

ASSESSMENT OF THE HYGIENIC CONDITION OF THE SLAUGHTERHOUSE BASED ON THE EVALUATION OF MICROBIOLOGICAL SWABS

Mária Vargová¹, Katarína Veszelits Laktičová¹, František Zigo^{*2}, Ibrahim F. Rehan^{3,4}

Address(es):

Slovakia.

¹ Department of Public Veterinary Medicine and Animal Welfare, University of Veterinary Medicine and Pharmacy, Košice, Komenského 73, 041 81, Slovakia.

² Department of Animal Nutrition and Husbandry, University of Veterinary Medicine and Pharmacy, Košice, Komenského 73, 041 81,

³Menoufia University, Faculty of Veterinary Medicine, Department of Husbandry and Development of Animal Wealth, Shebin Alkom, Menoufia, 32511, Egypt. ⁴Meijo University, Faculty of Pharmacy, Department of Pathobiochemistry, Yagotoyama 150, Tempaku-Ku, Nagoya-Shi, Aichi, 468-8503, Japan.

*Corresponding author: frantisek.zigo@uvlf.sk

https://doi.org/10.55251/jmbfs.9469

ARTICLE INFO ABSTRACT This study objective was to evaluate the hygienic condition of surfaces by microbiological swabs and the effectiveness of disinfectant Received 19. 9. 2022 Virkon S. Level of hygiene was evaluated in a small-capacity slaughterhouse located in the Košice region with a maximum weekly Revised 13. 10. 2023 capacity of 5 large livestock units. Microbiological swabs were taken from an area of 10 cm² before the process of slaughtering, during Accepted 17. 10. 2023 the process, and after disinfection. Disinfectant Virkon S was used in a 1% concentration during an exposure time of 30 minutes for Published 1. 2. 2024 disinfection of monitored surfaces. Disinfectant was effective on surfaces of cage, wall, floor, and (p<0.0001), where was determined significant decrease of microorganisms, but we recorded 1.0 x 10⁴ colony forming units (CFU)/10 cm² of the total count of bacteria (TCB) on the cage. 7.5 x 10³ CFU/10 cm² of TCB on the wall and 1.5 x 10⁴ CFU/10 cm² of TCB on the floor after disinfection, which indicates Regular article an insufficient level of disinfection. From the achieved results, it is clear, that the disinfection provided by the disinfectant Virkon S was not effective, as the evaluated disinfectant did not achieve a decrease in the number of microorganisms and thus did not ensure a sufficient level of hygiene. We concluded that it is crucial to effectively disinfect products using the right disinfectant at the right exposure period since it helps to stop the spread of several germs that could contaminate products and have a negative impact on consumers' health.

Keywords: disinfection, hygiene, microbiological swabs, slaughterhouse

INTRODUCTION

Slaughterhouse is a key source of bacterial contamination in meat and its products, which is a major health and economic concern for several public authorities (Klaham et al., 2022). The key hygiene controls at each stage are highlighted in the general principles of food hygiene, which represent hygiene practices from basic production through to final consumption. Food contamination can happen in a variety of ways and at any point throughout production, distribution, and storage (McBain et al., 2000). Slaughtering procedures in slaughterhouses play a significant influence in the transmission of foodborne microorganisms (Shang et al., 2019; Rasschaert et al., 2008). Slaughterhouse is a premise approved and registered by the controlling authority for hygienic slaughtering and inspection of animals, processing, effective preservation, and storage of meat products for human consumption (Alonge, 1991). Their general hygiene requirements are set out in Regulation of the European Parliament and the Council (EC) No. 853/2004 and Commission Regulation (EC) No. 2023/2006. A slaughterhouse is an establishment used for the slaughter and slaughtering of animals whose meat is intended for human consumption, must be equipped with equipment for disinfecting instruments with hot water (minimum temperature is 82 °C) (Lagin & Lopašovský, 2004).

The slaughtering of animals should take place under veterinary supervision and complete and appropriate hygienic precautions since this is the most crucial aspect in ensuring the production of meat products of high quality (Serda *et al.*, 2015; Zailani *et al.*, 2016).Bacterial contamination can occur on the surface of the meat during meat preparation, carcass cutting, manufacturing of meat products, packing, during transportation and storage until it reaches the consumer (Heyndrickx *et al.*, 2002; Ananchaipattana, 2003). This contamination can also occur during primary production at the farm from the first skin incision made to remove the blood, especially if the tools used by the operator are not sterile. In most slaughterhouses, carcasses are not skinned, but they are subjected to several steps that result in the skin being visibly clean and free from hair. Despite this, the carcasses may be heavily contaminated with microorganisms. The carcasses may be further contaminated during the next steps of evisceration and cutting (Gill *et al.*, 2000). Bacterial hazards represent a major concern in the production of food of animal origin. The major contamination points during slaughter are animal-related, such

as faecal and pharyngeal, and environmental. Hazard Analysis Critical Control Point (HACCP) and Good Manufacturing Practice (GMP) must be focused on maximal limiting this spread which leads to the prevention of microbial carcass contamination to ensure the health protection and meat safety (Lindblad & Berking, 2013). Because slaughter is an open process, there are several chances for the carcass to become contaminated with potentially harmful microorganisms. The procedure has some steps where the amount of bacteria may be decreased, but it doesn't include any steps where risks are completely removed. Only partial control can be obtained for the critical control points (CCPs) indicated for slaughtering techniques, and there is limited scope for risk prevention (Bakri et al., 2017). Some CCPs stand for control points (CPs) that GMP controls (Fig. 1). The general mechanisms of spread of the harmful bacteria differ. The main source of contamination for Yersinia enterocolitica, Campylobacter spp., and Salmonella spp. is the contamination of carcasses, which can be restricted if only stringent slaughtering protocols are followed. In the processing environment, other organisms such Aeromonas spp. and Staphylococcus aureus may be endemic, but they may be managed with proper cleaning and disinfection (Brown et al., 2000; Zweifel et al., 2008). These microorganisms serve as indicators for the GMP regulations.

Flawless hygienic production conditions are ensured by sanitation. The basic elements of a comprehensive sanitation activity are cleaning and disinfection. Thorough cleaning must be performed before applying disinfection. This step is essential for the disinfection to be as effective as it can be to remove organic and other undesirable matter, like biofilm, from the surfaces that may protect the microorganisms from the effects of the disinfectant (Dvorak, 2005). The goal of disinfection is to destroy microorganisms; this does not automatically mean killing all microorganisms but reducing the number to a level that is not normally harmful to health (Skaarup, 2011). To achieve effective disinfection the surfaces subject to disinfection must be thoroughly wet, and the disinfectant must be applied in the correct concentration. It must also be allowed to stay on the surfaces for the appropriate contact time (Dvorak, 2005). Inappropriate use but also overuse of biocides in different areas such as food industries, hospitals, and homes may lead to the emergence of resistance to various biocides (e.g.: quaternary ammonium compounds, triclosan, chlorhexidine or trisodium phosphate) (Brauodaki & Hilton, 2005; Romanova et al., 2002; Yuk & Marshall, 2006). The selective

pressure exerted by biocides is responsible for cross-resistance between antibiotics and biocides (**Davin-Regli & Pagès, 2012**) since the use of low concentrations of biocides may increase the risk of selection of resistant microorganisms.

Table 1 Hygienic a	aspects and preventive actions with re	espect to bacterial hazards at the	he slaughterhouse	
Process step	Hygienic aspect	Preventive actions	CP/CCP	
Lairage ↓ Stunning	Contamination between animals	Cleaning & disinfection	СР	
Killing	Contamination from tools	Cleaning & disinfection	СР	
Scalding ↓	Reduction of bacterial levels Contamination of lungs	Time/Temperature	СР	
Dehairing	Contamination from machines	Cleaning & disinfection	СР	
Flaming	Reduction of bacterial levels	Time/Temperature	СР	
Polishing	Contamination from machines	Cleaning & disinfection	СР	
Evisceration ↓	Contamination from intestines Contamination from the tongue, pharynx and tonsils Contamination from tools	Enclosure of rectum Working instructions Disinfection of tools	ССР	
Splitting ↓	Contamination via splitter/saw	Line-speed Water temperature	СР	
Meat inspection \downarrow	Contamination from inspection	Disinfection of tools	ССР	
Deboning of head	Contamination from head	Working instructions Disinfection of tools	ССР	

Legend: CP - control point, CCP – critical control points Source: Borch *et al.* (1996)

Evaluation of the disinfection process is one of the stages of disinfection. It should be done both during and after the process. Microbiological control is the most impartial way of disinfection evaluation. This technique shows whether the disinfection was effective. The total bacterial count or the presence of indicator bacteria are employed as a substitute for the arduous and unreliable process of detecting pathogenic germs (**Tuladhar** *et al.*, **2012**). Microbiological swabs are taken after the correct exposure time of the disinfection. A sufficient number of swabs according to the size of the disinfected area should be taken from different equipment and surfaces (**Griffith**, **2016**).

The objective of the study was to evaluate the hygienic condition of monitored surfaces by microbiological swabs and the effectiveness of disinfectant Virkon S used in a 1% concentration during an exposure time of 30 minutes.

MATERIALS AND METHODS

Characteristics of slaughterhouse and disinfectant

The study was performed in a small-capacity slaughterhouse with a maximum weekly capacity of 5 large livestock units. Slaughterhouse was located in the Košice region. The slaughter was divided into 3 parts - part I (slaughter and bleeding); part II (steambath and removal of bristles) and part III (evisceration) (Figure 1). In terms of slaughterhouse structure, the slaughterhouse in our experiment was a closed-system building and it had separated slaughtering lines for clean and unclean areas. Workers were assigned to a single location and did not rotate across other regions of the slaughterhouses. The equipment is rigorously confined to the authorized regions and is not combined within the designated zones. Workers donned safety gear, including boots and jackets, and cleansed their hands before going through the slaughtering line. The temperature of the scalding water was regulated, and the slaughtering and cutting knives were sterilized. To avoid cross-contamination between carcasses and the slaughtering floor, the slaughterhouse utilized hanging apparatus. For the purposes of the butchering, they used tap water. Every carcass treated in slaughterhouses had its internal organs removed.

For disinfection of different surfaces in evaluated slaughterhouses detergents and disinfectants were used. The detergent used for mechanical cleaning of surfaces was 2% Fint used as a degreaser by heating 50 - 60 °C with an exposure time. Disinfectants used for disinfection of surfaces were Fink – FC 21 and Virkon S which was evaluated. Fink FC 21 is a disinfectant used in liquid form, which is a high-foaming and cleaning agent, alkali, containing active chlorine. The disinfectant was applied by spraying at 2% concentration while being heated to 50 °C for 20 minutes of exposure. Virkon S was applied as a liquid in a 1% concentration by spraying, without being heated. The exposure lasted for thirty minutes. Oxone (potassium peroxymonosulfate, used as an oxidizing agent),

sodium dodecylbenzene sulfonate (anionic surfactant), sulfamic acid, and inorganic buffers are all components of this multipurpose disinfectant. It is a balanced, stabilized blend of peroxygen compounds, surfactant, organic acids, and inorganic buffer. This disinfectant is recommended for use as a hard surface disinfectant in livestock production and transportation facilities. Fink Kanalreiniger was used for disinfectants. It was used 200 grams in 0.5 liters of water with an exposure time of 30 minutes. Detergents and disinfectants used in the slaughterhouse were rinsed with water at the end of the processes of mechanical cleaning and disinfection.



Figure 1 From left: Division of slaughterhouse - part II (steambath and removal of bristles) and part III (evisceration)

Microbiological swabs

Assessment of the hygienic condition of the slaughterhouse was carried out by microbiological swabs. Swabs were taken from monitored places, from part I (slaughter and bleeding) before and during slaughtering and bleeding and after disinfection. Microbiological swabs were collected from 10 cm2 areas of the cage, wall, floor, and lift that were under evaluation (Figure 2). Six swabs were obtained from each location, totaling 24 samples for analysis. A sterile tube containing 10 ml of sterile saline solution and swabs was used. 0.1 ml of this mixture was put to the various agar plates. After being incubated in a thermostat, plates were used to analyze the colonies that had grown. Endo agar was used for coliform bacteria (CB), meat peptone agar was used for the total count of bacteria (TCB), and Sabouraud agar was made from molds. After 24 hours at 37 °C, the findings from Meat Peptone Agar and Endo Agar were obtained. After 3 to 5 days of incubation at room temperature, the findings from Sabouraud agar were obtained. For the determination of coliform bacteria, the total count of bacteria, and molds, the procedures according to the applicable ISO standards were used (ISO 18593; ISO

21257; ISO 4832). Numbers of microorganisms were expressed in CFU (colony forming units).



Figure 2 From left: Evaluated places for microbiological evaluation - cage, wall, floor, lift, and way of application of disinfectant

Statistical analysis

The results were statistically processed using descriptive statistical analysis of data and statistical method of the Student's t-test **for paired comparisons**. The differences in the numbers of the total count of bacteria, coliform bacteria, and molds were calculated between conditions before the process of slaughtering and after disinfection. The continuous variables were represented using mean (M) standard deviation (SD), whilst the categorical variables were described over the average of 5 samples from chosen surfaces (CFU/10 cm2). 0.05 was the threshold for significance (p).

RESULTS AND DISCUSSION

The effectiveness of disinfectants for microorganisms depends on many factors. On the one hand, these are the properties of the microorganisms themselves, on the other hand, the chemical and physical properties of the external environment (Vargová *et al.*, 2022). The concentration of the disinfectant, exposure time, pH, temperature, the presence of organic contaminants, such as blood, serum, or other body fluids, the microorganism or agent itself, their type (prions, viruses, gramnegative, gram-positive bacteria, microscopic fungi, protozoa, or spores), as well as their number and location, are factors that affect disinfection efficiency (Štefkovičová, 2007). Failure of the disinfection may be because of an ineffective disinfectant, or because of the effect of an environmental factor (Simões *et al.*, 2010). The insufficient exposure time was the reason for the failure of disinfection in our study.

The detection of pathogenic microorganisms in the outdoor environment is difficult and not sufficiently reliable, therefore, as part of the microbiological control of the effectiveness of disinfection, we determine the total count of bacteria or the presence of indicator bacteria – coliform bacteria (Vargová *et al.*, 2022). *Escherichia coli* is an example of a culturable coliform bacterium that can be utilized as a microbial surrogate for surface quality monitoring since it can be used to detect the presence of fecal material from warm-blooded animals. These bacterial species are part of the normal microflora that live in warm-blooded animals' intestines, and their presence on surfaces indicates the presence of bacterial pathogens. There are four different indicators of fecal contamination: total coliform, fecal coliform, *E. coli*, and *enterococcus* (Byappanahalli *et al.*, 2012). In Tables 2, 3, 4, and 5 is shown the effect of disinfectant Virkon S used at 1% concentration during exposure time 30 minutes on the evaluated surfaces - cage, wall, floor, and lift before the process of slaughtering, during the process of slaughtering and after disinfection.

Table 2 Effect of disinfectant Virkon S on monitored microorganisms present in the cage before the process of slaughtering, during the process, and after disinfection

		Cage		
		$(CFU/10 \text{ cm}^2)$		Р
	before	during	after	
ТСВ	1.5 x 10 ⁶	2.4 x 10 ⁴	$1.0 \ge 10^4$	p<0.0001
p-value			before vs after	p<0.0001
СВ	2.5 x 10 ⁴	3.8 x 10 ³	1.8 x 10 ²	p<0.0001
p-value			before vs after	p<0.0001
Molds	1.5×10^2	1.9 x 10 ⁴	8.4 x 10 ¹	p<0.0001
n-value			before vs after	$\mathbf{p} = \mathbf{n}\mathbf{s}$

Legend: CFU - colony forming units; TCB - total count of bacteria; CB - coliform bacteria. A level of 0.05 was considered significant (p), ns – not significant

Finding the total count of bacteria up to 10^3 on the floor and up to 10^2 on the other monitored surfaces is permissible and disinfection is considered effective. In Table 2, the number of TCB after disinfection was 1.0×10^4 CFU/10 cm² of TCB, which indicates insufficient disinfection (Sasáková *et al.*, 2020). According to

Ondrašovičová *et al.* (2013), the effectiveness of preventive disinfection is satisfactory if the number of indicator bacteria is within 10% of the original number. The numbers of CB on the cage after disinfection did not exceed 10% of their original number, which is considered a good result of disinfection.

 Table 3 Effect of disinfectant Virkon S on monitored microorganisms present on the wall before the process of slaughtering, during the process, and after disinfection

		Wall		
		(CFU/10 cm ²))	Р
	before	during	after	
ТСВ	3.5 x 10 ⁴	1.4 x 10 ⁴	7.5 x 10 ³	p<0.0001
p-value			before vs after p	< 0.0001
СВ	>1.0 x 10 ¹	1.1 x 10 ²	>1.0 x 10 ¹	p<0.0001
p-value		before vs after $p = ns$		
Molds	$1.0 \ge 10^3$	2.3 x 10 ³	1.1 x 10 ¹	p<0.0001
p-value			before vs after p	>0.0001

Legend: CFU - colony forming units; TCB - total count of bacteria; CB - coliform bacteria. A level of 0.05 was considered significant p)

 Table 4 Effect of disinfectant Virkon S on monitored microorganisms present on the floor before the process of slaughtering, during the process, and after disinfection

		Floor		
		(CFU/10 cm ²)	Р
	before	during	after	
TCB	1.5 x 10 ⁶	2.7 x 10 ⁴	1.5 x 10 ⁴	p<0.0001
p-value			before vs after p-	< 0.0001
СВ	1.1 x 10 ⁴	1.5 x 10 ⁴	1.4 x 10 ²	p<0.001
p-value		before vs after p<0.0001		
Molds	>1.0 x 10 ¹	2.2×10^3	>1.0 x 10 ¹	p<0.0001
p-value			before vs after p =	= ns

Legend: CFU - colony forming units; TCB - total count of bacteria; CB - coliform bacteria. A level of 0.05 was considered significant (p).

The bacterial contamination originates from the external animal surface, from the internal animal environment - from the gastrointestinal tract, as well as from the environment, including air, soil, water, equipment surfaces, and also humans. Control of the presence of pathogenic microorganisms on surfaces is based on the approaches of minimizing surface contamination through proper sanitation which includes mechanical cleaning and disinfection and the application of decontaminating procedures (Gebel *et al.*, 2013).

In Table 3 is shown the effect of disinfectant on evaluated microorganisms present on the wall. Virkon S was not effective enough on the monitored surface, because after disinfection we recorded 7.5 x 10³ CFU/10 cm² of TCB, which indicates an insufficient level of disinfection. In Table 4 is shown the effect of disinfectant on the presence of TCB, CB, and molds situated on the floor. In a comparison of several monitored microorganisms before the process of slaughtering and after disinfection, was recorded with a significant decrease in the number of TCB, CB, and molds (p<0.0001). Disinfectant Virkon S was effective against CB and molds, except TCB, where after disinfection we recorded 1.5 x 10⁴ CFU/10 cm².

 Table 5 Effect of disinfectant Virkon S on monitored microorganisms present on the lift before the process of slaughtering, during the process, and after disinfection

		Lift		
		(CFU/10 cm ²)		Р
	before	during	after	
ТСВ	1.1 x 10 ⁴	1.3 x 10 ²	1.0 x 10 ²	p<0.0001
p-value			before vs after p	< 0.0001
СВ	5.7 x 10 ²	8.4 x 10 ²	>1.0 x 10 ¹	
				p<0.0001
p-value			before vs after p-	< 0.0001
Molds	1.3 x 10 ²	8.5 x 10 ³	>1.0 x 10 ¹	p< 0.0001
p-value			before vs after p	< 0.0001

Legend: CFU - colony forming units; TCB - total count of bacteria; CB - coliform bacteria. A level of 0.05 was considered significant p).

In Table 5 is shown the effect of disinfectant on TCB, CB, and molds present on the lift. Virkon S was effective on the monitored surface where the number of evaluated microorganisms was permissible after disinfection and a significant decrease in microorganisms were obtained (p<0.0001).

In our experiment, disinfectant Virkon S caused a significant decrease of evaluated microorganisms but wasn't effective enough on each of the evaluated surfaces which leads to the conclusion that this disinfectant used at 1% of concentration during exposure time 30 minutes is not suitable for disinfection of surfaces in a slaughterhouse. However, according to a study by **Vargová** *et al.* (2021) Virkon S

used in a 1% concentration during an exposure time of 60 minutes was suitable for disinfection on the premises of slaughterhouses.

The finding of some of the monitored microorganisms on evaluated surfaces after disinfection is related to insufficient exposure time (30 minutes) and in some cases - Table 2: 1.0×10^4 CFU/10 cm² of TCB; Table 3: 7.5×10^3 CFU/10 cm² of TCB and Table 4: 1.5×10^4 CFU/10 cm² of TCB exceed the limit for preventive disinfection. Therefore, ensuring an appropriate contact time = exposure time can influence whether a pathogen is inactivated, killed, or unaffected. Contact times are usually dependent on the material of the surface and the concentration used.

CONCLUSION

We confirmed that the disinfectant Virkon S used in 1% concentration during an exposure time of 30 minutes was not effective enough against the total count of bacteria, coliform bacteria due to the insufficient exposure time. Our findings regarding the microbiological contamination of surfaces in the slaughterhouse may help hygienists in comparable settings establish appropriate hygienic practices for the prevention or reduction of microbiological contamination of surfaces.

Acknowledgement: This work was supported by Slovak grant VEGA no. 1-0162-23.

REFERENCES

Alonge, D. O. (2005). *Textbook of Meat Hygiene in the Tropics*. Ibadan, Nigeria: FarmCoe Press.

Ananchaipattana, C., Hosotani, Y., Kawasaki, S., Pongsawat, S., Latiful, B. Md., Isobe, S., Inatsu, Y. (2012). Prevalence of foodborne pathogens in retailed foods in Thailand. *Foodborne Pathogens and disease*. *9*(9), 835–40. https://doi.org/10.1089/fpd.2012.1169

Bakri, M., Maarof, A. G., Norazmir, M. N. (2017). Confusion determination of critical control point (CCP) via HACCP decision trees. *International Food Research Journal* 24(2): 747-754. https://www.researchgate.net/publication/317933369_Confusion_determination_

of critical_control_point_CCP_via_HACCP_decision_trees

Brauodaki, M., & Hilton, A.C. (2005). Mechanisms of resistance in *Salmonella enterica* adapted to erythromycin, benzalkonium chloride and triclosan.

International journal of antimicrobial agents, 25, 31-37.

Byappanahalli, M. N., Nevers, M. B., Korajkic, A., Staley, Z. R., Harwood, V. J. (2012). Enterococci in the Environment. *Microbiology and Molecular Biology Reviews*, 76(4), 685–706.

Borch, E., Nesbakken, T., Christensen, H. (1996). Hazard identification in swine slaughter respect to foodborne bacteria. *International Journal of Food Microbiology*, *30*, 9-25.

Commission Regulation No. 2023/2006 of the European Parliament and of the Council of 22 December 2006 on good manufacturing practice for materials and articles intended to come into contact with food. OJ L 314M, 723–726. <u>https://eur-lex.europa.eu/LexUriServ/LexUriServ.do?uri=OJ:L:2006:384:0075:0078:EN:PD</u> F

Commission Regulation No. 853/2004 of the European Parliament and the Council of 29 April 2004 establishing special hygiene regulations for food of animal origin. http://faolex.fao.org/docs/pdf/eur63427.pdf

Davin-Regli, A. D., & Pagès, J. M. (2012). Cross-resistance between biocides and antimicrobials: an emerging question. *Revue scientifique et technique* (*International Office of Epizootics*), *31*(1), 89-104.

https://pubmed.ncbi.nlm.nih.gov/22849270/

Dvorak, G. (2005). Disinfection 101. In: Center for Food Security and Public Health, Iowa State University, Ames, IA, 123-130.

Gebel, J., Exner, M., French, G., Chartier, Y., Christiansen, B., Gemein, S., Goroncy-Bermes, P., Hartemann, P., Heudorf, U., Kramer, A., Maillard, J. Y., Oltmanns, P., Rotter, M., Sonntag, H. G. (2013). The role of surface disinfection in infection prevention. *GMS Hygiene and Infection Control*, 8(1), ISSN 2196-5226. https://pubmed.ncbi.nlm.nih.gov/23967396/

Gill, C., Dussault, F., Holley, R., Houde, A. (2000). Evaluation of the hygienic performances of the processes for cleaning, dressing and cooling pig carcasses at eight packing plants. *International journal of food properties*, 58 (1-2), 65-72. https://pubmed.ncbi.nlm.nih.gov/10898463/

Heyndrickx, M., Vandekerchove, D., Herman, L., Rollier, I., Grijspeerdt, K., De Zutter, L. (2002). Routes for Salmonella contamination of poultry meat: epidemiological study from hatchery to slaughterhouse. *Epidemiology & Infection*, *129*(2), 253–65. <u>https://pubmed.ncbi.nlm.nih.gov/12403101/</u>

ISO (the International Organization for Standardization) 18593, 2004. Microbiology of food and animal feeding stuffs: Horizontal methods for sampling techniques from surfaces using contact plates. https://www.iso.org/standard/39849.html

ISO (the International Organization for Standardization) 21527, 2008. Microbiology of food and animal feeding stuffs: Horizontal method for the enumeration of yeasts and molds. <u>https://www.iso.org/standard/38275.html</u>

ISO (the International Organization for Standardization) 4832, 2006. Microbiology of food and animal feeding stuffs: Horizontal method or the enumeration of coliforms. <u>https://www.iso.org/standard/38282.html</u>

Klaharn, K., Pasquali, F., Pichpol, D., Meeyam, T., Harintharanon, T., Lohaanuku, P., Punyapornwithaya, V. (2022). Bacterial contamination of chicken meat in slaughterhouses and the associated risk factors: A nationwide study in Thailand. *Plos one*, <u>https://doi.org/10.1371/journal.pone.0269416</u>

Lagin, L., Lopašovský, Ľ. (2004). Technology of meat I. (Slaughter). Nitra: SPU, 100-104, ISBN 80-8069-425-7.

Lindblad, M, Berking, C. (2013). A meat control system achieving significant reduction of visible faecal and ingesta contamination of cattle, lamb and swine carcasses at Swedish slaughterhouses. *Food Control*, *30*, 101-105. https://doi.org/10.1016/j.foodcont.2012.07.040

Brown, M. H., Gill, C. O. Hollingsworth, J. Nickelson, R., Seward, S., Sheridan J. J., Stevenson, T., Sumner, J. L., Theno, D. M., Usborne, W. R., Zink, D. (2000). The role of microbiological testing in systems for assuring the safety of beef. *International Journal of Food Microbiology*, *62*, 7-16. https://pubmed.ncbi.nlm.nih.gov/11139024/

McBain, A., Allison, D. G., Gilbert, P. (2000). Population dynamics in bacterial biofilms. In: Community and Co-operation in Biofilms. Society for General Microbiology, Reading, 257-278.

Ondrašovičová, O., et al. (2013). Hygiena chovu zvierat/Animal hygiene (in Slovak). Košice: University of Veterinary Medicine and Pharmacy, Košice.

Rasschaert, G., Houf, K., Godard, C., Wildemauwe, C., Pastuszczak-Frak, M., De Zutter, L. (2008). Contamination of carcasses with Salmonella during poultry slaughter. *Journal of Food Protection*, 71(1), 146–52. https://pubmed.ncbi.nlm.nih.gov/18236675/

Romanova, N., Favrin, S., Griffiths, M. W. (2002). Sensitivity of *Listeria* monocytogenes to sanitizers used in the meat processing industry. Applied and Environmental Microbiology, 68, 6405-6409. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC134375/

Sasáková, N., Gregová, G., Veszelits Laktičová, K., Vargová, M., Hromada, R., Szabóová, T., Kachnič, J. (2020). *Veterinary care for the environment*. University textbook for VVL and HP - first edition. Košice: University of Veterinary Medicine and Pharmacy in Košice.

Serda, B., Ayalew, H., Berhanu, A., Sibhat, B. (2015). Microbiological assessment of meat contact surfaces at the abattoir and retail houses in Jigjiga town, Somali National Regional State of Ethiopia. *Journal of Food and Agricultural Science*, *5*(3), 21–26.

https://academicjournals.org/article/article1426595815_Ayalew%20%20et%20al %20.pdf

Shang, K., Wei, B., Jang, H. K., Kang, M. (2019). Phenotypic characteristics and genotypic correlation of antimicrobial resistant (AMR) Salmonella isolates from a poultry slaughterhouse and its downstream retail markets. *Food Control*, *100*, 35–45. <u>https://agris.fao.org/agris-search/search.do?recordID=US201900161561</u>

Simo^{es}, M., Simo^{es}, L., Vieira, M. J. (2010). A review of current and emergent biofilm control strategies. *LWT - Food Science and Technology*, *43*, 573–583.

Skaarup, T. (2011). *Slaughterhouse cleaning and sanitation*. Food and Agriculture Organization of the United Nations, Rome. https://canadianpreppersnetwork.com/cd3wd/disk5/ ag_slaughterhouse_cleaning_sanitation_53_unfao_en_lp_116710_.pdf

Štefkovičová, M. (2007). Disinfection and sterilization, theory and practice II., Žilina, 88-90.

Tuladhar, E., Hazeleger, W. C., Koopmans, M., Zwietering, M. H. (2012). Residual viral and bacterial contamination of surfaces after cleaning and disinfection. *Applied and environmental microbiology*, 78(21), 7769-7775. https://pubmed.ncbi.nlm.nih.gov/22941071/

Griffith, C. (2016). *Surface Sampling and the Detection of Contamination*. Handbook of Hygiene Control in the Food Industry. Chapter 44. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7152397/pdf/main.pdf

Vargová, M., Sasáková, N., Veszelits Laktičová, K., Zigo, F. (2021). Evaluation of the hygienic condition of the slaughterhouse. *Acta fytotechnica and zootechnica, 24*, 37-40.

Vargová, M., Veszelit Laktičová K., Sasáková, N., Gregová, G., Hromada, R., Szaboóvá, T. (2022). Hygiena zdravotníckych a farmaceutických zariadení. Vysokoškolská učebnica na praktické cvičenie pre študijný program Farmácia, 193 s. ISBN 978 – 80 – 8077 – 754 – 8.

Yuk, H. G., & Marshall, D. L. (2006). Effect of trisodium phosphate adaptation on changes in membrane lipid composition, verotoxin secretion, and acid resistance of *Escherichia coli* O157:H7 in simulated gastric fluid. *International Journal of Food Microbiology*, *106*(1), 39-44. https://pubmed.ncbi.nlm.nih.gov/16213051/

Zailani, A., Bello, M., Raji, A., Kabir, J., Yahuza, M. (2016). Microbial evaluation of meat contact surfaces in red meat abattoirs of Bauchi State, North-Eastern Nigeria. *Open Journal of Medical Microbiology*, *6*(1), 3–8. https://www.scirp.org/journal/paperinformation.aspx?paperid=64328

Zweifel, C., Fischer, R., Stephan, R. (2008). Microbiological contamination of pig and cattle carcasses in different small-scale Swiss abattoirs. *Meat Science*, 78(3), 225-31. <u>https://pubmed.ncbi.nlm.nih.gov/22062274/</u>