 385–96. [https://doi.org/10.1016/0003-9861\(87\)90116-0](https://doi.org/10.1016/0003-9861(87)90116-0)
- Shaw, A. P., Cecchi, G., Wint, G. R., Mattioli, R. C., & Robinson, T. P. (2014). Mapping the economic benefits to livestock keepers from intervening against bovine trypanosomiasis in Eastern Africa. *Preventive Veterinary Medicine*, 113, 197–210. <http://www.fao.org/3/dx.doi.org/10.1016/j.prevetmed.2013.10.024>
- Simarro, P. P., Cecchi, G., Franco, J. R., Paone, M., Diarra, A., Ruiz-Postigo, J. A., ... Jannin, J. G. (2012). Estimating and mapping the population at risk of sleeping sickness. *PLoS Negl Trop Dis*, 6, e1859. <http://dx.doi.org/10.1371/journal.pntd.0001859>
- Simarro, P. P., Cecchi, G., Paone, M., Franco, J. R., Diarra, A., Ruiz, J. A., ... Jannin, J. G. (2010). The Atlas of human African trypanosomiasis: a contribution to global mapping of neglected tropical diseases. *International Journal of Health Geography*, 9, 57. <https://doi.org/10.1186/1476-072X-9-57>
- Steverding, D. (2010). The development of drugs for treatment of sleeping sickness: a historical review. *Parasite and Vectors*, 3, 15. <https://doi.org/10.1186/1756-3305-3-15>
- Tesfaye, D., Speybroeck, N., De Deken, R. & Thys, E. (2012). Economic burden of bovine trypanosomiasis in three villages of Metekel zone, northwest Ethiopia. *Trop Animal Health Prod.*, 44(4), 873-879. <https://doi.org/10.1007/s11250-011-9981-3>
- Thompson, J.D., Higgins, D.G., & Gibson, T.J. (1994). CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position specific gap penalties and weight matrix choice. *Nucleic Acids Research*, 22:4673–80. <https://doi.org/10.1093/nar/22.22.4673>
- Ureta, T. (1982). The comparative isozymology of vertebrate hexokinases. *Comparative Biochemistry and Physiology*, 71B, 549–555. <https://doi.org/10.1016/j.ijcard.2017.04.04>
- Willson, M., Sanejouand, Y., Perie, J., Ve'ronique, H., & Opperdoes, F. (2002). Sequencing, Modeling, and Selective Inhibition of *Trypanosoma brucei* Hexokinase. *Chemistry & Biology*, 9, 839–847. [https://doi.org/10.1016/S1074-5521\(02\)00169-2](https://doi.org/10.1016/S1074-5521(02)00169-2)

SUPPLEMENTARY MATERIAL

Phosphate Buffered Saline (PBS)

To 800 ml of distilled water add
 488.8mg of NaH₂PO₄
 488.8mg of NaH₂PO₄
 2.55g of NaCl
 8.08g of Na₂HPO₄
 15g of D-glucose

The volume was then adjusted to 1000ml with distilled H₂O and the pH to 7.8 with 5N NaOH. It was sterilized by autoclaving and the pH rechecked before use. Each syringe used for blood collection contained 100µl of heparin mixed with 500µl of PSG